TFOS DEWS II Diagnostic Methodology report

James S. Wolffsohn, FCOptom PhD Chair [a, *], Reiko Arita, MD PhD [b], Robin Chalmers, OD [c],
Ali Djalilian, MD [d], Murat Dogru, MD PhD [e], Kathy Dumbleton, MCOptom PhD [f],
Preeya K. Gupta, MD [g], Paul Karpecki, OD [h], Sihem Lazreg, MD [i],
Heiko Pult, MSc (Optom) PhD [a, j, k], Benjamin D. Sullivan, PhD [l],
Alan Tomlinson, FCOptom PhD [m], Louis Tong, FRCS PhD [n], Edoardo Villani, MD [o],
Kyung Chul Yoon, MD PhD [p], Lyndon Jones, FCOptom PhD [q],
Jennifer Craig, MCOptom PhD [r]

[a] Ophthalmic Research Group, Aston University, Birmingham, UK
[b] Department of Ophthalmology, Itoh Clinic, Saitama, Japan
[c] Clinical Trial Consultant, Atlanta, GA, USA
[d] Illinois Eye and Ear Infirmary, UIC Department of Ophthalmology & Visual Sciences, Chicago, IL, USA
[e] Department of Opthalmology, Keio University School of Medicine, Shinjukuku, Tokyo, Japan
[f] School of Optometry, University of California, Berkeley, CA, USA
[g] Cornea & Refractive Surgery, Duke Eye Center, Durham, NC, USA
[h] Kentucky Eye Institute, KY, USA
[i] Cabinet Ophtalmologie, Alger Centre, Algiers, Algeria
[j] 'Dr Heiko Pult — Optometry and Vision Research', Weinheim, Germany
[k] School of Optometry and Vision Sciences, Cardiff University, Cardiff, UK
[l] Tearlab, San Diego, CA, USA
[m] Glasgow Caledonian University, Glasgow, UK
[n] Corneal and External Eye Disease, Singapore National Eye Center, Singapore
[o] Department of Clinical Sciences and Community Health, University of Milan & Eye Clinic San Giuseppe Hospital, IRCCS Multimedica, Milan, Italy
[p] Department of Ophthalmology, Chonnam National University Hospital, Gwangju, South Korea
[q] Centre for Contact Lens Research, University of Waterloo, Waterloo, Ontario, Canada
[r] New Zealand National Eye Centre, Department of Ophthalmology, The University of Auckland, Auckland, New Zealand

ARTICLE INFO

ABSTRACT

The role of the Tear Film and Ocular Surface Society (TFOS) Dry Eye Workshop (DEWS) II Diagnostic Methodology Subcommittee was 1) to identify tests used to diagnose and monitor dry eye disease (DED), 2) to identify those most appropriate to fulfil the definition of DED and its sub-classifications, 3) to propose the most appropriate order and technique to conduct these tests in a clinical setting, and 4) to provide a differential diagnosis for DED and distinguish conditions where it is a comorbidity. Symptom screening with the DEQ-5 or OSDI confirms that a patient might have DED and triggers the diagnostic tests of (ideally non-invasive) breakup time, osmolarity and ocular surface staining with fluorescein and lissamine green (observing the cornea, conjunctiva and eyelid margin). Prior to diagnosis, it is important to exclude conditions that can mimic DED with the aid of triaging questions. Meibomian gland dysfunction, lipid thickness/dynamics and tear volume assessment and their severity allow sub-classification of DED (predominantly evaporative or aqueous deficient) which informs the management of DED. Videos of these diagnostic and sub-classification techniques are available on the TFOS website. It is envisaged that the identification of the key tests to diagnose and monitor DED and its sub-classifications will inform future epidemiological studies and management clinical trials, improving comparability, and enabling identification of the sub-classification of DED in which different management strategies are most efficacious.

© 2017 Elsevier Inc. All rights reserved.

* Corresponding author.
  E-mail address: j.s.w.wolffsohn@aston.ac.uk (J.S. Wolffsohn).

1. Introduction

The Diagnostic Methodology Subcommittee set out to first identify tests used to diagnose and monitor dry eye disease (DED) from a comprehensive review of the academic literature, with a particular emphasis on changes since the original Tear Film and Ocular Surface Society (TFOS) Dry Eye Workshop (DEWS) [1]. Studies of test efficacy and/or performance are influenced by the fact that subjects have often been selected based on the same tests that are under scrutiny. Similarly, the performance of any "new" test may be compromised when the test is assessed in a population of DED patients who have been diagnosed using non-standardized criteria.

Secondly the committee identified those tests that are most appropriate to fulfil the definition of DED and its sub-classifications and the most appropriate order and technique to conduct these tests in a clinical setting. The committee also identified areas in which new tests are emerging, which may influence the future of DED diagnosis and monitoring. While the original TFOS DEWS recommended categories of tests that were considered appropriate to include in DED screening, diagnosis and monitoring, as well as a series of templates to standardize these tests [2], the variety of tests in some categories precluded easy comparison of epidemiological studies or clinical trials of potential DED management techniques. In addition, the previous definition of DED from the original TFOS DEWS "Dry eye is a multifactorial disease of the tears and ocular surface that results in symptoms of discomfort, visual disturbance, and tear film instability, with potential damage to the ocular surface. It is accompanied by increased osmolarity of the tear film and inflammation of the ocular surface" identified key elements presumed to be required for the diagnosis of dry eye (symptoms of discomfort, visual disturbance, tear film instability, increased osmolarity and inflammation of the ocular surface) which might all be expected to be present (perhaps sub-clinically) [1]. However, all these aspects are rarely inclusion criteria of studies. Also the definition implies that dry eye can occur without ocular surface damage, yet staining is often listed as an inclusion criterion.

The other main aim of the Diagnostic Methodology Subcommittee was to provide a differential diagnosis rationale chart for primary DED. 'Mystery patient' studies have identified that DED is poorly recognized by non-ophthalmic health professionals, who are often consulted on self-management [3]. Hence it is important to provide guidance as to the best questions to ask in order to differentiate primary DED from conditions that can mimic some characteristics of DED or cases when the dry eye is secondary to an underlying condition. Managing the underlying condition may alleviate the dry eye or change its severity and therefore its appropriate management. The chart also identifies when specialist tests and eye observation equipment are needed and, from this, determines when a referral to an appropriately equipped eye care practitioner is necessary.

2. Goals of the Diagnostic Methodology Subcommittee

The goals of the Diagnostic Subcommittee were to determine the most efficacious battery of tests for diagnosing and monitoring DED as per the revised definition, and to propose the most appropriate order and technique to conduct these tests in a clinical setting. Key diagnostic tests were to be differentiated from tests that inform subset aetiologies. Recommended differential diagnostic procedures for excluding other forms of disease that may mimic some of the signs and symptoms of dry eye were also to be articulated. To be widely adopted, a diagnosis must be based on tests available in clinical practice.

3. Definition of dry eye disease (DED)

The definition of dry eye has been amended by the TFOS DEWS II to "Dry eye is a multifactorial disease of the ocular surface characterized by a loss of homeostasis of the tear film, and accompanied by ocular symptoms, in which tear film instability and hyperosmolarity, ocular surface inflammation and damage, and neurosensory abnormalities play etiological roles." [4] Hence any indication that specific signs must be present for a patient to be diagnosed with dry eye has been removed and an emphasis has been placed on the homeostasis of the tear film. Loss of homeostasis implies the body has lost the ability to maintain equilibrium, resulting in a hyperosmolar, unstable tear film with associated sequelae, e.g., increased osmolarity, inflammation, neuropathy and reduced function (compromised lubrication, hydration). Hence diagnosis requires knowledge of what is considered normal, even though this may vary with patient demographics such as sex, age and ethnicity. There are many aspects of the tear film that could be considered abnormal, such as its stability, volume, osmolarity, pH and constituents, many of which are interrelated.

4. Classification of sub-categories of dry eye disease (DED)

The Definition report identifies that sub-categories of DED can be considered from those where the signs are predominantly evaporative (such as from a deficient lipid layer in meibomian gland dysfunction (MGD)) to those where the signs indicate aqueous deficiency (a reduced tear volume) more strongly, and the spectrum in between [4]. The severity of signs together with the evaporative-to-aqueous bias also form part of the sub-classification 'diagnosis' to aid the management of the patient's DED.

5. Diagnostic considerations

5.1. Diagnosis and monitoring

Forming an accurate clinical diagnosis is the mathematical equivalent to the problem of classification, where a multidimensional input vector of observed clinical parameters is mapped onto a discrete set of output classes, using joint probabilities and history to inform a pattern recognition algorithm. Optimal segregation of the variable space is determined by a combination of risk factors and training data. In one dimension, this concept is represented by the familiar overlapping histograms shown in Fig. 1a. True positives (TP) and false positives (FP) are represented by the portion of the affected and unaffected distributions to the right hand side of the cut-off. True negatives (TN) and false negatives (FN) are represented by the portion of the affected and unaffected distributions to the left hand side of the cut-off. In this example, the cut-off is set to achieve a high sensitivity, as defined by the ratio of true positives to the total number of affected subjects in the study. Accordingly, sensitivity $= TP/(TP+FN)$.

In Fig. 1b, the cut-off is set to achieve a higher specificity, as defined by the ratio of true negatives to the total number of unaffected subjects in the study. Accordingly, specificity $= TN/(TN+FP)$.

In any one dimension, sensitivity and specificity are inversely related, meaning that a more sensitive cut-off will cause a higher rate of false positives, and a more specific cut-off will cause a higher rate of false negatives.

5.2. Risk factor considerations — selecting an appropriate cut-off

The level of risk of an incorrect diagnosis generally governs the optimal cut-off for an individual sign or symptom. While there are a variety of valid, statistical risk models to choose an optimal cut-off,

for example, maximizing the ratio of true positives to false positives, receiver operator characteristic apex, etc, [5] clinical risk should supersede purely statistical methods when relying on a small number of signs or symptoms. For example, if a cataract surgeon understands the impact of an unhealthy ocular surface on biometry and visual outcomes [6–9], a more sensitive cut-off is preferable, as there is little to no safety hazard in treating a DED false positive with lubricants or other first line therapy. Conversely, the systemic costs of over-diagnosis must be considered in general practice, suggesting that a cut-off that produces equivalent risk of false positives and false negatives is more generally applicable for an individual marker. Equivalent risk results in a cut-off at the intersection between the affected and unaffected distributions if the measures of signs or symptoms are normally distributed.

5.3. Aspects of test validation

No single "gold standard" sign or symptom that correlates perfectly with the DED state has been established. If one existed, the distributions of this theoretical marker would be very similar to Fig. 1a and b, with a very small overlap in the affected and unaffected curves. Instead, there is a significant overlap between normal and DED distributions of currently available metrics, as all signs and symptoms fluctuate over time and vary significantly within different levels of disease severity [10,11]. Actual histograms are far more similar to Fig. 1c than to the idealized tests of 1a & 1b [12,13].

5.3.1. Sampling & spectrum bias
The lack of a gold standard makes it very difficult to establish true referent histograms when evaluating new diagnostic tests. The traditional approach to DED classification requires DED subjects to satisfy all criteria within a series of *sensitive* thresholds (such as

Ocular Surface Disease Index (OSDI) > 13, Schirmer < 10 mm/5 min, TBUT < 10 s, positive staining) and normal controls to satisfy all criteria within another, non-overlapping set (such as: OSDI < 7, Schirmer > 10 mm/5 min, TBUT > 10 s, negative staining) [14]. While this approach can produce strikingly high sensitivities and specificities of the diagnostic methods under evaluation, as has been done for the recent introductions of both matrix metalloproteinase-9 (MMP-9) (85% sensitivity) [14], and tear osmolarity (87% sensitivity) [15], this approach excludes a large number of DED patients, as signs and symptoms are uncorrelated across the broad population and do not move in synchrony [16–22]. For instance, it is very common to encounter a patient with a high level of symptoms and yet a lack of evidence of staining. Similarly, patients can be asymptomatic but exhibit obstructed meibomian glands, short breakup time and high osmolarity [22]. Excluding these uncategorized individuals prevents randomization across the broad population and describes sampling bias. Gaps in the inclusion criteria lead to spectrum bias, where normal patients are compared to more severe patients, to the exclusion of the mild to moderate subjects that are difficult to categorize [14]. Both sampling and spectrum bias will improve the sensitivity and specificity of a particular study, but will also increase the mean of the affected sample, shift the intersection of the two histograms to produce an unreasonably high cut-off, and result in unexpectedly poor sensitivity in the broad population. This is particularly relevant to regulatory trials, where labelled performance may not be replicated in the field, if tested on populations that are different to those included in the trial. For example, in the 510(k) summary of a new MMP-9 test, one site reported 97% sensitivity in diagnosing mild DED subjects at ≥ 40 ng/mL, while the other three sites reported 66%, 67% and 76% sensitivity (https://www.accessdata.fda.gov/cdrh_docs/pdf13/K132066.pdf). In milder populations that
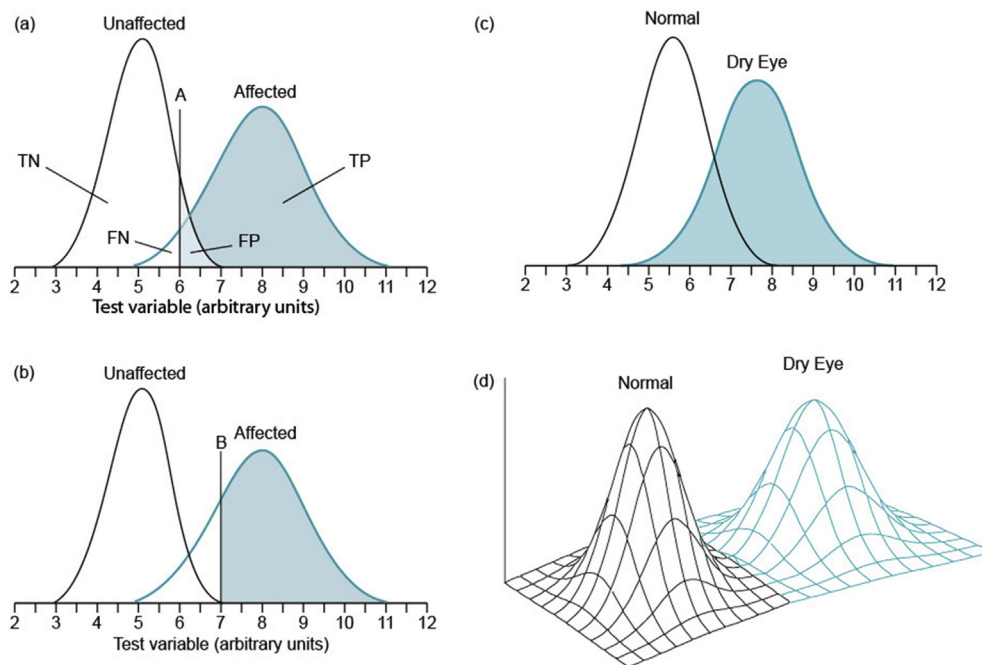

Fig. 1. Segregation of data (such as tear osmolarity or stain grade) for diagnosis concepts: a) representation by overlapping histograms True positives (TP) and false positives (FP) are represented by the portion of the affected and unaffected distributions to the right hand side of the cut-off (A). True negatives (TN) and false negatives (FN) are represented by the portion of the affected and unaffected distributions to the left hand side of the cut-off (A). In this example, the cut-off is set to achieve a high sensitivity, as defined by the ratio of TP to the total number of affected subjects in the study (TP+FN); b) cut-off (B) set to achieve a higher specificity, as defined by the ratio of TN to the total number of unaffected subjects in the study (TN+FP); c) in reality there is significant overlap between normal and DED distributions of currently available metrics, as all signs and symptoms fluctuate over time and vary significantly within different levels of disease severity; d) additional observations (represented by multiple dimensions to the diagnostic vector), increases sensitivity and specificity simultaneously, eventually allowing clear segregation of the affected and unaffected populations at higher orders, even if there is significant overlap in lower dimensions.

did not apply the regulatory trial inclusion criteria, the 40 ng/mL cut-off demonstrated an 11% sensitivity [23]. Similarly, using an equivalent risk threshold of ≥312 mOsm/L applied to a broad population segregated by uncorrelated clinical signs, tear osmolarity reported a sensitivity of 73% [13], and 67% in a milder population outside the trial setting [23].

As a counterpoint, in evaluating new diagnostic metrics, it is not clear whether spectrum bias is undesirable when there is no reliable gold standard to definitively diagnose DED. Without a competent benchmark for delineating affected and unaffected populations, histograms of subject populations will significantly broaden and overlap due to misclassification [11], leading to artificially low sensitivity and specificity of the new diagnostic metric under test. Because symptoms and classical DED signs are so variable over clinically relevant timescales [10,24], inclusion criteria that rely upon these metrics will result in a heterogeneous bias, impossible for even an ideal diagnostic metric to achieve good trial performance. When compared to uncorrelated inclusion criteria across the broad population, novel test sensitivities in the 40–70% range are statistically pre-determined for a single metric, regardless of how informative a test is for monitoring therapeutic efficacy or explaining mechanism of action. A good example of how subject misclassification can affect the evaluation of diagnostic metrics can be seen in Huang et al., 2012, which assessed interleukin (IL)-8 and IL-1 receptor agonist (IL-1Ra) as DED biomarkers [25]. In that study, the prospective criteria fully partitioned patients from controls in many DED measures (controls OSDI < 13, corneal staining = 0; DE1 OSDI ≥ 13, corneal staining < 4; DE2 OSDI ≥ 13, corneal staining = 4–7; and DE3 OSDI ≥ 13 and corneal staining >7), but resulted in sizeable overlap of IL-8 (inflammatory) and IL-1Ra (anti-inflammatory) levels between the tears of normal subjects and mild to moderate DED subjects. Conversely, a post-hoc partitioning of the patient space excluded the patients that are difficult to categorize with symptoms between OSDI 13–19 (OSDI < 13, corneal staining = 0, TBUT > 7; DE1 OSDI ≥ 20, corneal staining < 4 and TBUT ≤ 7; DE2 OSDI ≥ 20 corneal staining = 4–7, TBUT ≤ 7; and DE3 OSDI ≥ 20, corneal staining > 7, TBUT ≤ 7) that resulted in clear, significant differences between the subset of controls and mild subjects [25]. The true performance of a diagnostic metric is therefore somewhere in between the superlative performance in trials with spectrum bias and the compromised performance in trials across a broad population, using symptoms and traditional signs as inclusion criteria.

5.3.2. Selection bias

Selection bias occurs when efficacy of metrics that were used in the selection and differentiation of subjects are directly compared to a novel test that was *not* used as part of the inclusion criteria [26]. As clinical signs and symptoms are generally uncorrelated in DED, novel tests evaluated in this manner will necessarily fail. Many biomarkers (such as MMP-9, tear osmolarity, IL-1Ra, IL-8, interferon gamma-induced protein (IP)-10, S100 calcium binding protein A9) provide novel insight into disease pathogenesis [25,27–29], but because this information is unavailable from clinical observation, comparing performance of novel diagnostic metrics against the traditional signs such as staining, TBUT and symptoms will result in an apparently poor performance. This creates a paradox where, if a novel test is correlated to older metrics, it will have strong performance in a clinical trial – but there would be no need to measure the new information. Selection bias can also occur when a novel test is compared in subjects defined as having a history of DED, as these are usually based on established diagnostic tests, which the novel test is then compared to. Furthermore, trials that evaluate new markers must also prospectively align the time-courses of therapy or wash out subjects, as

different markers respond at different rates. Failure to account for therapeutic timing is also a type of selection bias that artificially rewards lagging indicators if leading indicators have already responded [30].

5.3.3. Clinically important difference

In order for a new diagnostic metric to be most useful for monitoring, the marker should a) play a direct role in the pathogenesis of the disease, b) significantly improve upon treatment with an effective therapy, with the best markers traversing a large dynamic range and c) be specific to DED. Given the inherent temporal variation in all DED signs and symptoms, knowing whether a therapy has in fact altered the distribution of a sign or symptom at a single visit is non-trivial, especially with a single additional observation on follow up.

The term Minimal Clinically Important Difference was first described by Jaeschke and colleagues in 1989 as "the smallest difference in score in the domain of interest which patients perceive as beneficial and which would mandate, in the absence of troublesome side effects and excessive cost, a change in the patient's management" [31]. Some changes are found to be statistically significant, but of a magnitude too small to be noticed by a patient or to influence clinical management decisions. While clinically important differences are subjective, possibly changing with circumstances and time, they inform sample size calculations. In the field of DED research, only the Impact of Dry Eye on Everyday Living (IDEEL) – Symptom Bother scale [32] and (OSDI) [24] questionnaires have been assessed to determine a clinically important difference (12 and 7.0–9.9 points, respectively). The differences required in clinical studies for signs and symptoms of DED and the resulting sample sizes needed for clinical studies using these metrics to determine a significance difference are described in Table 1.

Another statistical approach, in the form of the reference change value (RCV), provides a confidence interval that new observations are not simply within the statistical error of the original distribution. To calculate RCV, the percentage analytical variation of a method as measured on traceable control materials (denoted CVA; or for clinical observations CVA would be intra-observer variation) and the percentage intra-individual (within-subject) variation (denoted CVI) within a population are needed.

$$RCV = 2^{1/2}Z(CV_A^2 + CV_I^2)^{1/2}$$

The Z-score for a one-sided, 95% probability that the change in sign or symptom is "very likely real" is 1.65 [48,49]. In applying an RCV, the CVI is typically derived from the literature [48]. Since most DED metrics do not have published data with thousands of subjects from which to draw reliable CVI data (unlike clinical chemistry studies), it is recommended that one should subtract the CVA from the CVI before calculating the RCV if the two CV measures are derived from the same study, so as to avoid double-counting. If the change in a sign or symptom between visits exceeds the calculated RCV, there is a high probability that the therapy is working. Very few papers have endeavoured to estimate the RCV of different tests for DED. Fortes et al., estimated the RCV for tear osmolarity to be 13 mOsm/L [49], although they did not correct the CVI for the CVA in the same study. The Fortes estimate would require a patient with a 338 mOsm/L reading upon initial visit to measure 325 mOsm/L or below for a clinician to believe a therapy was very likely having an effect. A corrected CVI would result in a RCV of about 10 mOsm/L. The authors are not aware of any peer-reviewed studies that reported RCV for common clinical DED tests, but applying published longitudinal data [10], the CVI can be derived from the average and standard deviation of the subjects (n = 52), applying a zero CVA for

Table 1
Clinical differences to detect and resulting sample size calculation based on 2-sample *t*-test comparison with 80% power and p < 0.05 significance level http://www.statisticalsolutions.net/pssTtest_calc.php. Note that in more complex experiments, such as those requiring repeated measures analysis of variance (ANOVA), it is better to consider the number of degrees of freedom (based on both the number of treatments/visits and the number of replicates), with at least 15 recommended (such as 5 subjects being followed up 4 times during treatment [33]. As dry eye metrics often deviate from a normal distribution, it is recommended that the subject numbers are increased by 10% to compensate [34].

Test	SD of repeated measures	Healthy population mean	Clinical difference to detect	Minimum sample size per group
OSDI	6.7 on 100 point scale [35]	9.6 ± 12.2 [35] 7.8 ± 3.1 [24] 3.7 ± 6.9 [36]	MCID 4.5 to 7.3 mild/moderate & 7.3 to 13.4 severe disease [24]	14-35 mild-moderate; 4-14 severe
DEQ-5	Unknown	2.7 ± 3.2 [37]	6 (based on variation between severity classifications) [37]	Not possible
NITBUT	7.2 [38] 2.0 [36]	11.2 ± 6.8 [38] 10.4 ± 4.2 [36]	5 s	33 Tearscope 3 Keratograph 5m
FBUT	2.9 average of 2 repeats [39]	7.6 ± 10.4 [38] 9.1 ± 3.5 [36]	5 s	6
Lipid quality (Tearscope)	Unknown			Not possible
TMH	0.15 *(slit lamp)* [39] 0.05 *(Keratograph 5m)* [36]	0.29 ± 0.13 mm *(slit lamp)* [39] 0.29 ± 0.04 mm *(Keratograph 4)* [40] 0.27 ± 0.12 *(Keratograph 5m)* [36] 0.19 ± 0.02 mm *(with OCT)* [41] 0.34 ± 0.15 mm*(with OCT)* [42]	0.1 mm	36 slit lamp 4 Keratograph 5m
Bulbar Redness (Efron)	0.4 [43] 0.4–0.7 [44]	No reported means- clinically normal taken as grade 0-1	0.5 grading	6–16
Staining grading (Efron)	Only weighted k/ICC [39]	No reported means- clinically normal taken as grade 0-1	Not reported	Not possible
LWE	No repeatability studies	Grade 1 (2–4 mm horizontal staining, 25–50% sagittal staining) [45]	Not reported	Not possible
LIPCOF	No repeatability studies	Grade 1	Not reported	Not possible
Schirmer's Test (without anaesthetic)	3.9 [39] 11.3 [46]	16.8 [46] 15.5 ± 8.7 [36]	5 mm	5–41
Phenol Red	7.5 [39] 10.0 [46]	29.0 [46]	5 mm	18–32
Osmolarity (tearlab)	4.8 [47]	301mOsm/L [15] 299mOsmol/L [47]	5mOsm/L	15

Footnotes: OSDI = Ocular Surface Disease Index; DEQ-5 = Dry Eye Questionnaire − 5 item; NITBUT = non-invasive breakup time; FBUT = fluorescein breakup time; TMH = tear meniscus height; LIPCOF = Lid Parallel Conjunctival Folds; LWE = Lid Wiper Epitheliopathy.

convenience. RCVs for TBUT = 6.3 s (average over 3 months = 4.8 ± 2.7 s), 9.6/15 for Oxford corneal fluorescein staining (2.9 ± 4.1), 14.2/27 for Foulks/Bron meibomian gland grading (11.0 ± 6.1), and 55.3/100 for OSDI (34.7 ± 23.7) [10]. Like other statistical tests, the RCV should be used only as a guide and not an absolute value − the clinician still needs to take into account all available information when making a determination about therapeutic efficacy. Comparing the statistically derived RCVs to the published minimal clinically important difference suggests that the actual value is somewhere between these two approaches, less than the RCV and greater than the minimal clinically important difference.

5.3.4. Parallel testing

In order to increase sensitivity and specificity simultaneously, it is necessary to expand a diagnostic input vector to include multiple dimensions. As shown in Fig. 1d, extra observations eventually allow a clear segregation of the affected and unaffected populations at higher orders, even if there is significant overlap in lower dimensions. For example, if one wanted to classify trees based on leaf colour alone, it would be a very noisy, imprecise way to separate elm trees from oak trees. If you add in leaf shape, tree height, sap characteristics, bark texture and so forth, the task becomes more straightforward. Sensitivity is optimized in parallel testing by diagnosing disease if any one of a series of highly specific signs is measured to be abnormal [50]. Also known as a logical "OR" operator in computing, parallel sensitivity is calculated by subtracting the product of the two individual sensitivities from their sum (Sensitivity A + Sensitivity B − (Sensitivity A x Sensitivity B)), while parallel specificity is simply the product of the individual test specificities (Specificity A x Specificity B). Each additional metric will increase sensitivity, while multiplicatively reducing specificity. Low specificity tests quickly degrade the combined specificity. Three parallel tests at 50% sensitivity and 97% specificity achieve 87.5% sensitivity and 91.3% specificity, which is far better than any one individual test. However, if the three tests had only 90% specificity, the parallel specificity would degrade to 72.9%. Therefore, when adding markers in parallel, more specific diagnostic metrics allow for greater confidence − which is somewhat paradoxical, as most clinicians judge new diagnostic metrics based on their sensitivity, not their specificity. As an example, parallel testing of multiple tear proteins has been shown to be very effective in diagnosing DED, despite each protein marker being individually quite insensitive (≈40−60% sensitive); when used in parallel as part of a panel, the combined measurements produce greater than 90% combined sensitivity and specificity [27,50,51].

5.4. Sequence of testing

As even non-invasive tests of DED require alternation of blinking or bright illumination, the sequence of testing can affect the results. It is recommended that the tests are performed from the least to the most invasive [52].

6. Recommendations of appropriate tests for diagnosis and assessment of dry eye

This section reviews the development and enhancement of diagnostic metrics of DED, particularly since the previous TFOS DEWS report. The order in which the tests are reviewed is not a

reflection of their importance. Due to the issues highlighted in Section 5 with regard to comparing the sensitivity and specificity of tests, recommendations are based on the level of evidence combined with the invasiveness of the test and its ability to be conducted in a standard clinical setting, ideally without highly specialist instrumentation. The recommended diagnostic 'homeostasis marker' tests are the minimum data set to be collected from all patients identified by the screening questionnaire (as many patients do not elicit symptoms unless specifically asked) and in all DED clinical trials. However, additional DED metrics should be applied to identify the subtype of DED and the specific aspects (such as inflammatory markers or environmental triggers) relevant to a clinical trial.

6.1. Symptoms

As in the previous TFOS DEWS definition of DED [53], the current TFOS DEWS II definition for DED mentions the presence of ocular surface symptoms and other signs of DED [4]. Although the relationship between symptoms and signs of DED is not linear and varies across individuals and types of DED [54], the ability to accurately quantify ocular surface symptoms is an important screening tool that can assist in establishing the medical necessity for additional DED evaluation. It is also critical for monitoring the progression of the condition and response to treatments. In this regard, symptom measurements are very similar to clinical signs of DED. It is therefore recommended that a validated symptom questionnaire be administered at the beginning of the patient interaction.

6.1.1. Current questionnaires

In the clinical setting, symptoms or other subjective reports are typically captured through the patient case history [55,56]. Symptoms reported during non-scripted verbal interviews are very difficult to standardize and quantify. To enhance standardization in clinical research, symptoms are typically gathered through the use of questionnaire instruments that are most often self-administered by the patient or research subject without input from the clinician or researcher. In DED, these instruments either measure ocular surface or vision symptoms associated with DED, the impact of DED on everyday function and on health-related quality of life. Table 2 gives a summary of the most frequently used DED questionnaires, their original and recent citations, and the forms of validation supported by the literature cited.

For questionnaires that are additionally intended as outcome measures for registration studies at the US Food and Drug Administration (FDA), an FDA guidance document describes a path for the development of a Patient Reported Outcome (PRO) [84]. For most DED research and clinical care, the majority of symptom tools focus primarily on the measurement of symptoms associated with DED, and these instruments, while valid, do not follow the full psychometric development plan for PROs. However, even for symptom questionnaires that are not supporting FDA claims, it is critical that they be validated for their discriminative ability. A recent thorough review by Guillemin and co-workers in 2012 covers the topic of questionnaire validation, and strengths and weaknesses of many DED questionnaires [85].

It is helpful if instruments are also shown to be reproducible and responsive to change in the DED condition. For clinicians, it can be helpful to have a published diagnostic score criteria to screen patients who may need further testing. Table 2 covers these aspects of the DED instruments currently in use. A few of these questionnaires are undergoing translation for use in other populations [65,67]. New DED questionnaires are undoubtedly in development, and they can all be assessed for the features cited here.

6.1.2. Diagnostic test recommendation and technique

In general, the Ocular Surface Disease Index (OSDI) is the most widely used questionnaire for DED clinical trials. The OSDI measures frequency of experiencing symptoms, environmental triggers and vision related quality of life. Many other questionnaires establish concurrent validity against the OSDI in recent publications. The consensus view of the committee was to use the OSDI due to its strong establishment in the field or the DEQ-5 due to its short length and discriminative ability [37]. The continuous nature of visual analogue scales is attractive for clinical trials compared to discrete Likert-based question rating, so questionnaires such as the severity scale of the Symptoms Analysis in Dry Eye (SANDE) should be considered for repeated comfort assessment.

6.2. Visual disturbance

6.2.1. Current tests

6.2.1.1. Symptoms.
A number of patient-reported outcome questionnaires have been developed which have items or subscales that assess patients' visual experiences of DED. These include:

6.2.1.1.1. Ocular Surface Disease Index (OSDI). The OSDI includes 6 questions related to visual disturbance (blurred vision, or poor vision) or visual function (problems reading, driving at night, working on a computer, or watching TV). A study showed that the DED group of 87 patients had worse OSDI composite and subscale scores for vision-related function, compared to a group of 71 patients without DED [86].

6.2.1.1.2. Dry Eye Questionnaire (DEQ-5). The DEQ has 4 questions related to visual disturbance, including the frequency of visual changes, how noticeable the visual disturbance is in the morning and at night, as well as how much the visual fluctuation bothers the patients. Visual symptoms generally increase in intensity over the day, suggesting that open-eye conditions might affect symptom progression [58]. One study using the DEQ found that 10% of patients with non-Sjögren syndrome DED and 30% of patients with Sjögren syndrome complained of impaired vision while others reported that between 42% and 80% of patients with primary Sjögren syndrome experienced "disturbances in daily vision" [18,87,88].

6.2.1.1.3. Impact of Dry Eye on Everyday Living (IDEEL). The IDEEL questionnaire has 2 items related to visual disturbance including the extent to which a person is bothered by "blurry vision" or "sensitivity to light, glare, and/or wind". Statistically significant differences in responses to the IDEEL questionnaire scores across varying levels of DED severity have been observed [89].

6.2.1.1.4. National Eye Institute's Visual Function Questionnaire (NEI VFQ-25). The National Eye Institute's Visual Function Questionnaire (NEI VFQ-25) is a generic visual function questionnaire with seven visual domains including general vision, distance vision, peripheral vision, driving, near vision, color vision, and ocular pain. DED patients have poorer NEI VFQ-25 scores for the subscales of general health, general vision, ocular pain, short distance vision activities, long distance vision activities, vision related social function, vision related mental health, vision related role difficulties, vision related dependency, and driving [86,90].

6.2.1.1.5. Dry eye-related quality-of-Life Score (DEQS). The Dry Eye-Related Quality-of-Life Score (DEQS) questionnaire developed in Japan has shown strong correlations with 4 subscales (Ocular Pain, Near Vision, Distance Vision, and Mental Health) of the NEI VFQ-25 [62].

6.2.1.1.6. Computer-vision symptom scale (CVSS17). The Computer-Vision Symptom Scale (CVSS17) is a Rasch-based linear-scale that contains 17 items exploring 15 different symptoms of computer-related visual and ocular symptoms. The CVSS17 includes a broad range of symptoms such as photophobia (items A33

Table 2
Features of Dry Eye Questionnaires & Supporting Literature. Clinical utility of these questionnaires is summarised on Table 6 of the Epidemiology subcommittee report of TFOS DEWS II [57].

Name	Primary & Recent References	Dry Eye Screening Criteria	Type of Validation	Other Comments
Dry Eye Questionnaire (DEQ)	**Primary:** Begley et al. (2002) [58]	No	Discriminant ADDE	Indiana University Frequency & Intensity
5-Item Dry Eye Questionnaire (DEQ-5)	**Primary:** Chalmers et al. (2010) [37] **Recent:** Camp et al. (2015) [59] Galor et al. (2015) [60] Fernandez et al. (2013) [61]	≥6 KCS ≥12 suspect SS	Discriminant ADDE Subgroup Glaucoma Across post traumatic stress disorder, Depression	Indiana University Frequency & Intensity
Dry Eye-Related Quality-of-Life Score (DEQS)	**Primary:** Sakane et al. (2013) [62]	No	Content Face Psychometric Reproducibility	Frequency & Degree
Impact of Dry Eye on Everyday Life (IDEEL)	**Primary:** Abetz et al. (2011) [63] **Recent:** Fairchild et al. (2008) [32]	Mild 40–50 Moderate 51–63 Severe >64	Content Psychometric Discriminant ADDE Responsiveness CID = 8 Symptom Bother	Alcon Research, Ltd., MAPI Values Symptom bother only
McMonnies' Questionnaire (MQ)	**Primary:** McMonnies & Ho (1987) [64] **Recent:** Tang et al. (2016) [65]	>14.5 Dry Eye	Chinese Translation & Validation	Frequency only
Ocular Comfort Index (OCI and OCI-C)	**Primary:** Johnson & Murphy (2007) [66] **Recent:** Chao et al. (2014) [67] Golebiowski et al. (2016) [68]		Rasch scaled items Item reduction Responsiveness CID = 3 Chinese Translation & Validation MGD Female Cross-section	Frequency & Intensity
Ocular Surface Disease Index (OSDI)	**Primary:** Schiffman et al. (2000) [35] **Recent:** Amparo et al. (2015) [69] Asiedu et al. (2016) [70] Baudouin et al. (2014) [71] Finis et al., 2014 [72] Galor et al. (2015) [60] Miller et al. (2010) [73] Ogawa et al. (2013) [74]	Mild 13–22 Moderate 23-32 Severe ≥ 33	CID = 7.0–9.9 Concurrent with SANDE Concurrent with SPEED Severe ≥ 33 Concurrent with SPEED Concurrent with DEQ5 GVHD Subgroup	Allergan, Inc. Better for Research vs. SANDE Better for ATD Dry Eye vs. SPEED Frequency & Intensity
Symptom Assessment in Dry Eye (SANDE)	**Primary:** Schaumberg et al. (2007) [75] **Recent:** Amparo et al. (2015) [69] Saboo et al. (2015) [76]		Concurrent with OSDI Concurrent with OSDI, NEI-VFQ	Frequency & Intensity Visual Analogue Scale Better for Clinical vs. OSDI
Standard Patient Evaluation of Eye Dryness (SPEED)	**Primary:** Blackie et al. (2009) [77] **Recent:** Asiedu et al. (2016) [70] Finis et al. (2014) [72]		Concurrent with OSDI Concurrent with OSDI	Frequency & Intensity Better for MGD Dry Eye
Developed for Use with Contact Lens Wearers				
Contact Lens Dry Eye Questionnaire (CLDEQ)	**Primary:** Begley et al. (2001) [78] Nichols et al. (2002) [79]	Yes Screening		Frequency & Intensity
8-Item Contact Lens Dry Eye Questionnaire (CLDEQ-8)	**Primary:** Chalmers et al. (2012) [80] **Recent:** Chalmers et al. (2016) [81]	≥12 = CLD	Discriminant Concurrent with Overall Opinion of CLs CID = 3 Responsiveness Concurrent with Overall Opinion of CLs, Eye Dryness & Eye Sensitivity	Frequency & Intensity Soft Contact Lenses
Contact Lens Impact on Quality of Life (CLIQ)	**Primary:** Pesudovs et al. (2006) [82] **Recent:** Erdurmus et al. (2009) [83]	Yes QoL Keratoconus Only	Rasch scaling Across CL types	Frequency of bundled symptoms More of a contact lens related QoL questionnaire than a direct measure of symptoms

Footnotes: Abbreviations in alphabetical order: ADDE = Aqueous Deficient Dry Eye, CLD = Contact Lens Discomfort, MGD = Meibomian Gland Dysfunction, QoL = Quality of Life. CID = clinically important difference, GVHD = Graft Versus Host Disease, NEI-VFQ = National Eye Institute - Visual Function Questionnaire.

and C23) and "blinking a lot" (item A20), and has been reported to be valuable in the evaluation of computer related visual and ocular symptoms [91].

6.2.1.2. Functional tests. Conventional distance and near visual acuity testing, employing Early Treatment Diabetic Retinopathy Study (ETDRS) and Lighthouse near vision charts, showed significant deterioration in symptomatic and asymptomatic ocular surface disease (OSD) subjects, which improved temporarily with instillation of artificial tear drops [92,93]. Similar static tests that require reporting the orientation of sine wave gratings of varying contrast have also been utilised pre- and post-artificial tear instillation [94,95]. Dynamic methodologies to assess visual function in DED patients include detection of randomly located targets of differing contrast during a driving simulation [96]. Ridder et al. employed computer-generated sine-wave gratings that were briefly presented (16 msec duration), and demonstrated that DED patients exhibit a decrease in contrast sensitivity with tear film breaks [97].

Functional visual acuity (FVA) was first defined by Goto and colleagues, as functional vision for daily activities [98]. It corresponded to the visual acuity measured with the patient's habitual prescription, during 10–20 s of sustained eye opening without blinking, aided by anesthesia. To better standardize the test, a commercialized system was developed by Ishida and colleagues, with Landolt optotypes presented in one of four orientations; increasing optotyope size occurs when a previous presentation is incorrectly identified or when there is no response within the set display time (selectable from 1 to 5 s); decreasing size occurs when the answer is correct (SSC-350; Nidek, Gamagori, Japan) [99]. The visual maintenance ratio is the average FVA divided by the baseline visual acuity. FVA is reduced in DED patients, Sjögren syndrome and Steven Johnson syndrome, more than in controls, due to irregularity of the ocular surface and induced higher order aberrations, and it improves with treatment [98–101]. The application of FVA measurements in other types of DED has identified a significant decline of FVA relating to decreased tear clearance in the elderly and associated with the short BUT type of DED in office workers, atopic keratoconjunctivitis, conjunctivochalasis and in elderly drivers [102–106].

6.2.1.3. Aberrations. Initial work examined the optical and visual impact of tear breakup during periods of non-blinking by quantifying vessel contrast in the fundus images and by monitoring the psychophysical contrast sensitivity and the spatial distribution of tear thickness changes by retroillumination [107,108]. Advances in wavefront aberrometers enabled assessment of real-time changes in the ocular optics by evaluating refractive anomalies at multiple sites over time. Laser-Assisted in situ Keratomileusis (LASIK)-related dry eyes had greater optical aberrations due to increased tear film irregularity, compared to healthy controls [109]. Serial measurements of higher order and double pass (objective scatter) aberrations after a blinking in patients with DED is associated with increased HOAs resulting, in part, from superficial punctate keratitis (SPK) overlying the optical zone [110–112].

6.2.1.4. Light scatter. Scheimpflug imaging has been used to show that the ocular forward light scattering and corneal backward light scattering from the anterior cornea are greater in dry eyes than in normal eyes and that increased corneal backward light scattering in dry eyes, at least partially, again resulted from central SPK overlying the optical zone [113].

6.2.2. Diagnostic test recommendation
Visual disturbance is currently assessed subjectively through ocular symptomology questionnaires. Until well-established objective clinical measures of visual disturbance become widely available, there is no specific additional vision test that can be recommended by TFOS DEWS II for the diagnosis of DED. This does not preclude use of vision tools that are currently under development being used to enhance understanding of individual cases of dry eye.

6.3. Tear film stability

The Definition and Classification subcommittee of TFOS DEWS II have included "tear film instability" in their revised definition of DED [4]. Impaired tear film stability has been one of the fundamental diagnostic criteria for diagnosing abnormality of the tear film and many ways of evaluating tear film stability have been described [114].

6.3.1. Current tests
6.3.1.1. Tear film break-up time. In clinical practice, the most frequently employed test of tear film stability is the measurement of the tear film breakup time (TBUT); this is the interval of time that elapses between a complete blink and the appearance of the first break in the tear film [115,116].

6.3.1.2. Fluorescein breakup time. Sodium fluorescein may be instilled to enhance visibility of the tear film, when the test is referred to as the fluorescein breakup time (FBUT); however, fluorescein reduces the stability of the tear film and therefore the measurement may not be an accurate reflection of its status [117,118]. The fluorescein can be instilled in varying volumes and concentrations using either a micropipette, or more commonly impregnated strips [66]. Since controlling the volume instilled with strips may be difficult, the use of narrow (1 mm) strips and dry sterile applicators have been proposed [118–121]. A standardized methodology is also important and instructions are generally given to blink naturally three times and then to cease blinking until instructed [66]. The reference value for DED diagnosis when fluorescein is used range from a cut-off time of less than 10 s [122], to less than 5 s when smaller, more controlled volumes of fluorescein are used [123,124]. The sensitivity and specificity of the test have been reported to be 72.2% and 61.6%, respectively, in individuals with Sjögren Syndrome [88]; however, mild and moderate DED patients have a broad range of FBUT values and the diagnostic value is less certain for these DED sufferers [13,125]. A significant downfall of the measurement of FBUT is its dependence on subjective assessment of the observer and attempts have consequently been made to automate the measurement [126,127]. Despite the drawbacks of using fluorescein to assess tear film stability, FBUT still remains one of the most commonly used diagnostic tests for DED in clinical practice [128–132].

6.3.1.3. Non-invasive tear breakup time. Since tear film stability can be affected by fluorescein, temperature, humidity and air circulation, non-invasive breakup time (NIBUT) measurements have become more popular in both clinical practice and research. Many of these techniques involve the observation of the specular reflection of an illuminated grid pattern from the tear film [133], and these typically result in longer measured values of time to breakup than stability assessment techniques involving fluorescein instillation [118,134,135]. NIBUT can also be measured through observations of placido disk images that are reflected from the anterior ocular surface with many of the currently marketed corneal topography systems [136], and specific software has been developed to assess localized changes in corneal power, as an indication of surface irregularities and breakup of the tear film, with some

instruments [137–140]. Automated assessment of tear film stability is also possible with specific software on instruments such as the Keratograph (Oculus, Wetzlar, Germany), which detects and maps locations of tear breakup over time [141,142], The NIBUT recorded with automated systems was initially reported to be shorter than other subjective measurements of NIBUT, and even conventional FBUT measurements [140–142], however, a recent study described the reverse finding [143]. A standardized methodology is also important when conducting NIBUT measurements with similar instructions to blink naturally three times and then to cease blinking until instructed to blink again [66].

A different approach has been used by other groups of researchers in which high-speed videokeratoscopy is used to estimate the variance of the number of rings detected radially from the centre of the videokeratoscopic image [144–146]. The changes in this variance indicate the instability in image quality, which is directly related to the quality of the tear film, and this has been used as an estimate of the NIBUT. This technique has been further refined by Downie using the E300 corneal topographer (Medmont International Pty Ltd., Victoria, Australia) to measure Tear Film Surface Quality Break-up Time [147]. The algorithm used identifies and eliminates images with excessive movement and is able to recognize shadows arising from eyelashes.

Interferometry is also used to assess the stability of the tear film in a non-invasive manner [148]. Using this technique, the time between the blink and the first appearance of a discontinuity in the lipid layer can be measured, and instruments have been developed specifically for this purpose [38,149–152]. More recently an instrument employing interferometry has been developed to measure the thickness of the lipid layer (TearScience® LipiView®, TearScience, Morrisville, NC) [77,153]; however, this cannot be used to measure the tear breakup time since only the tear film over the lower half of the cornea is analyzed and the area of initial break can occur anywhere across the cornea, and is noted frequently at the upper lid margin [154]. Instruments that do not allow the assessment of the entire area of the cornea exposed during eye opening may fail to detect areas of tear film abnormality.

The sensitivity and specificity of the NIBUT vary according to the specific technique used, with values of 82–84% sensitivity and 76–94% specificity being reported [134,142,147]. A cut-off value of less than or equal to 10 s has been reported to be indicative of DED when viewing the reflection of an illuminated grid pattern [134]; The absolute values for breakup time have been reported to be longer for non-invasive techniques, with a mean difference of 3.7 s being reported [38]; however, when breakup times are shorter, the differences between the two techniques have been reported to be of less magnitude [155].

6.3.1.4. Thermography. Evaporation of the tear film results in a cooling of the ocular surface [156], therefore measuring the absolute temperature and the spatial and temporal changes in temperature during the inter-blink period, may be used to evaluate tear film stability. Infrared thermography is able to measure the temperature of the ocular surface in a non-invasive manner and provide an objective, quantitative output [157]. Purslow and Wolffsohn demonstrated the ocular surface temperature measured using infrared thermography is principally related to the tear film [158]. The evidence in the literature indicates that the cooling rate of the ocular surface is faster in individuals with DED than in normal eyes, which is assumed to be as a result of a greater rate of tear film evaporation [156,159–161].

Advances in instrumentation have allowed measurement of the ocular surface temperature with increasing accuracy, resolution, and speed [160–163]. Recently, thermography has been used to differentiate between DED of differing aetiologies, with the lowest temperatures and greatest cooling rates being reported for presumed aqueous deficient dry eyes, and lower rates in dry eyes of presumed evaporative aetiology [164].

Studies have also been conducted in which ocular surface temperature and FBUT have been measured concurrently [165,166]. Su et al., demonstrated that areas of ocular surface cooling and breakup were co-localized [166], and Li et al., reported a direct relationship between FBUT and ocular surface cooling, implying that localized increases in evaporation are contributing to tear film thinning and breakup [165]. Using a customized ocular surface thermography device, a method has been demonstrated in which the exact area showing temperature reduction can be determined by analysing a series of images over a period of 9 s [167]. From this analysis, a "thermal breakup area" and "thermal breakup time" can be reported. Furthermore, the subjective sensation of discomfort has been reported to occur earlier in the interblink period in patients with DED than in controls (during forced eye opening), and that the subjective symptoms were correlated to low corneal temperatures and enhanced tear evaporation [168]. Sensitivity and specificity values of around 80% have been reported [160,161].

6.3.1.5. Osmolarity variability. An in-depth review of the evidence relating to osmolarity testing in the diagnosis of DED is provided in Section 6.5.1.1; however, it is also important to consider how spatial and temporal variations in tear osmolarity might affect tear film stability. There is greater inter-eye variability of osmolarity in DED than in normals [12,15,169]. and the inter-eye differences increase with disease severity [13]. Moreover, this inter-eye variability has been shown to substantially reduce over time with successful treatment of DED [125].

While repeated measurements over a period of time were shown to be low and stable in normal subjects, DED subjects showed relatively elevated and unstable readings [170]. This finding is termed heteroscedasticity, or increasing variation with increasing value [171]. Keech et al., further reported that the variability of tear osmolarity of normal subjects was indistinguishable from the analytical variability of measurements of a control solution of known osmolarity, suggesting that normal individuals retain an effective tear film with little variation from blink-to-blink and day-to-day [170]. In contrast, the tears of individuals with DED demonstrated increasing variation and the authors speculated that this was due to "a combination of chaotic or incomplete mixing between blinks and spatially variable tear film breakup, leading to a stochastically increased evaporation rate."

In a small study conducted by Liu et al., a link was reported between hyperosmolarity and tear instability, suggesting that transient increases in tear osmolarity may be observed under conditions of tear instability [172]. More recently, Peng et al., purported that increases in evaporation, that resulted during prolonged interblink periods or as a result of environmental factors (such as increased humidity and wind speed), drive tear film breakup, and predicted "massive" increases in osmolarity at the centre of areas of rupture of the tear film [173].

Indeed variability of osmolarity has been recommended to be something that clinicians should specifically be looking for when trying to identify patients with DED [13]. Sullivan advocates that between-eye differences beyond the threshold of 8 mOsm/L should be considered an indication of the loss of tear film homeostasis that occurs with DED [171].

6.3.1.6. Tear evaporation rate. An intact lipid layer may be necessary to prevent tear film evaporation [174]. The tear film evaporation rate is used as an indicator of tear film stability [175]. Evaporation of the tear film has been measured using a number of different techniques including a vapour pressure gradient [176,177],

and the velocity of relative humidity increase (resistance hygrometry) within a goggle cup placed over the eye [178—181]. Using these techniques, higher evaporation rates between blinks have been reported to be associated with poor tear film stability [148], and DED symptoms [179,182,183]. An absent, or non-confluent lipid layer has been determined to be associated with a four-fold increase in evaporation rate [148], and a two-fold increase in evaporation rate has been reported in patients with keratoconjunctivits sicca [179]. The rate of evaporation of the tear film has also been shown to be higher in the presence of a contact lens, and the effect remains 24 h after ceasing contact lens wear [184,185]. Since the evaporation rate is dependent on ambient temperature [186], humidity [175,180,187], and time of day [181,188], and can be affected by evaporation from the skin surrounding the eye, use of tear evaporation rate as a diagnostic and monitoring tool is challenging due to variable measurements.

In an attempt to address these issues, further techniques to measure tear evaporation rate have been proposed [189—191]. Using an infrared thermography camera [192], tear evaporation rate can be measured non-invasively while excluding the influences of the surrounding skin and sealed chambers [189,191]. Rohit et al. have recently described the modification and recalibration of a dermatology instrument by attaching a swim goggle cup [190]. Using this instrument, the authors reported being able to obtain absolute rather than relative evaporation rates both with, and without, contact lens wear. Despite these developments, a "normal" tear evaporation rate has yet to be established questionning the diagnostic relevance of this measurement at the current time; in addition individual differences in evaporation rate contribute to the challenge.

6.3.2. Diagnostic test recommendation and technique

It should be emphasised that tear film stability test results are highly variable [125]. When performing tests to assess tear film stability, clinicians need to be meticulous about the procedures and factors that may influence the measurements. Thermography and tear evaporation rate evaluation are not well-established clinical techniques. Measurement of the tear breakup time with a non-invasive technique (NIBUT) is considered preferable to the FBUT [193] and the two techniques are well correlated [118,194]. Since there are several different methods for conducting the measurement, standardization is needed for consistency. The measurement should be made before any other invasive tests are conducted (such as eyelid manipulation or staining of the ocular surface). The patient should be instructed to blink naturally three times and then to cease blinking until instructions are given to blink again, and then to blink freely between measurements [66]. Where possible, an automated measurement system is recommended [193], since subjective measurements taken with a videokeratoscope and the Tearscope/Tearscope Plus (Keeler, Windsor, UK) have been shown to vary between measurement sessions and observers [38,195]. A NIBUT cut-off value of less than, or equal to, 10 s has been reported to be indicative of DED in Caucasians, when viewing the reflection of an illuminated grid pattern [134], but the cut-off value with automated measurement systems is generally shorter [141]. The difference might be attributable to the slower response rate of the observer in subjective techniques as well as the objective software detecting interference in the image capture process and interpreting these as breaks in the tear film.

6.4. Tear volume

Although not mentioned directly within the definition of DED, the tear film volume is important for ocular surface health and its loss of homeostasis (aqueous deficiency) may be at the same time a key pathogenic mechanism and a diagnostic sign in DED patients, independent of evaporative dry eye.

6.4.1. Current tests

6.4.1.1. Meniscometry (tear meniscus assessment). Meniscometry describes assessment of the tear meniscus and may take the form of a height, or a cross-sectional volume metric. The tear menisci serve as reservoirs, supplying tears to the precorneal tear film [196]. The majority of tear fluid is contained within the menisci [197], formed by the tears lying at the junctions of the bulbar conjunctiva and the margins of both the upper and lower eyelids. The quantitative assessment of the tear menisci is, at present, the most direct approach to study the tear film volume. Slit-lamp techniques to study tear meniscus height (TMH), curvature (TMR), and cross-sectional area (TMA) are widely used in clinical practice and show good diagnostic accuracy and correlations with other DED tests [198,199]. However, this approach is operator-dependent and has important limitations, mainly related to fluorescein instillation and dependence on time-from-blink, which have potential impact on the tear film characteristics. The simplest type of slit-lamp meniscometry, based on judging the meniscus height by comparison to the variable slit-lamp beam height, has shown poor inter-visit repeatability [39]. Specialized meniscometry systems, equipped with a rotatable projection system that includes a target comprising a series of black and white stripes, a half-silvered mirror, and a digital video recorder, have been developed to facilitate simple and dynamic visualization of the tear meniscus, without the need for fluorescein instillation [200—202]. Meniscometry can be influenced by time after a blink, measurement locus along the lid margin, time of day, temperature, humidity, air speed, and illumination [2,66,203].

Application software for the iPod touch (Apple Inc., Cupertino,CA) has been recently developed to create a portable digital meniscometer that generates a grating of parallel black and white bands on the display, and which is reflected from the tear film at a working distance of 50 cm. This new slit-lamp mounted digital meniscometer exhibits good reproducibility, good agreement with both conventional video-meniscometry [204] and optical coherence tomography meniscometry [205], and an ability to facilitate detection of tear meniscus changes following the instillation of artificial tears [206].

Optical Coherence Tomography (OCT) assessment of the tear meniscus, described as an emerging technology in the TFOS DEWS 2007 report [2], has been extensively studied in the last ten years [207—226]. Upper and lower TMH, TMA, TMR and tear meniscus depth are, at present, the most commonly studied parameters. Spectral-domain OCT meniscometry has shown good intra-observer and inter-observer repeatability [212,219,224], that is superior to time-domain OCT [213,226]. The measurements are instrument-dependent [213,216], and can be biased by conjunctivochalasis, LIPCOF, disorders of lid margin congruity, and apposition between the lid and ocular surface [218,227]. The main advantages of OCT meniscometry are that it is non-invasive and image acquisition is rapid and simple, however analysis of the image may be complex, time-consuming and operator-dependent [224]. The development of validated measurement software is needed, ideally allowing dynamic image analysis to minimize interfering factors related to head, eye and eyelid movements [211,224].

6.4.1.2. Phenol red thread test. The phenol red (PRT) test that received brief mention in TFOS DEWS report [2], and was removed more than 10 years ago from the Japanese DED diagnostic criteria, consists of a thin cotton thread soaked with phenol red, a pH-sensitive dye. When dry, the thread assumes a yellow color, but

when moistened by tears the thread turns red as a consequence of the slightly alkaline physiological pH of tears (between pH 7 and 8) [228]. The test is performed by hooking the folded end over the lateral one-third of the lower eyelid margin for 15 s. The small dimensions of the cotton thread should limit the chance of eliciting substantial reflex tearing [229], and the minimal amount of pH indicator soaked on the thread should minimize the irritating effect of the test, as shown by the repeatability of multiple PRT tests performed during the same session [230]. These elements suggest that PRT test provides an indirect but realistic measure of the resting tear volume [231,232]. However, some authors have reported no significant correlation between the PRT test and tear volume determined with previously established methods such as tear meniscus height measurement or fluorophotometry [230], and poor correlation between PRT and DED symptoms [17]. Conflicting data, from weak [46], to strong [233], agreement, have been recently published on the correlation between the PRT test and Schirmer test. In clinical practice, an arbitrary cut-off value of 20 mm has been adopted to differentiate DED with and without aqueous deficiency using the PRT test [234]. A cut-off of 10 mm gives a sensitivity of 25% and specificity of 93% [235]. Doughty et al. reported small and not statistically significant differences between PRT performed with open or closed eyes [236].

6.4.1.3. Schirmer test. The Schirmer test is performed by folding the Schirmer paper strip (5 × 35 mm) at the notch and hooking the folded end over the lateral one-third of the lower lid margin. The score is the measured length of wetting from the notch, after a period of 5 min. The Schirmer test without anesthesia is a well-standardized test, providing an estimation of stimulated reflex tear flow. Although some authors have reported that the Schirmer test with topical anesthesia or nasal stimulation might be more objective and reliable in DED detection [237,238], there is a lack of high level evidence data on repeatability, sensitivity and specificity for these approaches [39]. Administering the test with the patient's eyes closed may minimize the variability of results [239], reducing the influence of the vertical gaze position [240], and horizontal eye movements [241]. Several diagnostic cut-off values have been proposed, from ≤5 mm/5 min [2], to ≤ 10 mm/5 min [228], and a range of sensitivity (77% [88] − 85% [242]) and specificity (70% [88] - 83% [242]) have been reported. The combination of Schirmer and PRT tests has been proposed to improve the diagnostic accuracy, at least in patients with aqueous deficient dry eye [228].

A variation of this test, termed strip meniscometry, involves dipping a strip (made of a 25-mm polyethylene terephthalate covered with a urethane-based material with a 0.4 mm central ditch containing a nitrocellulose membrane filter paper strip impregnated in natural blue dye reservoir) for 5 s into the tear meniscus [243]. Strip meniscometry with a cut-off of ≤4 mm has a sensitivity of 84% and specificity of 58% used in isolation and up to 81% sensitivity and 99% specificity when combined with TBUT [244].

6.4.2. Diagnostic test recommendation and technique

Meniscometry (volume or height) provides a non-invasive method to indirectly assess tear volume, with moderate repeatability especially if digital imaging rather than observational techniques are adopted. It is traditional to image the meniscus in the centre of the lower eyelid without lid manipulation shortly after a blink [66]. The Schirmer test without anaesthetic remains a diagnostic test recommended for confirmed severe aqueous deficiency (such as in Sjögren syndrome) [245], but its variability and invasiveness, precludes it use as a routine diagnostic test of tear volume, especially in cases with evaporative dry eye secondary to MGD where tear quality rather than quantity is affected and any subtle

reduction in resident tear volume in the interpalpebral space will likely be masked by the reflex tearing response on insertion of the strip.

6.5. Tear film composition

6.5.1. Current tests

6.5.1.1. Tear film osmolarity. A recent review of the literature identified 163 articles published since the year 2000 relevant to the use of tear osmolarity in the diagnosis of DED [246]. Hyperosmolarity of the tear film on the ocular surface causes a significant increase in interferon gamma, in the absence of large increases from other Th1, Th2 and Th17 cytokines, which can induce epithelial cell apoptosis through the JAK/STAT signalling pathway to induce cell death [247]. Tear osmolarity has been demonstrated to have the highest correlation to disease severity of clinical DED tests [11], and has been frequently reported as the single best metric to diagnose and classify DED [12,13,246]. However, other studies have indicated current measurement techniques to be highly variable [248]. Osmolarity generally increases with disease severity [174], classified as normal (302.2 ± 8.3 mOsm/L), mild-to-moderate (315.0 ± 11.4 mOsm/L) and severe (336.4 ± 22.3 mOsm/L). More severe subjects exhibit both an increased average and increased variability between eyes and over time [11,170], making the marker heteroscedastic [170]. Various cut-off values for DED have been proposed in the literature, from 305 mOsm/L [249], to 316 mOsm/L [12], with reported sensitivities ranging from 64% to 91% [15,23,249,250], specificities from 78% to 96% [249,251], and positive predictive values ranging from 85% to 98.4% [249,252]. These data support the 316 mOsm/L cut-off as a specific threshold to better differentiate moderate to severe DED, or when used in parallel with other specific tests, while the 308 mOsm/L cut-off has become a widely accepted, more sensitive, threshold for use in general practice to help diagnose mild to moderate subjects [13,15].

6.5.1.2. Tear film ferning. Ferning occurs when the tear film is dried, typically on a glass plate. As the pattern of the tear fern depends on the composition of the tear sample, tear ferning may be a simple test for tear film quality at a gross biochemical level. The process requires a slow crystal growth rate, low solution viscosity and low impurity levels to permit free-solute diffusion. Seven to 10 min under normal room temperature (20–26 °C) and room humidity (RH up to 50%) has been recommended [253]. The crystallisation begins with the formation of a nucleus, due to the supersaturation of ions with solvent evaporation at the peripheral edge of the drop. When the sample solute is able to diffuse into areas with a lower solute concentration, normal crystals can form [253].

Healthy tear samples produce compact, dense ferning patterns, while in dry eye samples, the pattern is fragmented or absent [254]. Electrolytes may play a role in ferning as hyperosmolarity has been found to result in deteriorated ferns [249,255]. It has a high reported sensitivity and specificity in Sjögren's syndrome [256–258]. and rheumatoid-induced keratoconjunctivitis sicca [259], but the results are more variable in DED [249,254]. Tear ferning is correlated with tear film volume and weakly with tear film stability, but seems to be independent of individual tear proteins [260]. Tear ferning changes with contact lens wear have been found to have a moderately high sensitivity (78.4%) and specificity (78.4%) for predicting contact lens tolerance in a clinical setting [261]. However, other studies have found that the tear ferning test had a poor correlation with tear film stability and symptoms in contact lens wearers [262].

6.5.2. Diagnostic test recommendation and technique

Despite some potential diagnostic ability, the underlying

mechanisms responsible for producing tear ferning and their interaction with dry eye sub-types are still poorly understood and hence this cannot currently be recommended as a diagnostic test [253].

Recent data have reinforced that two values are important to note in tear osmolarity testing: the higher of the two eyes, which is more indicative of the DED process, and the difference between the two eyes, which provides insight about the instability of the tear film [13]. Using the maximum value between both eyes has been shown to provide a higher dynamic range and larger observable change after effective therapy than using the average or single eye [10,170], and this approach is approved by the FDA for commercially available tests [263]. Normal subjects have little to no diurnal change, with repeat testing at time intervals of 1 min, 15 min, 1 day, 5 days, demonstrating variation largely indistinguishable from the analytical precision of a commercial instrument ($\approx \pm 3-6$ mOsm/L) [10,170,264]. Moreover, a longitudinal study showed that tear osmolarity is the least variable of all the common signs for DED over clinically relevant time scales [10], which might seem counterintuitive, since tear osmolarity has the highest frequency of variation, changing blink-to-blink depending on the stability of the tear film and severity of disease, however the actual amplitude of variation is strongly dependent on disease severity. Inter-eye differences of normal, mild to moderate and severe DED patients were 6.9 ± 5.9 mOsm/L, 11.7 ± 10.9 mOsm/L, and 26.5 ± 22.7 mOsm/L, respectively [13]. The low variation of normal subjects contributes to the high specificity of the marker and makes it a good candidate for parallelization and therapeutic monitoring. Accordingly, normal subjects don't display elevated osmolarity, so a value over 308 mOsm/L in either eye or a difference between eyes ≥ 8 mOsm/L are good indicators of a departure from tear film homeostasis and represent a diseased ocular surface [265].

6.6. Damage to ocular surface

6.6.1. Current tests
6.6.1.1. Ocular surface staining. Punctate staining of the ocular surface is a feature of many ocular diseases and instilled dyes are extensively used in the diagnosis and management of DED. In addition, the distribution of micropunctate staining may provide an etiological clue [266]. The most frequently used dyes are sodium fluorescein, rose bengal, and lissamine green. The clinical appearance of fluorescein staining occurs whenever viable cells experience a compromise to their integrity such as a disruption in superficial cell tight junctions or defective glycocalyx [266,267]. It is suggested that there is some weak background fluorescence of health corneal epithelial cells [268]. Rose bengal stains ocular surface epithelial cells that are unprotected by mucin or glycocalyx, as well as dead or degenerated cells [269,270]. However, it stings on instillation and induces reflex tearing. In addition, it has been shown to suppress human corneal epithelial cell viability *in vitro* [271]. On the other hand, lissamine green is less toxic to the ocular surface and consequently is as well tolerated as fluorescein [272]; it stains epithelial cells only if the cell membrane is damaged (a vital dye), irrespective of the presence of mucin, whereas rose bengal, because of its cytotoxicity, produces staining irrespective of the state of cell health, once mucin is absent [273,274]; therefore lissamine green has largely replaced the use of rose bengal in evaluating ocular surface disorders [13,275]. There have been also several reports using mixtures of these dyes for simultaneous staining of the cornea and conjunctiva [272,276,277]. A solution of 2% fluorescein and 1% lissamine green has been found to be optimal in terms of comfort and staining efficacy, but is not commercially available [272]. Sequential staining and/or using more than one paper strip will increase the likelihood of observing ocular surface

damage [277,278]. Fluorescein has a peak excitation wavelength of 495 nm, whereas the commonly used 'cobalt blue' light filters of slit lamp biomicroscopes have a peak of around 450 nm [279]. The fluorescence peak is around 515 nm within the pH range of the tear film, so the yellow barrier filter required for optimum observation should band pass at around 500 nm [279]. For lissamine green, a red filter (567–634 nm) to enhance contrast against the sclera may enhance staining visibility [280]. For consistent recording of staining severity of the ocular surface, there are various grading systems including the van Bijsterveld system [242], the National Eye Institute/Industry Workshop guidelines [281], the Collaborative Longitudinal Evaluation of Keratoconus (CLEK) schema [282], the Oxford Scheme [283], the area–density combination index [284], and the Sjögren's International Collaborative Clinical Alliance ocular staining score (Table 3) [285]. Corneal and conjunctival staining have been shown to be informative markers of disease severity in the severe DED; however, staining of the ocular surface in mild/moderate DED showed poor correlation to disease severity [11]. Therefore, observing staining of the cornea and conjunctiva is considered an important aspect in the clinical analysis of severe DED.

6.6.1.2. Impression cytology. Impression cytology is a relatively simple and practical technique that has been used in the diagnosis of the ocular surface disorders such as DED, limbal stem-cell deficiency, ocular surface neoplasia, and specific viral infections [287]. During the past decade, it has become standard to study squamous metaplasia and goblet cell density of the conjunctiva for the diagnosis and monitoring of DED [288]. Cells from the first to third most superficial layers of the epithelium are removed by application of cellulose acetate filters or biopore membranes, and the cells can be subsequently analyzed by various methods including microscopy, immunocytochemistry, immunoblotting analysis, polymerase chain reaction, and flow cytometry depending on the objective of the investigation [289]. Specific examination procedures for impression cytology are described elsewhere [290]. For analyzing conjunctival impression cytology, several squamous metaplasia grading systems based on qualitative or quantitative cytological criteria are applied. The best-known methods include the systems by Nelson [291], Tseng [292], and Blades [293]. Among them, the Nelson classification system, considering the density, morphology, cytoplasmic staining affinity and nucleus/cytoplasm ratio of conjunctival epithelial and goblet cells, remains widely used [294].

6.6.1.3. Lid Parallel Conjunctival Folds (LIPCOF). Lid-parallel conjunctival folds (LIPCOF) are folds in the lateral, lower quadrant of the bulbar conjunctiva, parallel to the lower lid margin. Even though LIPCOF may represent the first mild stages of conjunctivochalasis and thus may share the same aetiology [295], they display slightly different characteristics clinically. The cross-sectional area of LIPCOF is much smaller than that of conjunctivochalasis [227,296]. LIPCOF [235,297–299] do not occur centrally as does conjunctivochalasis, and does not seem to be age related [297]. While conjunctivochalasis can be induced or increased by forceful blinks or digital pressure towards the lid margin or gaze [300], this does not appear to happen in the case of LIPCOF.

LIPCOF occur behind the temporal and nasal tear meniscus along 2/3 of the total length of the inferior tear meniscus [203], and may cause tear meniscus height measurements to be underestimated [227]. Decreased mucin production is associated with the severity of LIPCOF [298], and LIPCOF are significantly correlated with lid wiper epitheliopathy [235,298]. LIPCOF may be related to the completeness of blinking [301], blink speed and tear film viscosity [295].

Table 3
Grading scales for ocular surface staining.

Scale	Cornea	Conjunctiva	Features
van Bijsterveld system [242]	1: few separated spots 2: many separated spots 3: confluent spots	Nasal and temporal zones: 1: few separated spots 2: many separated spots 3: confluent spots	Focus of Sjögren syndrome Out of 9
National Eye Institute/Industry Workshop guidelines [281]	divided into five sectors (central, superior, inferior, nasal and temporal), each scored 0—3	divided into superior paralimbal, inferior paralimbal & peripheral area both nasally & temporally, each scored 0—3	Total 15 corneal and 9 conjunctival
Collaborative Longitudinal Evaluation of Keratoconus (CLEK) schema [282]	divided into five sectors (central, superior, inferior, nasal and temporal), each scored 0—4 in 0.5 steps	divided into four sectors (superior, inferior, nasal and temporal), each scored 0—4 in 0.5 steps	Fluorescein ICC = 0.76 Rose Vengal ICC = 0.40 [39]
area—density combination index [284]	area (A0: no punctate staining; A1: >1/3; A2: 1/3 to 2/3; A3 >2/3) & density (D0: no punctate staining; D1: sparse; D2: moderate; D3: high with lesion overlap).	NA	combined in single index e.g. A2D3
Oxford staining score [283].	Fluorescein, lissamine or rose bengal can be used; 0 to V grade dependent on intensity of punctate staining displayed pictorially across a combination of the cornea and conjunctiva.		Dots increase on a log scale between grades
ocular staining score [285].	Fluorescein 0: 0 dots 1: 1—5 dots 2: 6—30 dots 3: >30 dots	Lissamine green 0: 0—9 dots 1: 10—32 dots 2: 33—100 dots 3: >100 dots	Fluorescein extra points +1 for confluent patches, staining within pupil or filaments Out of 12 ICC ~0.90 [286]

Patients with increased LIPCOF grades are likely to suffer from DED [297,302—304]. One study, showed that combining nasal LIP-COF and non-invasive breakup time using an algorithm appeared to be the most predictive DED test combination [235]. Sensitivity of LIPCOF Sum (nasal + temporal LIPCOF) to discriminate between normal and symptomatic DED patients was reported to be 70% and specificity, 91%, for a cut-off value of 2, using a revised LIPCOF grading scale (Table 4), where the LIPCOF score is derived from the number of folds rather than the height of the folds [298,299,302]. Another group evaluated a medium predictive ability of temporal LIPCOF using the Höh et al. fold height based grading scale [297], and defined the cut-off value as 2, giving a sensitivity of 52% and specificity of 64% [303].

LIPCOF are observed, without fluorescein, on the bulbar conjunctiva in the area perpendicular to the temporal and nasal limbus, above the lower lid (temporal and nasal LIPCOF, respectively), with a slit-lamp microscope using ~25× magnification (Fig. 2) [298,299,302]. LIPCOF can be classified by different grading scales, such as a recent scale counting the number of folds [235]. Care must be taken to differentiate between parallel, permanent, conjunctival folds (LIPCOF, single folds height ~0.08 mm) and disrupted microfolds (height~0.01 mm) [298,299,302,305—307].

Researchers have also used OCT [308,309] or Scheimpflug photography to observe LIPCOF [297]. Using these instruments, additional criteria such as cross-sectional area of LIPCOF or LIPCOF coverage by the tear meniscus can be evaluated [297,308,309]. Conjunctival shrinkage has been proposed as a diagnostic feature of dry eyes [310],and has been shown to occur more in patients with dry eye symptoms, less stable tears and with ocular surface staining, but not those with MGD [311].

Table 4
Example of a LIPCOF grading scale [235].

	Grade
No conjunctival folds	0
One permanent and clear parallel fold	1
Two permanent and clear parallel folds, (normally <0.2 mm)	2
More than two permanent and clear parallel folds, (normally 0.2 mm)	3

6.6.1.4. In vivo confocal imaging. In-vivo confocal microscopy (IVCM) is a non-invasive technique that allows the evaluation of signs of ocular surface damage in DED [312,313], including decreased corneal (apex and lower periphery) [314], and conjunctival epithelial cell density [315—317], conjunctival squamous metaplasia (increased mean individual epithelial cell area, decreased nucleocytoplasmic ratio and goblet cell density) [318], and corneal nerve changes (decreased sub-basal nerve density, increased tortuosity and increased number of bead-like formations) [315,317,319—325]. Laser scanning IVCM allows easy identification of conjunctival goblet cells (although some concerns have been reported with regard to tarsal evaluation [326]), suggesting it may be a valuable tool in assessing and monitoring DED-related ocular surface damage [312,327—330]. The confocal approach seems less invasive, but as effective as impression cytology [318,331], however it has not yet been widely adopted in clinical practice and its predictive ability in the diagnosis of DED is unknown.

6.6.1.5. Ocular surface sensitivity. Cochet-Bonnet or non-contact air-jet esthesiometers have been employed to evaluate ocular surface sensitivity. Loss of corneal sensation can give rise to severe corneal epithelial disorders such as neurotrophic keratopathy [332,333]. The palpebral conjunctival sensitivity appears to be more critical than corneal sensitivity when assessing DED [334]. Corneal esthesiometry is weakly correlated to other DED tests, but increases with severity of the disease and has achieved a reported specificity of 96%, but a sensitivity of just 19% [249].

6.6.2. Diagnostic test recommendation and technique

While corneal staining is perhaps a later stage feature of DED, combination staining with fluorescein and lissamine green instilled by a moistened and saturated filter paper strip to highlight corneal and conjunctival/eyelid margin tissue damage, respectively [272,276,277,279], is recommended as the most appropriate diagnostic technique for ocular surface damage. Ophthalmic stain strips are registered as medical devices rather than pharmaceuticals in some countries and lissamine green is not a licensed product in other countries so it is acknowledged that access to these dyes in a
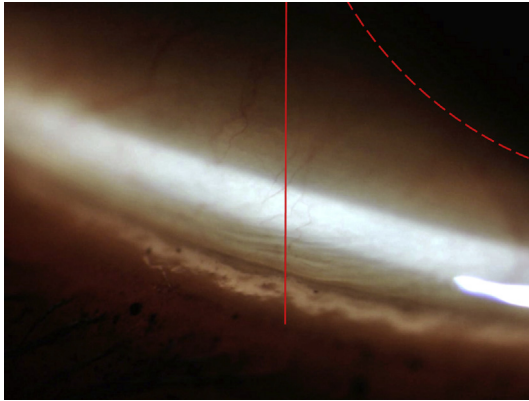
Fig. 2. LIPCOF degree 2 (Pult Scale) - dashed line indicates the corneal limbus - vertical perpendicular line indicates the appropriate area for observation.

clinical setting can be challenging; however, there have been no reported adverse effects, the benefits would appear to outweigh any risks.

6.7. Inflammation of the ocular surface

Inflammation is a recognized component of the pathophysiological mechanism of DED [4] and has been proposed to offer a stable indicator of DED severity [335]. However, inflammation is not specific to DED and can occur in other ocular or systemic disease [336,337]. In autoimmune disease, inflammation occurs in the eye as well as specific sites around the body, for example, the joints in rheumatoid arthritis. Autoimmune serum markers, including SSA and SSB are most often evaluated in Sjögren syndrome. Other relevant autoimmune diseases include systemic lupus erythematosis, mixed connective tissue disease, chronic hepatitis, Stevens Johnson syndrome and chronic graft versus host disease (GVHD). Systemic investigations used to differentially diagnosing the cause of inflammation might involve biopsy of salivary glands, flow cytometry of peripheral mononuclear blood cells, radiology or imaging of joints.

6.7.1. Current tests
6.7.1.1. Ocular/conjunctival redness. The most common clinical sign that is suggestive of ocular surface inflammation is conjunctival redness [338—340]. This is a consistent sign of conjunctival vascular dilatation and reactive change to pathological stimuli. It can occur in any disease with inflammation, not just DED, for example, in response to chemical injury, infective conjunctivitis or allergic conjunctivitis. Ocular redness can be easily detected with a pen torch or standard slit lamp biomicroscopic examination. For the purpose of diagnosis and documentation of treatment effects, more quantitative documentation methods using digital imaging analysis have been developed [341—344].

6.7.1.2. Matrix metalloproteinases. The matrix metalloproteinases (MMPs) are one of many classes of proteases secreted into the tears in DED [174,345—347]. The level of MMPs reflect the loss of ocular surface barrier function, since MMPs can destroy tight junctions in the ocular surface epithelium. MMPs are produced as inactive proenzymes and can be cleaved to become active enzymes. It is therefore important for the diagnostic test to detect enzyme activity levels and not just total tear protein levels. One development is the availability of a commercial 'point of care' diagnostic device (InflammaDry®, Rapid Pathogen Screening, Inc, Sarasota, FL, USA) which assays tear MMP-9 levels in 10 min [348]. In its current form, this assay produces a dichotomous outcome, with levels above

40 ng/ml producing a positive result, and is non-specific to the source of inflammation.

6.7.1.3. Cytokines and chemokines. The levels of tear cytokines and chemokines are important and reflect the level of epithelial disease. Certain cytokines can highlight a specific disease process, for example, elevation of Th1 and Th17 subclasses of cytokines suggest involvement of particular T lymphocyte differentiation pathways in the disease [349]. Elevation of tear Th2 cytokines, on the other hand, may suggest a more allergic-based disease, although recent evidence suggests various aspects of T cell Th1, Th2 and Th17 exist across aqueous deficient, evaporative and mixed forms of DED, with a propensity towards Th1 type T cell responses as a more global indicator of DED [350]. Since collection of tear fluid is relatively non-invasive compared to biopsies and venipuncture for serum assays, it is an attractive idea to include these as diagnostic tools [174,351].

A recent report on standard operating instructions for the tear assay of tumor necrosis factor alpha, interferon gamma, interleukin 1 beta and interleukin 6 has been published [352]. This refers to the collection, storage and repeatability of the tear assay, but with laboratory testing rather than a 'point of care' device. The operating instructions could be made even more cost-effective by reducing the need for reagents [353]. However, it has been found that tear IL-10 and IL-1β levels had significant inter-day variations, while epidermal growth factor, fractalkine, IP-10 and vascular endothelial growth factor were consistently higher in the evening compared to the mid-day measurements [354]. Such issues will affect how readily these tests are adopted in routine clinical practice. Tear chemokines such as CXCL9, -10, -11, and CXCR3 are important in the tear fluid, as they serve as ligands for specific chemokine receptors on immune cells [29,355—357]. The elevation of specific ligands may therefore imply the involvement of the specific lymphocytes in the ocular surface, without actually measuring the presence of these lymphocytes.

6.7.1.4. Ocular surface immune markers. The most commonly used ocular surface immune marker is the HLA-DR expression, a Class-II MHC antigen, which indicates a loss of the normally immune-suppressive environment of the ocular surface. Epstein has recently published standard operating instructions for impression cytology, for use in clinics and in clinical trials [358]. It was reported that sufficient conjunctival epithelial cells could be harvested for the quantification of HLA-DR using a suitable impression membrane, for example, the commercially available Eyeprim™ membrane (Opia Technology, Paris, France). The precision/repeatability of HLA-DR expression was studied and it was noted that collection, storage and shipment of specimens from distant sites were successful and storage of specimens for up to 30 days (with refrigeration) before processing did not affect results. Since the centralized laboratory was able to track large number of masked samples reliably, the authors suggest that this tool is suitable for use in randomized controlled trials of DED.

Although the authors found an increased expression level of HLA-DR associated with increased clinical severity of DED [358], a comparison with six other studies showed that the normal levels of expression of HLA-DR are very variable (ranging from 5% to 54%), and the correlation of HLA-DR expression with traditional clinical signs of DED is weak [359]. This may suggest that not all DED cases are equally inflammatory, or that the marker is non-specific for DED and indeed can involve any ocular surface inflammation. Nevertheless use of impression cytology can be useful in the documentation of specific immune cells in specific contexts of DED. For example, the quantification of neutrophil involvement in Stevens Johnson syndrome has been published [360]. Other relevant

markers of apoptosis include CAM-1, CD14$^+$, CD8$^+$ and CD4$^+$ cells [361,362].

6.7.1.5. *In vivo confocal imaging.* Corneal sub-epithelial and stromal IVCM signs of inflammation have been hypothesized and studied in DED more than 10 years ago [363,364]. More recently, IVCM has allowed examination to be extended to a number of components of the ocular surface morpho-functional unit [315,365]. Recent literature has shown significant differences between patients with DED and controls, and among different types of DED, for many presumed inflammatory parameters, including corneal dendritic cells (DC), stromal hyper-reflective (activated) cells [317,320,321,366], conjunctival hyper-reflective roundish or ovoidal (inflammatory) cells [316,326,367], and meibomian gland (MGs) acinar wall and inhomogeneous appearance between 'slices' (inflammatory infiltration) [329,368]. Some of these parameters have shown good repeatability and correlate with tear film inflammatory mediators, and other signs of DED [369]. Inflamed ocular surfaces, in immune-mediated diseases and in DED, show not only increased DC density, but also morphological DC changes, which may indicate cell maturation [364,370]. In recent research, IVCM imaging of DCs in DED was able to predict, as well as monitor, the response to anti-inflammatory drugs [370,371], and to detect sub-clinical ocular surface inflammation [372].

6.7.2. *Diagnostic test recommendation and technique*

As described, practitioners need to be aware that the ocular inflammation tests mentioned are not specific for DED. For a clinical test to be acceptable, it should be readily performed without excessive demands on technical manpower or time [373]. For this reason, research techniques such as mass spectrometry [374,375], have not been included in this section. The technical challenge involved in assessment of tear protein levels should not be underestimated. Only a very minute amount of tears can be sampled from DED patients, and since the linear range of many analytes is different, differential dilution of the collected tears may be necessary. Some of the tests may be problematic when used in a population without normal reference values. For example, many tear cytokines and even MMPs tend to increase with age [376], and age specific upper limits of the normal values have not been published, thereby potentially limiting the usefulness of the tools as diagnostic devices. However, multiplexed cytokine systems are increasingly becoming available [353]. Currently most practitioners do not include one of these tests for inflammation as a prerequisite for clinical diagnosis of DED. Certain clinical tools have been available for a long time, but the recent availability of a standard commercial platform, such as the ocular redness index within the Oculus Keratograph 5M software suggests that tools for measuring inflammation may now be within reach of many clinicians [340].

With the availability of newer immunosuppressive medications and trials concerning these drugs [377,378] it is logical that inflammation should be assessed. The exact modality used may need to be varied depending on the pathway or target cell upon which the immunosuppressive drug acts, and such diagnostic tools should be used for refining patient selection as well as monitoring after commencement of treatment. Costs of these diagnostic tests should be considered, but these should be calculated from a holistic standpoint. For example, if the tests can assist the channelling of patients to appropriate healthcare services there may be cost savings for reduced referrals.

6.8. *Eyelid aspects*

6.8.1. *Current tests*

6.8.1.1. *Anterior.* Anterior eyelid features, such as anterior

blepharitis and demodex blepharitis, are differential diagnoses and comorbidities of DED rather than diagnostic criteria and therefore are discussed in Section 9.

6.8.1.2. *Posterior*

6.8.1.2.1. *Lid wiper epitheliopathy (LWE).* A small portion of the marginal conjunctiva of the upper and lower lid acts as a wiping surface to spread the tear film over the ocular surface [379,380]. This contacting surface at the lid margin has been termed the 'lid wiper' [379]. The normal lid wiper is rich in goblet cells [381], and appears to be the most sensitive conjunctival tissue of the ocular surface [382]. The lid wiper staining with dyes such as fluorescein and lissamine green, which occurs principally in DED patients [298,299,379,383,384], has been termed lid wiper epitheliopathy (LWE) or upper lid margin staining [379,385,386]. It has been proposed that LWE is related to increased friction (direct contact between surfaces) throughout blinks [298,379,383,384], although modelling of the tribology suggests that tear film viscosity-induced hydrodynamic forces at the start of each blink are the principal cause [295]. Boundary lubrication may therefore play a key role in reducing dry eye [387]. LWE occurs on the upper and lower lids, but most studies report only upper LWE. Lower LWE in contact lens wearers has been found to be associated with DED symptoms in some studies [388], but not others [298,299].

Korb and colleagues reported that 88% of symptomatic patients had LWE but only 16% of asymptomatic patients presented with LWE [45]. Shiraishi et al. reported a higher prevalence of LWE in younger than older contact lens wearers [389]. The predictive ability of upper LWE is reported to be 48% (sensitivity) and 96% (specificity) in non-lens wearers using a cut-off value of grade 1 (based on the Korb grading scale; Table 5). In their protocols, Korb et al. recommend the use of fluorescein and lissamine green in combination to stain LWE with repeated instillation of lissamine green before the evaluation of LWE [45,278]. However in another study, LWE increased following repeated lid eversion, but not dual instillation [390] LWE can be observed immediately adjacent to the lid margin of the everted eyelid using a slit lamp biomicroscope and is most commonly classified by combining the extent of its staining, in terms of length in mm, and width relative to the lid margin width [45,235,278,298,302,379,391]. Another grading system has proposed observing the area and staining pattern [392]. A more advanced method may be to use confocal microscopy, where small hyperreflective dots, assumed to highlight inflammation, have been observed in lens wearers wearing high coefficient of friction lenses [384].

6.8.1.2.2. *Interferometry.* Oily substances spread to form a thin layer on the surface of water. Exposure of such an oily layer to adequate light results in the generation of an interferometric fringe pattern from interference from the front and back surface refractive index change reflections (from the interface with the air and the muco-aqueous tear film phase respectively). The superficial oily layer of the tear film is thought to retard evaporation of the tears, and, with the rest of the tear film, provides an optically smooth surface over the cornea [148,175,393]. The lipids produced by the meibomian glands usually distribute dynamically from the inferior to the superior region over the ocular surface and then stabilize shortly thereafter [394].

In conjunction with the surface reflection pattern and dynamics, interferometry can allow the thickness of the lipid layer of the tear film to be estimated [148]. Using slit lamp photometry to measure reflectivity, Olsen first estimated the thickness of the lipid layer of the tear film to be approximately 40 nm [395]. Since this initial analysis, single-wavelength interferometry has been applied to such measurements [396–399]. Guillon et al. developed a clinical interferometer (Tearscope; Keeler, Windsor, UK) that uses

broadband illumination to visualize the kinetics of the lipid layer of the tear film, showing that different patterns of interferometric fringe are generated according to the lipid layer thickness [154]. Goto et al. developed an algorithm for quantifying lipid layer thickness from interferometric fringe patterns [398]. The DR-1 system (Kowa, Nagoya, Japan) was also developed as an interferometer for evaluation of the kinetics of the lipid layer of the tear film in both normal subjects and patients with DED (Fig. 3). This system has revealed that lipid layer kinetics are related to the tear film condition or blink pattern [398]. Interferometry is now an established technique for clinical examination that allows visualization of the kinetics of the oily layer of the tear film.

The LipiView interferometer (TearScience, Morrisville, NC) was recently introduced as the first instrument to allow automated measurement of the thickness of the lipid layer of the tear film [153] This instrument has a sensitivity of 65.8% and a specificity of 63.4% with a cut-off value of 75-nm for the detection of MGD, but its diagnostic contribution to DED has not been established [153]. The lateral shearing interferometer has also recently been introduced for research purposes [400–403]. This latter system relies on illumination with a helium-neon laser, and analysis by fast Fourier transform, to evaluate surface irregularities of the tear film related to breakup of the lipid layer. Such instruments are likely to provide new insights into the lipid layer of the tear film and the pathophysiology of dry eye.

6.8.1.2.3. Meibography. Meibography allows observation of the silhouette of the meibomian gland morphological structure. The original technique involved white-light transillumination of everted eyelids from the skin aspect, with imaging based on black-and-white film [404], infrared film [405–407], and a near-infrared charge-coupled device (CCD) video camera [408]. Arita et al. developed a non-contact, slit lamp mounted meibography system that relies on an infrared filter and an infrared CCD video camera, in which imaging is less time-consuming than other systems (Fig. 4) [409]. Recent advances in technology have led to the development of several mobile, handheld, pen-shaped and multi-functionality systems with infrared light-emitting diodes (LEDs) fixed to infrared cameras that allow the capture of videos and images of similar quality to those obtained with earlier meibography systems [410–412].

Several different scoring scales, such as the meiboscore, have been proposed for the evaluation of meibography [409,411,413–416]. In addition, quantitative evaluation of meibomian gland area visualized by meibography has been performed [417–420]. Such quantitative evaluation has been applied to the diagnosis of MGD [419] as well as to evaluation of the effects of treatment [421,422]. Meibography alone does not appear to be sufficient for the diagnosis of MGD, but instead should be interpreted in the context of other clinical parameters [411,423–425]. The thickness of the lipid layer of the tear film measured by interferometry (LipiView) was found to be related to meibomian gland area determined by meibography [426]. Tear fluid secretion has also been shown to be positively correlated, as a compensatory mechanism, with the area devoid of meibomian glands in patients with MGD [427].

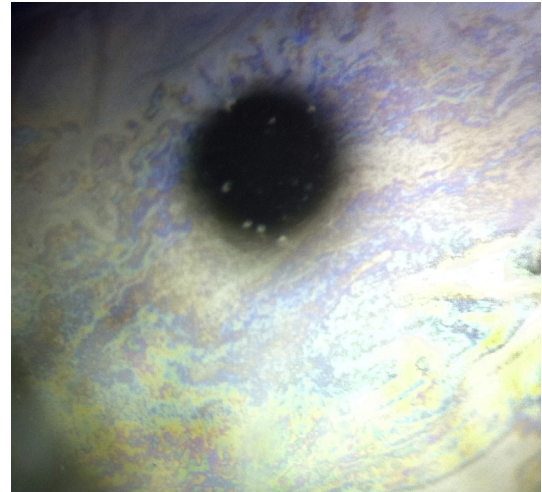Diagnostic cut-off values for the meiboscore in combination

Fig. 3. Interferometric image of the tear film lipid layer in a patient with dry eye. A multicolored interferometric fringe is observed. A video of lipid layer imaging is also available on the TFOS website.

with symptoms and lid margin abnormalities demonstrated a sensitivity of 84.9% and specificity of 96.7% for the diagnosis of MGD, in a study comparing normal eyes with those affected by obstructive MGD [423]. Meibography scales have been found to be highly reproducible [413,428], Meibography has revealed that changes in meibomian gland morphology are less pronounced in patients with ADDE than EDE [427,429]. However, shortening of meibomian gland ducts was frequently detected in wearers of contact lenses who complained of DED symptoms [430]. Establishing the diagnostic value of meibography in DED requires further study.
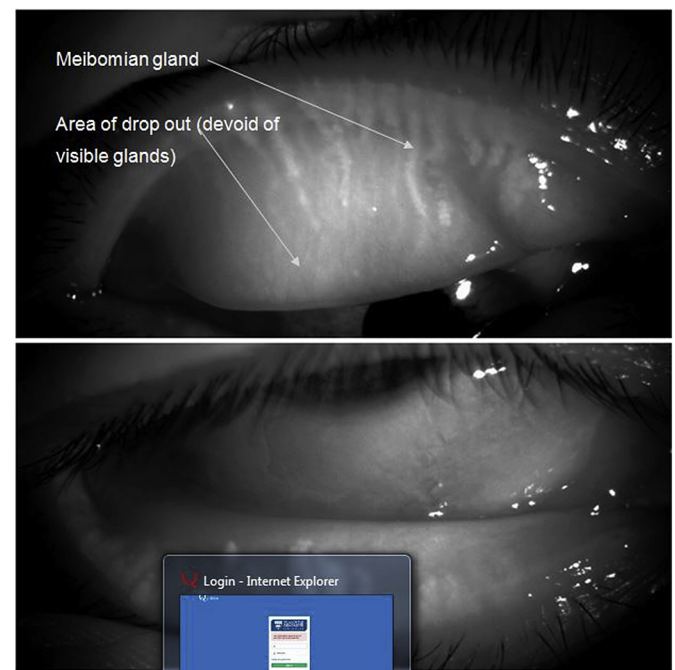

Fig. 4. Infrared images of the upper and lower eyelids obtained by non-invasive meibography in a patient with MGD. Hyper-illuminated regions correspond to meibomian glands. Note that dark areas presumed to indicate gland dropout, as well as gland shortening, are apparent.

Table 5
LWE grading scale [391].

Horizontal length of staining	Grade	Sagittal width of staining	Grade
<2 mm	0	<25% of the lid wiper	0
2-4 mm	1	25% - <50% of the lid wiper	1
5-9 mm	2	50% - <75% of the lid wiper	2
>10 mm	3	≥75% of the lid wiper	3

6.8.1.2.4. Meibomian gland expressibility/duct assessment.
Meibomian glands secrete meibum, which contains components of the lipid layer of the tear film. Meibum quantity, quality and expressibility are thought to reflect meibomian gland function. The expressibility of meibum as an indicator of meibum secretion is commonly determined by the application of digital pressure to the glands, along the length of the eyelid, through the skin surface of the eyelid [406,431,432], although more standardized procedures for expression have been reported [433]. In the normal eyelid, meibum is clear and readily expressed with gentle pressure. Conversely, the condition of meibum in patients with MGD is varied. In such individuals, meibum can lose its clarity to become cloudy and then opaque and its viscosity can be increased, becoming toothpaste-like and difficult to express in patients with severe MGD. The ranging qualities of meibum as well as its expressibility have been evaluated in various grading schemes. The number and location of expressible glands, as well the response of the glands to different levels of digitally applied pressure, have thus been scored and graded, providing information directly related to meibomian gland condition [416,433−439]. However, the diagnostic value of meibomian gland expressibility and duct appearance has not been established in DED.

6.8.1.3. In vivo confocal imaging. IVCM can be used to study the eyelid margin, to diagnose eyelid mite infestation [440,441], and to assess meibomian gland changes [329,442]. This technology has shown diagnostic benefits in obstructive MGD, providing new information about meibomian gland morphology related to specific conditions, such as contact lens wear, GVHD and atopic kerato-conjunctivitis [368,443−447], and could detect the response of meibomian glands to treatment [448,449].

6.8.1.4. Dynamic
6.8.1.4.1. Blink/lid closure analysis. Blinking is vital in maintaining optical performance and the health of the ocular surface. The blinking action clears debris, provides mechanical protection and re-forms the tear film [107,393,450−459]. Furthermore, blinking appears to be vital for meibum distribution [460], and in re-forming a proper tear film lipid layer [450,451,454]. The percentage of almost complete blinks is correlated to DED symptoms and LIPCOF, perhaps due to physical interference with spontaneous blinks [298,301], and may be related to MG morphology [461]. However, there is a broad spectrum of reported results, between 10% and 80%, for the percentage of incomplete blinks in a population of healthy individuals [454,455,462−464]. This may be due to the different measurement protocols and procedures, or variations in the visual task, or the eyelid motion detection method.

The normal spontaneous blink rate is reported to occur from 10 to 15 blinks per minute [301,465−467]. It is higher in females than in males [301,463,464], but the effect of age is controversial [301,467,468]. Incomplete blinking can result in DED and exposure keratopathy [301,452,469]. The inter-blink interval is variable between subjects, is decreased in DED and can be increased with artificial tear instillation [301,452,470]. However, the blink rate is also affected by systemic conditions such as Parkinson disease [471], and tasks such as computer work [472].

Blink speed is faster in the closing phase than the opening phase and faster for the upper lid than for the lower lid [473]. There appear to be no correlation between blink speed and either DED symptoms or tear film stability. However the upper lid velocity is positively related to LIPCOF [295,473].

Incomplete blinks can result in DED symptoms and corneal staining observable by slit lamp biomicroscope. Using fluorescein, the incomplete blink can be highlighted by a "tide line" visible as a dark line in the fluorescein pattern indicating the lower limit of movement of the upper eyelid during a recent incomplete blink [464]. More advanced methods utilise high speed video, possible now even on smart phones [474], observed from an inferior-temporal angle [301,475] However appropriate diagnostic cut-off values and sensitivity and specificity figures still require investigation.

6.8.1.4.2. Lid sensitivity. Ocular surface sensitivity plays a role in the maintenance of ocular surface homeostasis. A Cochet-Bonnet esthesiometer has been applied to evaluate lid sensitivity in several studies. Norn found that lid sensitivity was intermediate between corneal sensitivity and conjunctival sensitivity in healthy subjects [476,477], and others have reported the lower eyelid is more sensitive than the upper eyelid [478,479]. Lid margin sensitivity was found to be normal in patients with chronic blepharitis or DED [477]. It thus remains unclear whether lid sensitivity may show disease-dependent changes or whether it is unaffected in eyelid diseases.

6.8.2. Diagnostic test recommendation and technique
For subtype classifying of DED and to inform appropriate management, the presence of blepharitis, and their blink rate and completeness when a patient is performing a task such as completing a DED questionnaire unaware that the eye care practitioner is observing them, should be noted. Lipid thickness should be observed with an interferometric technique and the pattern graded. Ideally meibography should be performed along with duct observation and expressibility [480].

7. Monitoring dry eye disease progression and management

Few studies have monitored changes in DED signs and symptoms over time. New electronic technologies, such as smartphones or other handheld devices, have been tested recently to capture symptom information in "real time" rather than rely on reports from a recall period, thus aiding patient monitoring [481].

The Women's Health Study and Physicians' Health Study cohorts, revealed worsening of vision-related symptoms in 29% of the subjects. In multivariable logistic regression models for visual symptoms, spending >$20 (USD) per month on DED treatments, presence of a history of severe DED symptoms, and use of systemic beta-blockers were significantly associated with patient-reported visual worsening. Patients who reported severe symptoms of DED in the past were more likely to report worsening and to have corneal staining, suggesting that this might be a clinically relevant indicator of the probability of visual/OSD progression [482]. More prospective studies monitoring visual changes during the natural course of DED, and following treatment, are needed in the future.

8. Clinical protocol for dry eye diagnostic test battery

From Section 6, the recommended diagnostic and monitoring test battery is collated in Fig. 5. Symptoms and at least one positive result of the markers of homeostasis listed below should constitute the diagnosis of DED. If a patient has dry eye symptoms, DED is diagnosed when at least one homeostasis test result is positive. This can occur even if the practitioner does not have access to the full battery of recommended tests. However, if the practitioner has access to only a limited number of the homeostasis marker tests and these show negative results, a referral may be necessary to confirm the results of the remaining measures, to which the practitioner does not have access, before a diagnosis of DED can be excluded.

In situations where there are chronic symptoms but limited signs, that are refractory to treatment, then neuropathic pain rather than DED should be considered. Asymptomatic patients with DED type signs, unattributable to other conditions via the differential
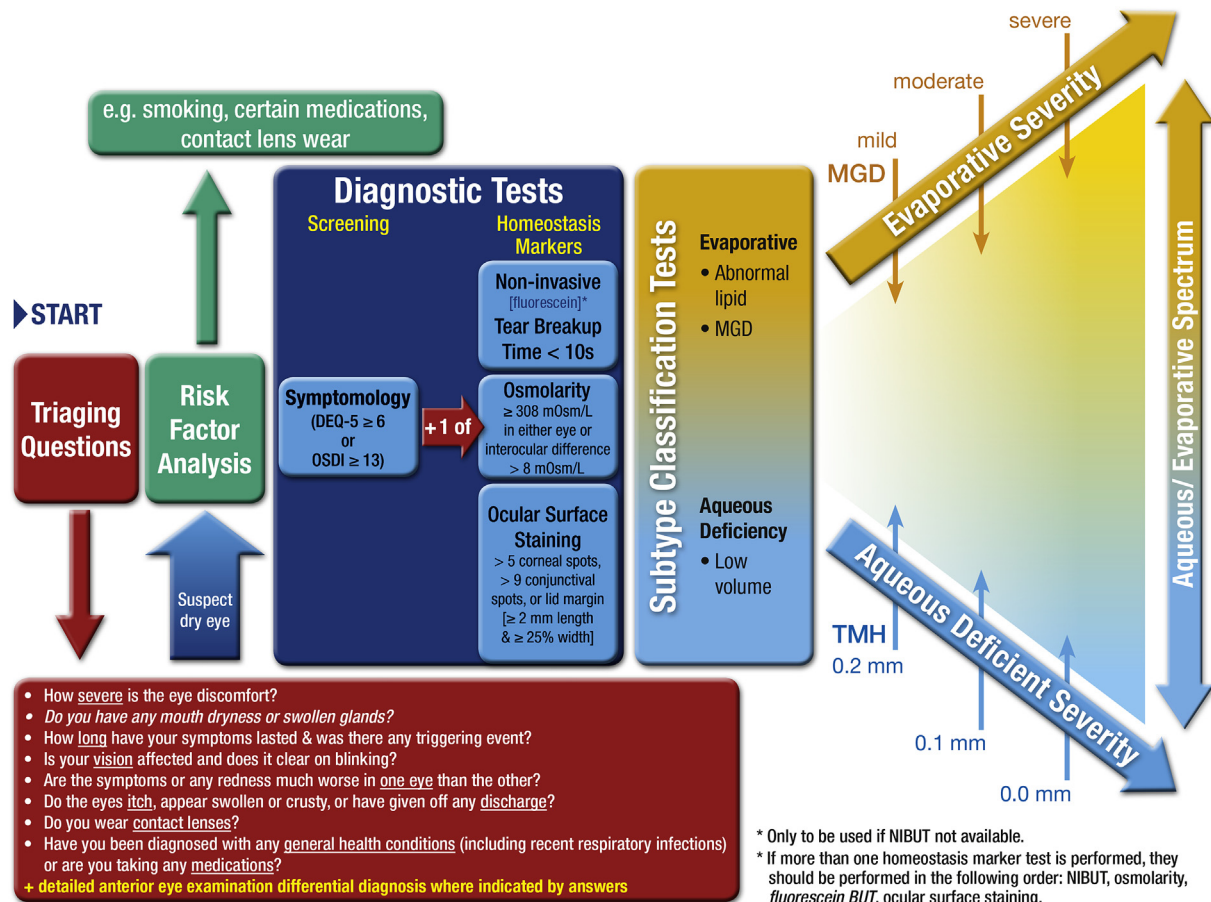
Diagnostic Tests

e.g. smoking, certain medications, contact lens wear

▶ START

Triaging Questions

Risk Factor Analysis

Suspect dry eye

Screening — Homeostasis Markers

Symptomology (DEQ-5 ≥ 6 or OSDI ≥ 13)

+1 of

Non-invasive [fluorescein]* Tear Breakup Time < 10s

Osmolarity ≥ 308 mOsm/L in either eye or interocular difference > 8 mOsm/L

Ocular Surface Staining > 5 corneal spots, > 9 conjunctival spots, or lid margin [≥ 2 mm length & ≥ 25% width]

Subtype Classification Tests

Evaporative
• Abnormal lipid
• MGD

Aqueous Deficiency
• Low volume

mild / moderate / severe MGD — Evaporative Severity

Aqueous/ Evaporative Spectrum

Aqueous Deficient Severity

TMH 0.2 mm / 0.1 mm / 0.0 mm

- How severe is the eye discomfort?
- Do you have any mouth dryness or swollen glands?
- How long have your symptoms lasted & was there any triggering event?
- Is your vision affected and does it clear on blinking?
- Are the symptoms or any redness much worse in one eye than the other?
- Do the eyes itch, appear swollen or crusty, or have given off any discharge?
- Do you wear contact lenses?
- Have you been diagnosed with any general health conditions (including recent respiratory infections) or are you taking any medications?
- + detailed anterior eye examination differential diagnosis where indicated by answers

* Only to be used if NIBUT not available.
* If more than one homeostasis marker test is performed, they should be performed in the following order: NIBUT, osmolarity, fluorescein BUT, ocular surface staining.

Fig. 5. DED diagnostic test battery. The screening DEQ-5 or OSDI confirms that a patient might have DED and triggers the diagnostic tests of non-invasive breakup time, osmolarity [measured prior to breakup time if FBUT used] and ocular surface staining with fluorescein and lissamine green (observing the cornea, conjunctiva and eyelid margin). On initial diagnosis, it is important to exclude conditions that can mimic DED with the aid of the triaging questions (Section 9) and to assess the risk factors which may inform management options [380]. Marked symptoms in the absence of clinically observable signs suggest that there may be an element of neuropathic pain. DED is a subset of OSD; signs alone may still warrant management to prevent DED manifestation and to optimise the optical corneal surface such as prior to refractive surgery or contact lens wear [4]. MGD [483] lipid thickness/dynamics and tear volume assessment and their severity inform the subtype classification of DED (as predominantly evaporative or predominantly aqueous deficient) which helps inform the management of DED. MILD MGD is indicated by a secretion grade 4-7, an expressibility grade of 1 and an amorphous/color fringes lipid pattern. MODERATE MGD is indicated by meibomian gland orifice plugging, lid margin vascularity, a secretion grade 8-12, an expressibility grade of 2 and a meshwork or wave (flow) lipid pattern. SEVERE MGD is indicated by lid margin meibomian gland orifice drop-out or displacement, a secretion grade ≥ 13, an expressibility grade of 3 and an absent, globular or abnormal colored fringes lipid pattern. Videos of these diagnostic and sub-classification techniques are available on the TFOS website. Sjögren syndrome should be suspected if the DEQ-5 score is > 12. Further testing will help identify treatment mechanisms worthy of targeting, but are beyond the scope of this Diagnostic Methodology report.

diagnosis and comorbidities triaging questions in Section 9, might still warrant prophylactic ocular surface treatment. Videos of these diagnostic as well as sub-classification techniques of MGD, lipid thickness/dynamics and tear volume are available on the TFOS website.

Tables of severity describing several signs and symptoms and (often arbitrary) cut-offs for different levels are of limited use, as features of dry eye often do not show strong association. Hence it is recommended that severity, for the purpose of selecting treatment, is based on subtype classification features (MGD, lipid thickness/dynamics and non-invasive tear volume) along with symptomology.

The recommended order and clinical practice procedural recommendations are as follows:

8.1. Symptoms

DEQ-5 (Fig. 6a) or OSDI (Fig. 6b) — self-administered [35,37]. Positive result is a DEQ-5 score ≥6 [37], or OSDI score ≥13 [35].

8.2. Tear breakup time

8.2.1. Non-invasive breakup time

Non-invasive breakup time should be performed with a method where as much of the naturally exposed cornea as possible is specularly illuminated with a light source allowing observation of breakup after a blink. Objective methods are preferred with three measurements being performed and the median value recorded. Following training, if a patient can no longer refrain from blinking before the tear film breaks up, this is typically counted as the breakup time for that measurement [194]. The lower breakup value of the two eyes should be considered in making the diagnosis. The cut-off for a positive finding can be as low as 2.7 s for automated algorithms [142], and up to 10 s for subjective observation techniques [134].

8.2.2. FBUT

FBUT can be considered when non-invasive techniques are not available, but this should follow after osmolarity measurement. Fluorescein should be instilled at the outer canthus to avoid ocular

DEQ 5

1. Questions about **EYE DISCOMFORT**:

    a. During a typical day in the past month, how often did your eyes feel discomfort?

        0 ☐ Never
        1 ☐ Rarely
        2 ☐ Sometimes
        3 ☐ Frequently
        4 ☐ Constantly

    b. When your eyes felt discomfort, how intense was this feeling of discomfort at the end of the day, within two hours of going to bed?

Never have it	Not at all intense				Very intense
0 ☐	1 ☐	2 ☐	3 ☐	4 ☐	5 ☐

2. Questions about **EYE DRYNESS**:

    a. During a typical day in the past month, **how often** did your eyes feel dry?

        0 ☐ Never
        1 ☐ Rarely
        2 ☐ Sometimes
        3 ☐ Frequently
        4 ☐ Constantly

    b. When your eyes felt dry, **how intense was this feeling of dryness** at the end of the day, within two hours of going to bed?

Never have it	Not at all intense				Very intense
0 ☐	1 ☐	2 ☐	3 ☐	4 ☐	5 ☐

3. Question about **WATERY EYES**:
During a typical day in the past month, **how often** did your eyes look or feel excessively watery?

        0 ☐ Never
        1 ☐ Rarely
        2 ☐ Sometimes
        3 ☐ Frequently
        4 ☐ Constantly

Score:	1a	+	1b	+	2a	+	2b	+	3	=	Total
	____	+	____	+	____	+	____	+	____	=	_____

Fig. 6a. Five-item Dry Eye Questionnaire (DEQ-5) reproduced with permission (Indiana University) [37].

surface damage (see below), with the excess saline on the strip shaken off or a reduced area fluorescein strip used [118]. Optimal viewing is between 1 and 3 min after instillation [279]. A positive finding has been reported to be < 10 s [13] although in some studies the average in healthy middle aged patients is noted to be lower than this [244].

8.3. Osmolarity

Osmolarity should be assessed with a temperature stabilised, calibration checked device. In the case of the Tearlab, temperature stability is achieved by having the device powered on for a sufficient period of time with test cards adjacent to the device for at least 30 min. Seat the patient with chin tilted upward and eyes directed toward the ceiling. Place one hand on the face for stabilization. Do not pull the eyelid down or away from the eye. Sample

from just above the lower eyelid tear meniscus, being careful not to press inward to avoid contact with the globe during collection. The difference between the eyes as well as the absolute measures can be diagnostic [170,171]. A positive result is considered to be ≥ 308 mOsm/L with the currently available device in either eye [13,15], or an interocular difference >8 mOsm/L [171].

8.4. Ocular surface staining

Staining (A finding in either eye is considered positive, as staining is considered a late sign of DED):

8.4.1. Lissamine green staining

Principally for assessing conjunctival and lid margin damage, a lissamine green strip is wet with saline, with the whole drop retained on the strip for at least 5 s to elute the dye. A 10 μL or ~ ¼ to

OCULAR SURFACE DISEASE INDEX©

Please answer the following questions by checking the box that best represents your answer.

Have you experienced any of the following during **the last week**:

	All of the time	Most of the time	Half of the time	Some of the time	None of the time
1. Eyes that are sensitive to light?	☐	☐	☐	☐	☐
2. Eyes that feel gritty?	☐	☐	☐	☐	☐
3. Painful or sore eyes?	☐	☐	☐	☐	☐
4. Blurred vision?	☐	☐	☐	☐	☐
5. Poor vision?	☐	☐	☐	☐	☐

Have problems with your eyes limited you in performing any of the following during **the last week**:

	All of the time	Most of the time	Half of the time	Some of the time	None of the time	N/A
6. Reading?	☐	☐	☐	☐	☐	☐
7. Driving at night?	☐	☐	☐	☐	☐	☐
8. Working with a computer or bank machine (ATM)?	☐	☐	☐	☐	☐	☐
9. Watching TV?	☐	☐	☐	☐	☐	☐

Have your eyes felt uncomfortable in any of the following situations during **the last week**:

	All of the time	Most of the time	Half of the time	Some of the time	None of the time	N/A
10. Windy conditions?	☐	☐	☐	☐	☐	☐
11. Places or areas with low humidity (very dry)?	☐	☐	☐	☐	☐	☐
12. Areas that are air conditioned?	☐	☐	☐	☐	☐	☐

Scoring Instructions

Item scoring

The total OSDI score is calculated based on the following formula:

$$OSDI = \frac{(\text{sum of severity for all questions answered}) \times (100)}{(\text{total \# of questions answered}) \times (4)}$$

where the severity was graded on a scale of
0 = none of the time,
1 = some of the time,
2 = half of the time,
3 = most of the time,
4 = all of the time.

Interpretation

A score of 100 corresponds to complete disability (a response of "all of the time" to all questions answered), while a score of 0 corresponds to no disability (a response of "none of the time" to all questions answered). Therefore, change from baseline of −12.5 corresponds to an improvement by at least one category in half of the questions answered.

Subscale Scoring

Subscales scores are computed similarly with only the questions from each subscale used to generate its own score.
Therefore, any subscales analyzed separately would also have a maximum possible score of 100.

The three subscales (vision-related function, ocular symptoms and environmental triggers) are broken out as follows:

Subscale	Questions
Vision-Related Function	4, 5, 6, 7, 8, 9
Ocular Symptoms	1, 2, 3
Enviromental Triggers	10, 11, 12

Fig. 6b. Ocular Surface Disease Index (OSDI©) Version 1 Copyright 1995 Allergan Inc. Irvine, CA, USA. All rights reserved.

½ of a drop appears to be an optimal volume if pipetting a pre-determined concentration solution [272,280]. Otherwise, a drop from the strip is instilled inside the far lower temporal lid in upgaze with the lower eyelid of the eye pulled slightly temporally to avoid damage to the conjunctival or lid wiper tissue (Fig. 7). Studies have suggested that observation should occur between 1 and 4 min post-instillation, and that observation through a red filter potentially aids visualization [272,280]. A positive score is > 9 conjunctival spots [285].

8.4.2. Fluorescein staining

Principally for assessing corneal damage, fluorescein should be instilled in a similar way, but with the excess saline on the strip shaken off to instil a minimal volume. Optimal viewing is between 1 and 3 min after instillation [279]. A positive result is > 5 corneal spots [285].

Lid wiper epitheliopathy can be observed stained with fluorescein, rose bengal or lissamine green dyes, although there seems to be a preference for just lissamine green in recent research, with viewing recommended 3–6 min after repeat instillation using 2 separate strips wet with 2 saline drops [484]. Positive is LWE of ≥ 2 mm in length and/or ≥ 25% sagittal width (excluding Marx's line) [391].

DED severity can change with the time of day so this should be

Fig. 7. Recommended location to apply ophthalmic dyes in strip form to avoid confounding damage to the conjunctiva and lid margins observed for the diagnosis of DED and its sub-classification. See video on TFOS website for further guidance.

considered in interpreting results and in monitoring DED over time [485,486].

9. Differential diagnosis & comorbidities

Based on the conditions that can mimic the signs and symptoms of DED outlined in the subsections below, administering a series of questions (Table 6) will aid in the differential diagnosis. While further investigation of possible comorbidities should not negate immediate relief management of DED type symptoms, failing to fully investigate possible comorbidities can lead to non-optimized treatment and the delayed diagnosis of causative conditions that could have serious consequences, such as the higher risk of malignancy in Sjögren syndrome [487]. If questioning by non-eye care professionals suggests DED, but recommended treatments do not result in a marked improvement in symptoms within about a one-month period, a detailed eye examination is recommended.

For those patients where the differential diagnosis history and symptoms suggests that this might not be primary DED, a full differential diagnosis should be performed using a slit lamp biomicroscope to examine the:

- eyelashes for both anterior blepharitis and signs of demodex infestation
- eyelid palpebral conjunctiva for MGD and the presence of follicles or swelling
- bulbar conjunctiva for redness pattern and signs of swelling
- cornea for ulceration, and staining should be applied to detect possible trauma

Table 6
Initial questions for the differential diagnosis of DED, indicating where more detailed observation of the ocular surface and adnexa is warranted. Medications which can cause DED are noted in the TFOS DEWS II epidemiology report [57]. Sjögren syndrome is a subtype of DED, but is included in the differential diagnosis questioning to ensure it is considered from the outset.

Question	Guidance
How severe is the eye discomfort?	• Unless severe, dry eye presents with signs of irritation such as dryness and grittiness rather than 'pain;. If pain is present, investigate for signs of trauma / infection /ulceration.
Do you have any mouth dryness or enlarged glands?	• Trigger for Sjogren's syndrome investigation
How long have your symptoms lasted & was there any triggering event?	• Dry eye is a chronic condition, present from morning to evening but generally worse at the end of the day, so if sudden onset or linked with an event, examine for trauma / infection / ulceration.
Is your vision affected and does it clear on blinking?	• Vision is generally impaired with prolonged staring, but should largely recover after a blink; a reduction in vision which does not improve with blinking, particularly with sudden onset, requires an urgent ophthalmic examination.
Are the symptoms or any redness much worse in one eye than the other?	• Dry eye is generally a bilateral condition, so if symptoms or redness are much greater in one eye than the other, detailed eye examination is required to exclude trauma & infection
Do the eyes itch, are swollen, crusty or have given off any discharge?	• Itching is usually associated with allergies while a mucopurulent discharge is associated with ocular infection
Do you wear contact lenses?	• Contact lenses can induce dry eye signs and symptoms and appropriate management strategies should be employed by the contact lens prescriber.
Have you been diagnosed with any general health conditions (including recent respiratory infections) or are you taking any medications?	• Patients should be advised to mention their symptoms to the health professionals managing their condition, as modified treatment may minimise or alleviate their dry eye.

- anterior chamber for the presence of cells or flare, indicating inflammation

9.1. Conjunctivitis

9.1.1. Allergic conjunctivitis

Symptoms of DED may be very similar to those of allergic conjunctivitis and the conditions can occur simultaneously [380]. In one study of 689 patients, clinically significant itching was found in 194 (28.2%) cases; DED was reported to be a symptom in 247 (35.8%) cases; and redness was documented in 194 (28.2%) cases [487]. Systemically, the presence of immunoglobulin E (IgE) antibodies to seasonal or perennial allergens can be documented in most cases of allergic conjunctivitis [488], and there are now some diagnostic tests available to indicate the presence of IgE biomarkers in the tear film or on the ocular surface. In addition, classical allergic conjunctivitis clinical findings, such as conjunctival chemosis, eyelid edema and conjunctival papillae, differentiate allergic from DED [489,490]. Also, allergic rhinitis is present in more than 80% of ocular allergy cases [491,492], but is not a symptom known to be associated with DED. Other findings frequently detected in allergy include a strong family history, atopic dermatitis and/or the presence of asthma [493]. Common oral pharmaceutical agents for allergy treatment have a significant drying effect on the ocular surface and may actually induce DED in patients [494,495]. A diminished tear volume, in turn, permits allergens to remain on the surface longer and may induce or exacerbate allergic conjunctivitis [496].

Giant papillary conjunctivitis (GPC) is associated with trauma to the upper tarsal plate. Contact lens wear is the primary contributor, although an exposed suture following a corneal transplant, a foreign body or ocular prosthesis also could induce GPC [497]. Symptoms of GPC and DED can overlap, including decreased contact lens wear time and mucin discharge. The key differentiating findings include large upper tarsal papillae and hyperemia with usually minimal corneal or bulbar conjunctival involvement [497]. Further, in most instances, the cause of the trauma usually is identifiable.

Atopic keratoconjunctivitis (AKC) is a chronic and potentially severe, visually threatening form of allergic eye disease. As AKC it is a bilateral, chronic, inflammatory disease, the signs and symptoms may be similar, and DED may actually be present in many of these patients. Additionally, signs of inflammation are noted on the cornea, conjunctiva and eyelids. Common symptoms include photophobia, burning, tearing, itching, mucoid discharge, and eyelid hyperemia and hypertrophy, often with greater lower eyelid involvement. Some of the more common signs that are found in both AKC and DED include SPK, conjunctival injection or hyperemia, blepharitis/MGD and tear dysfunction [498–501]. The OSD in AKC patients is characterized by greater epithelial damage and SPK [502]. Prolonged inflammation plays an important role in the progression of OSD in patients with longstanding, active AKC [415]. The hallmark findings that may help differentiate AKC from DED include conjunctivitis (potentially cicatrizing), periorbital eczema [503], corneal neovascularization that could lead to eventual conjunctivalization of the cornea, symblepharon, keratoconus and anterior polar cataracts [504,505]. Other key findings that may aid in the differential diagnosis include a strong family history of multiple allergies, atopic dermatitis, the presence of asthma and periorbital eczema [506]. In fact, it is estimated that atopic dermatitis and asthma are present in 95% and 87% of AKC patients, respectively [497].

Vernal keratoconjunctivitis (VKC) causes rapid fluorescein breakup time, SPK associated with sodium fluorescein staining and increased conjunctival lissamine green staining [507]. Patients with VKC often report severe symptoms, including intense itching, burning, epiphora, conjunctival injection and photophobia [508,509]. Clinically, VKC is associated with the presence of large cobblestone papillae and/or Horner-Trantas dots [510]. The condition can lead to debilitating corneal damage, including shield ulcers and scarring. Another key differentiator from DED is that this condition tends to occur in younger male patients—most notably those under age 18 [511].

9.1.2. Viral conjunctivitis

Viral conjunctivitis is a relatively common presentation that affects patients of all ages, including the ages during which DED is most frequent. The majority of viral conjunctivitis cases involve the highly contagious adenovirus (65–90%) [512]. Adenovirus is capable of surviving for long periods on environmental surfaces and takes a long time to shed, giving it an incubation period of 4–10 days before it is clinically observable [513]. In addition to the two types of adenovirus; pharyngoconjunctival fever (PCF) and epidemic keratoconjunctivitis (EKC), other viral conjunctivitis causes include herpes viruses, picornaviruses, and several systemic viral infections.

Although viral conjunctivitis has a number of findings in common with DED, such as tearing, burning, redness, irritation, photophobia and blurred vision, a number of differentiating factors also exist. Patients with viral conjunctivitis usually experience redness and irritation in one eye initially, often spreading to the fellow eye within a few days. When asked, patients also often report recent upper respiratory tract infection or close contact with someone with a red eye. Morning crusting is also common. Exam findings usually reveal a watery, mucoid discharge and red, edematous lids. Preauricular lymphadenopathy is also commonly present [514].

The term EKC is used when adenoviral eye infections invade the cornea. EKC, in particular, tends to be accompanied by periorbital edema and significant inflammation that may also involve the extaocular muscles. A follicular response is often noted on the palpebral conjunctiva. Early stage EKC presents with positive preauricular lymphadenopathy on the ipsilateral side to the eye that first manifested the conjunctivitis. Approximately one week later, the cornea typically exhibits sub-epithelial infiltrates, which account for symptoms of irritation and pain, often leading to decreased visual acuity that can last months or even years after the infection subsides [513].

PCF is a highly infectious illness with systemic symptoms including sweats, sore throat, fever and headache. Myalgia, malaise, pharyngitis, and gastrointestinal disturbances also are typical in patients with PCF. Upper respiratory tract symptoms may precede ocular findings, but not in all cases. Acute follicular conjunctivitis and regional lymphoid hyperplasia with tender, enlarged preauricular adenopathy are often also found in patients with PCF. PCF is most commonly observed in children and in groups living in close quarters, such as schools, prisons, ships, military bases and families. It is self-limiting and often dissipates within a week [514].

Herpes viruses that cause conjunctivitis include the herpes simplex virus, varicella-zoster virus, which also causes chickenpox and shingles, and Epstein-Barr virus, which also causes infectious mononucleosis. Herpes simplex virus in its primary form typically affects children and presents as a unilateral red eye. It is sometimes accompanied by a vesicular rash around the eyelid area. In the absence of ulceration or vesicles, herpes infection can be more difficult to diagnose. Secondary herpes simplex virus forms typically involve some form of keratitis in addition to the conjunctivitis. Interestingly, research suggests that dry eye is a stressor that may contribute to stromal keratitis in patients who have herpes [515].

Herpes zoster conjunctivitis is also unilateral and typically is accompanied by a rash that involves pustules, vesicles and edema/hyperemia of the surrounding skin, respecting the midline. Conjunctivitis sometimes precedes the appearance of lesions, making diagnosis more challenging in patients with this inflammatory condition [516].

The Epstein-Barr virus infects >90% of the population [517]. Initial exposure generally occurs during infancy or early childhood and produces subclinical infection. However, if exposure occurs in adolescence, it often manifests as infectious mononucleosis. Epstein-Barr virus infection of ocular structures most often results in transient follicular conjunctivitis [518] but can also manifest as DED, keratitis, uveitis, choroiditis, retinitis, oculoglandular syndrome, papillitis, and ophthalmoplegia [519]. Picornaviruses, such as enterovirus 70 and coxsackievirus A24, are highly contagious and often are the cause of epidemics. Like adenoviral conjunctivitis, picornaviruses cause an acute hemorrhagic response, although the clinical appearance is usually more severe. A number of systemic viruses—including as rubeola (measles), rubella (German measles), mumps, and influenza also frequently involve conjunctival infection [514]. In cases where clarification is desired, diagnostic tests with high sensitivity and specificity can help identify forms of viral conjunctivitis in minutes [520].

9.1.3. Bacterial conjunctivitis

Acute bacterial conjunctivitis is less common than viral and allergic conjunctivitis, but also shares several findings in common with DED. Bacterial conjunctivitis can affect patients of any age, but is most commonly found in children [521]. In adults, the more common culprits are gram-positive organisms such as *staphylococcus*, while in children bacterial conjunctivitis tends to be caused by *Haemophilus influenzae* and *streptococcus* species, with more than one organism causative in some cases [521]. As with DED, patients who have bacterial conjunctivitis may complain of irritation, foreign body sensation, burning, stinging and photophobia. However, they are often most concerned with the redness and discharge. Symptoms of bacterial conjunctivitis usually include a greater degree of conjunctival injection compared to conjunctivitis caused by viruses or DED. Also the discharge is wet and mucopurulent, rather than dry and crusty, and patients often complain of matting or adherence of the eyelids, especially in the morning. Bacterial conjunctivitis can be can be unilateral or bilateral and can sometimes be accompanied by systemic findings, especially in children. Systemic symptoms might include fever, malaise, purulent rhinorrhea and a respiratory infection. Otitis media is also common in children and is highly indicative of *H. influenzae* infection [522]. In some cases, bacterial conjunctivitis is accompanied by a red sheen around the eyelids, which is indicative of preseptal cellulitis.

9.2. Anterior blepharitis

Inflammation of the eyelids can result from infection by, or allergic reaction to, external agents. The clinical features of blepharitis include redness, exanthema, sores, eschar, swelling, and bullous formation. Blepharitis is classified according to its anatomic location. Anterior blepharitis affects the base of the eyelashes, eyelash follicles, and/or eyelid skin. Inflammation of follicles is categorized as marginal blepharitis, whereas that of eyelid skin is blepharo-dermatitis. The pathogenesis of anterior blepharitis is infectious or noninfectious in nature, and so the location and cause of the condition should be considered for diagnosis [523]. Clinical features of anterior blepharitis often overlap those of DED [524]. Recurrent blepharitis can cause DED, thus observation of the eyelid is important for adequate diagnosis of DED. Tear meniscus, tear film breakup time and pattern, foamy discharge and debris in the tear film should be observed [524], along with eyelids position (i.e., ectropion and entropion), eyelid closure (i.e., lagophthalmos), blink response and the anterior eyelid margin (noting any collarettes around eyelashes). Staphylococcal or seborrheic anterior blepharitis are linked to ADDE [482,524] in 50—75% of cases [525,526], perhaps due to the decreased tear volume supporting less lysozyme or immunoglobulins [526]. Definitive diagnosis is made by identification of the responsible microorganism or allergen. There are no specific clinical diagnostic tests for blepharitis. However, cultures of the eyelid margins may be indicated for patients who have recurrent anterior blepharitis with severe inflammation as well as for patients who are not responding to therapy [524].

9.3. Demodex

Demodex mites are common elongated microscopic ectoparasites that live on the surface of the human body. Demodex infestation is related to age with 84% of the population at age 60 and 100% of those older than 70 years showing Demodex infestation [527]. Demodex can spread from the face to the eyelids, perhaps leading to blepharitis and also rosacea [527—530], which may be the link between DED and meibomian gland dysfunction [528,531—533]. However Demodex infestation can also be found in asymptomatic patients [529]. Contact lens wearers do not show higher rates of Demodex infestation than non-wearers, but the relationship with DED symptoms and signs has not been investigated [534]. Two species, *Demodex folliculorum* and *Demodex brevis* have been identified in human eyelids [529,535,536]. *Demodex folliculorum* are typically found in the lash follicles of the eyelids, whereas *Demodex brevis* burrow deep into sebaceous and meibomian glands. Sebum is thought to be their main food source and Demodex mites may consume follicular and glandular epithelial cells, which may lead to direct damage of the lid margin [529]. Demodex mites can cause blepharitis by carrying bacteria on their surface including *streptococci* and *staphylococci* [529,537]. Also the protein inside the Demodex mites and their waste products may trigger inflammatory responses likely via a delayed hypersensitivity or an innate immune response [538]. Demodex-based lid margin inflammation may result in blepharoconjunctivitis [529]. Proper treatment of ocular demodicosis may resolve blepharoconjunctivitis in adults [529,539], however its role in children remains unclear [529]. Severe cases of demodex with inflamed lid margins can affect the cornea [529,540].

Demodex can sometimes be observed in situ with high magnification slit lamp microscopy, on epilated lashes using standard light microscopy or using more advanced techniques, such as IVCM [329,440,528,529,541]. Liu et al. [529] recommend the following clinical procedure based on a comprehensive literature review:

1. Clinical history: high index of suspicion when blepharitis, conjunctivitis or keratitis in adult patients or blepharoconjunctivitis or recurrent chalazia in young patients are refractory to conventional treatments, or when there is madarosis or recurrent trichiasis.
2. Slit-lamp examination: typical cylindrical dandruff at the root of eyelashes.
3. Microscopic confirmation: detection and counting of Demodex eggs, lavae and adult mites on epilated lashes.

To avoid epilating eyelashes it has also been reported that Demodex leave the follicle and are visible by slit lamp microscopy after gentle tension is applied to the lash and the lash manually rotated with forceps, encouraging exodus of the mites and allowing the lash to "scrape out" Demodex deep within the follicle [542]. As

Demodex infestation can also occur in non-DED patients [527], its diagnostic contribution is limited.

9.4. Parasitic infections

Chlamydia is an obligate intracellular parasite and one of the most common sexually transmitted infections [543]. Trachoma (or granular conjunctivitis) is caused by chlamydia trachomatis which results in inflammation, corneal inflammation and scarring of the conjunctiva, obliterating the meibomian gland ductules and goblet cells, and inducing DED complications [544]. A genital infection with chalmydia trachomatis is also the main predisposing factor for adult inclusion conjunctivitis, which is most common in young adults who are usually asymptomatic. The key differential signs from typical DED include the generally unilateral infectious nature, which can be accompanied by corneal ulcers, subepithelial infiltrates or opacity, superior epithelial keratitis, superior pannus, conjunctival scarring, mucopurlent discharge and follicles.

9.5. Corneal and conjunctival abnormalities

The corneal epithelial barrier can be compromised in the setting of DED, and manifest clinically as punctate epithelial keratopathy/ erosions by fluorescein staining, most prominently in the interpalpebral zone. Other epithelial changes in DED can include filaments, epithelial ridges and, in late stages, keratinization. The epithelial barrier integrity, however, may be compromised due to other non-DED etiologies, which can also lead to epithelial changes and corneal fluorescein staining (Table 7). These conditions often co-exist with DED and may contribute to the OSD. It can sometimes be challenging to determine whether the main underlying reason for the epithelial disease is DED, other etiologies or both.

Certain clinical features help to distinguish these other epithelial abnormalities from those that are directly related to the loss of tear film homeostasis. The history is sometimes helpful; particularly, patients might have a history of contact lens wear [545], use of multiple eye drops or exposure to toxic chemical agents [495,546]. More importantly, the clinical examination often provides additional information to alert the clinician. Specifically, the pattern and location of the epithelial changes (particularly fluorescein staining) can provide critical diagnostic clues that help distinguish DED from other alternative (or concomitant) conditions affecting the corneal epithelium. For instance, fluorescein staining in a "whorl" pattern can be seen in the setting of epithelial stress (such as toxicity from medications) [495] or conjunctivalization of the cornea due to limbal stem cell deficiency [545,547]. Likewise, fluorescein staining in the superior cornea, which is not typical for DED, may be seen in conditions such as superior limbic keratoconjunctivitis [548], floppy eyelid syndrome [549], and contact lens wear [545].

Conjunctival disease may be another co-morbid condition in patients with DED. One important disorder that can symptomatically mimic DED, and often co-exist and contribute to the patient's tear film instability, is conjunctivochalasis [550,551]. In addition to the clinical findings, the lack of response to standard DED therapies

further raises the suspicion and the need to address this co-existing condition. Other critical signs of co-existing conjunctival disease are cicatricial changes (sub-epithelial scarring, fornix foreshortening, cicatricial entropion/trichiasis, and in later stages symblepharon and keratinization) [380,552]. These findings may be a manifestation of underlying systemic diseases such as mucous membrane pemphigoid (also known as ocular cicatricial pemphigoid) and chronic Stevens-Johnson syndrome [552–554]. While these conditions universally have dry eyes as part of the clinical picture, an early diagnosis is critical, as the management often requires more advanced therapies including systemic immunomodulatory therapy.

9.6. Filamentary and other keratitis, and keratopathies

Filamentary keratitis is generally a chronic corneal condition, characterized by fine strands of degenerated epithelial cells and mucus attached to the cornea at one or both ends [555]. Patients often experience foreign body sensation, grittiness, discomfort, photophobia, blepharospasm, and increased blinking. ADDE is the most common ocular condition associated with filamentary keratitis and best-practice management involves treating the underlying DED and potential mechanical removal of the corneal filaments [556]. Interstitial keratitis is any non-ulcerating inflammation of the corneal stroma, often with vascularisation, but without involvement of either the epithelium or endothelium. The underlying causes are generally infectious or immune-mediated [557]. Neurotrophic keratitis from dysfunction of the ophthalmic division of the trigeminal nerve caused by conditions such as diabetes mellitus, ocular herpes simplex, neoplasia, and ophthalmic surgery is associated with reduced aqueous production [558]. However, treatment after the early stages of the disease requires more radical treatment than primary DED such as antibiotics, antivirals, autologous serum and steroids [559]. Bullous keratopathy is a pathological condition in which small vesicles, or bullae, form in the cornea due to endothelial dysfunction. These blister-like formations undergo painful ruptures and disrupt vision. Treatment can include hyperosmotic eye drops to reduce swelling (5% sodium chloride), amniotic membranes, bandage contact lenses to reduce discomfort, antiglaucoma medications to reduce the flow of fluid into the cornea, and corneal transplants to replace the damaged tissue [560]. Hence while filamentary and other keratitis, and keratopathies can mimic some of the signs of DED, slit lamp detection of vascularisation, anterior chamber cells and flare; stromal edema generally set them apart from primary DED.

9.7. Rheumatological conditions

Eye involvement represents a common finding in patients with systemic autoimmune diseases, particularly rheumatoid arthritis, Sjögren syndrome, seronegative spondyloarthropathy, and anti-neutrophil cytoplasmic antibody-associated vasculitis. The eye is a privileged immune site, but commensal bacteria are found on the ocular surface. Eye injury may be inflammatory, vascular or

Table 7
Common causes of corneal epithelial abnormalities.

Epithelial Trauma	lid margin keratinization, trichiasis/entropion, foreign body, superior limbic keratoconjunctivitis, floppy eyelid, contact lens wear (including hypoxia)
Epithelial Toxicity	preservatives from topical medications; such as glaucoma drops, vidarabine; mitomycin-C; fluorouracil (5-FU); other chemical/environmental exposure
Limbal Stem Cell Disease	autoimmune diseases (Stevens-Johnson syndrome, mucous membrane pemphigoid), contact lens wear, chemical injury, aniridia, ectodermal dysplasia
Epithelial Dystrophies	epithelial basement membrane dystrophy. Meesman's dystrophy
Conjunctival Scarring	mucous membrane pemphigoid, chronic Stevens-Johnson syndrome, chronic atopic keratoconjunctivitis

infectious, as well as iatrogenic, but DED can also be a presenting symptom. Over half of newly presenting DED cases to a tertiary centre were secondary to a known (48%) or undiagnosed (5%) inflammatory disease, primary thyroid disorder, Sjögren syndrome or rheumatoid arthritis [561]. Sjögren syndrome is considered a sub-classification of DED [380], but requires specific diagnostic differentiation from other forms of DED to facilitate appropriate treatment and allow monitoring of potentially life-threatening complications. Unfortunately the average time to diagnose primary Sjögren syndrome from symptom onset is 6.5 years [562], despite being an independent risk factor for non-Hodgkin lymphoma [563], and the most highly associated risk factor among all rheumatic diseases for malignancy [487]. The revised international classification criteria for Sjögren syndrome, by the American-European Consensus Group Criteria, 2002 [245,564] includes one criterion of daily feeling of dry mouth for more than 3 months, recurrent or persistent swollen salivary glands as an adult, or a need to drink liquids to aid swallowing dry food, thus any of these symptoms in a patient reporting DED should instigate a referral. There are also now serological biomarker tests for Sjögren syndrome [565]. It should be noted that tests not recommended for the diagnosis of DED, such as the Schirmer test, are still recommended for the diagnosis of Sjögren syndrome [245].

9.8. Lid related disease

Lid related disease such as chalazion or infectious hordeolum, may result in DED symptoms. Other eyelid conditions such as anterior blepharitis and MGD can inform the management of DED and therefore the eyelid should always be carefully observed when DED is investigated.

9.9. Visual asthenopia

General symptoms of visual discomfort may include those linked to DED [566]. DED is the predominant cause of computer vision syndrome [567], resulting in the reporting of general visual symptoms after prolonged use of digital screens compared to equivalent paper copy tasks [568]. Incomplete blinks rather than a reduction in blink rate appears to be associated with these symptoms [569]. Differentiation from primary DED is on the basis of history informed triggers of dryness and more general symptoms such as the eyes being tired, hurting, feeling heavy, burning, straining, stinging and experiencing photophobia [91].

9.10. Graft versus host disease (GVHD)

GVHD is an immune-mediated inflammatory disease following allogeneic hematological stem cell transplantation that causes destruction of host tissues by immunocompetent cells from the donor. Typical ocular complications in the acute form of the condition are pseudomembranous conjunctivitis and acute hemorrhagic conjunctivitis in 12—17% of cases [570,571], whereas 60—90% with the chronic form develop ocular symptoms of DED [572], perhaps due to tear fluid levels of receptor agonist IL-8/CXCL8 and interferon inducible protein IP-10/CXCL10 [28]. Ocular symptoms can be minimised by a stepwise approach to treatment involving topical anti-inflammatory medications and autologous serum tears, but patients must be monitored closely, as they are prone to serious ocular complications such as corneal perforation and endophthalmitis [573].

9.11. Contact lenses

Contact lenses can induce dry eyes (termed CLIDE) and

appropriate management strategies should be employed to minimize these [495,574]. This should be distinguished from people who have diagnosed primary DED and wish to wear contact lenses where, as well as the selection of lens modality and material, non-preserved DED treatments should be considered [377].

9.12. Psychological factors

Concomitant psychosocial issues have been associated with DED. Patients with DED have been shown to have increased prevalence of sleep and mood disorders [575]. Anxiety and depression have also been reported with increased frequency in DED patients in a variety of studies [576—578]. In one population-based cross-sectional study, of over 6000 women, these findings were similarly confirmed. Subjects with a diagnosis of DED were more likely to experience severe psychological stress [odds ratio (OR) 2.5], depressive mood [OR 1.5], and anxiety [OR 1.5] [579]. In another large series of over 7000 DED patients, the adjusted OR of DED and anxiety was 2.8 and DED and the OR for depression was 2.9 [580]. Beyond depression and anxiety, it has been suggested that DED can lead to neuropathic ocular pain and this has been shown to occur with greater frequency in patients who also have comorbid chronic pain syndromes [333,581]. Post-traumatic stress disorder has also been associated with DED and may have a link via treatment medication use or the underlying disease process [61,582]. Neuropathic pain can be differentiated from a disease mechanism through the use of anaesthetic [583], although this has not been reported in relation to DED symptoms.

Specialized forms of DED, such as Sjögren syndrome, has been associated with cognitive and mood disorders [584]. Signs of these disorders signify central nervous system involvement, which is an emerging area within Sjögren syndrome understanding. Other studies have noted that patients with Sjögren syndrome self-report greater fatigue and depression, however when compared to matched controls showed no greater dysfunction on objective tests of cognition and psychomotor function [585]. Hence, a patient's perception of disease and function can be powerful. Health related quality of life has been studied in Sjögren syndrome, showing that these patients often worry about the consequences of their illness [586].

10. Emerging technologies

Lab-on-a-chip systems capable of evaluating multiple biomarkers simultaneously are being developed by several companies and hold promise for the differential diagnosis of DED as well as systemic diseases [587]. While regulators to date have shown reluctance in approving diagnostic panels in the case of OSD, the availability of these technologies are anticipated to be of transformative value to the ophthalmic communities. Future developments will include the creation of a multiplex tear assay device that incorporates the collection and handling of sub-microliter amounts of tear [588,589]. Since ocular surface oxidative stress is an important trigger of inflammation [590], another exciting development would be the evaluation of diagnostic tools for the assessment of reactive oxygen species or oxidised products in DED. Technology is needed to determine key pathophysiological indicators of dry eye, such as osmolarity and inflammation, over the whole ocular surface in real-time within the inter-blink interval to better understand the predicated localized changes and how they impact DED [591].

An additional non-invasive assessment of tear film stability has been proposed by Varikooty et al. [592] Using this technique, tear film spread and stability is quantified through the measurement of tear film particle dynamics. Video recordings are made using a slit

lamp over a ten second period and customized software allows the velocity of particles in the tear film to be calculated. How well the particles move, depends on the ease of spread of the tear film across the ocular surface and interactions between the different layers of the tear film. Currently this method of assessment is not commercially available; however, it is possible that it could be incorporated into current or future instrumentation.

Although not yet widely used in DED, IVCM is an emerging technology that appears to have several potential applications in research and in clinical practice and might prove to be a good candidate to develop and to validate predictive biomarkers and surrogate endpoints for clinical research on DED. DED can cause corneal damage and the reverse can also occur [593].

11. Summary and conclusions

The report has determined, through scientific evidence and consensus, the most appropriate (efficacious) battery of tests to diagnose and monitor DED (Fig. 5), as per the revised definition [4]. The most appropriate test order and techniques to conduct these tests in a clinical setting have been proposed. Critical, diagnostic tests (symptoms, NIBUT, osmolarity and corneal/conjunctival/lid margin staining) have been differentiated from tests that inform subtype classification aetiologies (MGD imaging/observation and expression, lipid thickness, and tear volume tests). If the diagnostic tests suggest the presence of dry eye, differential diagnostic questioning (Table 7) and further ocular examination as indicated are essential on the initial diagnostic occurrence to exclude other forms of disease which might mimic some of the signs and symptoms of DED. By managing the underlying condition, there is the potential for symptoms of dryness to be resolved or minimised.

Financial disclosures

Acknowledgements

References

[1] The definition and classification of dry eye disease: report of the definition and classification subcommittee of the international dry eye WorkShop. Ocul Surf 2007;2007(5):75—92.

[2] Methodologies to diagnose and monitor dry eye disease: report of the diagnostic methodology subcommittee of the international dry eye WorkShop. Ocul Surf 2007;2007(5):108—52.

[3] Bilkhu PS, Wolffsohn JS, Tang GW, Naroo SA. Management of dry eye in UK pharmacies. Cont Lens Anterior Eye 2014;37:382—7.

[4] Craig JP, Nichols KK, Akpek E, Caffery B, Dua HS, Joo CK, et al. TFOS DEWS II definition and classification report. Ocul Surf 2017;15(3):276—83.

[5] Ruopp MD, Perkins NJ, Whitcomb BW, Schisterman EF. Youden Index and optimal cut-point estimated from observations affected by a lower limit of detection. Biom J 2008;50:419—30.

[6] Epitropoulos AT, Matossian C, Berdy GJ, Malhotra RP, Potvin R. Effect of tear osmolarity on repeatability of keratometry for cataract surgery planning. J Cataract Refract Surg 2015;41:1672—7.

[7] Jung HH, Ji YS, Oh HJ, Yoon KC. Higher order aberrations of the corneal surface after laser subepithelial keratomileusis. Korean J Ophthalmol 2014;28:285—91.

[8] Donnenfeld ED, Solomon R, Roberts CW, Wittpenn JR, McDonald MB, Perry HD. Cyclosporine 0.05% to improve visual outcomes after multifocal intraocular lens implantation. J Cataract Refract Surg 2010;36:1095—100.

[9] Lohmann CP, Guell JL. Regression after LASIK for the treatment of myopia: the role of the corneal epithelium. Semin Ophthalmol 1998;13:79—82.

[10] Sullivan BD, Crews LA, Sonmez B, de la Paz MF, Comert E, Charoenrook V, et al. Clinical utility of objective tests for dry eye disease: variability over time and implications for clinical trials and disease management. Cornea 2012;31:1000—8.

[11] Sullivan BD, Whitmer D, Nichols KK, Tomlinson A, Foulks GN, Geerling G, Khanal S, Ramaesh K, Diaper C, McFadyen A. Tear film osmolarity: determination of a referent for dry eye diagnosis. Invest Ophthalmol Vis Sci 2010;47:4309—15.

[13] Lemp MA, Bron AJ, Baudouin C, Benitez Del Castillo JM, Geffen D, Tauber J, et al. Tear osmolarity in the diagnosis and management of dry eye disease. Am J Ophthalmol 2011;151. 792—8 e1.

[14] Sambursky R, Davitt 3rd WF, Latkany R, Tauber S, Starr C, Friedberg M, et al. Sensitivity and specificity of a point-of-care matrix metalloproteinase 9 immunoassay for diagnosing inflammation related to dry eye. JAMA Ophthalmol 2013;131:24—8.

[15] Jacobi C, Jacobi A, Kruse FE, Cursiefen C. Tear film osmolarity measurements in dry eye disease using electrical impedance technology. Cornea 2011;30: 1289—92.

[16] Kallarackal GU, Ansari EA, Amos N, Martin JC, Lane C, Camilleri JP. A comparative study to assess the clinical use of Fluorescein Meniscus Time (FMT) with Tear Break up Time (TBUT) and Schirmer's tests (ST) in the diagnosis of dry eyes. Eye (Lond) 2002;16:594—600.

[17] Moore JE, Graham JE, Goodall EA, Dartt DA, Leccisotti A, McGilligan VE, et al. Concordance between common dry eye diagnostic tests. Br J Ophthalmol 2009;93:66—72.

[18] Begley CG, Chalmers RL, Abetz L, Venkataraman K, Mertzanis P, Caffery BA, et al. The relationship between habitual patient-reported symptoms and clinical signs among patients with dry eye of varying severity. Invest Ophthalmol Vis Sci 2003;44:4753—61.

[19] Nichols KK, Nichols JJ, Mitchell GL. The lack of association between signs and symptoms in patients with dry eye disease. Cornea 2004;23:762—70.

[20] Mizuno Y, Yamada M, Miyake Y. Dry Eye Survey Group of the National Hospital Organization of J. Association between clinical diagnostic tests and

health-related quality of life surveys in patients with dry eye syndrome. Jpn J Ophthalmol 2010;54:259—65.

[21] Fuentes-Paez G, Herreras JM, Cordero Y, Almaraz A, Gonzalez MJ, Calonge M. Lack of concordance between dry eye syndrome questionnaires and diagnostic tests. Arch Soc Esp Oftalmol 2011;86:3—7.

[22] Sullivan BD, Crews LA, Messmer EM, Foulks GN, Nichols KK, Baenninger P, et al. Correlations between commonly used objective signs and symptoms for the diagnosis of dry eye disease: clinical implications. Acta Ophthalmol 2014;92:161—6.

[23] Schargus M, Ivanova S, Kakkassery V, Dick HB, Joachim S. Correlation of tear film osmolarity and 2 different MMP-9 tests with common dry eye tests in a cohort of non-dry eye patients. Cornea 2015;34:739—44.

[24] Miller KL, Walt JG, Mink DR, Satram-Hoang S, Wilson SE, Perry HD, et al. Minimal clinically important difference for the ocular surface disease index. Arch Ophthalmol 2010;128:94—101.

[25] Huang JF, Zhang Y, Rittenhouse KD, Pickering EH, McDowell MT. Evaluations of tear protein markers in dry eye disease: repeatability of measurement and correlation with disease. Invest Ophthalmol Vis Sci 2012;53:4556—64.

[26] Szalai E, Berta A, Szekanecz Z, Szucs G, Modis Jr L. Evaluation of tear osmolarity in non-Sjogren and Sjogren syndrome dry eye patients with the TearLab system. Cornea 2012;31:867—71.

[27] Soria J, Duran JA, Etxebarria J, Merayo J, Gonzalez N, Reigada R, et al. Tear proteome and protein network analyses reveal a novel pentamarker panel for tear film characterization in dry eye and meibomian gland dysfunction. J Proteomics 2013;78:94—112.

[28] Cocho L, Fernandez I, Calonge M, Martinez V, Gonzalez-Garcia MJ, Caballero D, et al. Biomarkers in ocular chronic graft versus host disease: tear cytokine- and chemokine-based predictive model. Invest Ophthalmol Vis Sci 2016;57:746—58.

[29] Enriquez-de-Salamanca A, Castellanos E, Stern ME, Fernandez I, Carreno E, Garcia-Vazquez C, et al. Tear cytokine and chemokine analysis and clinical correlations in evaporative-type dry eye disease. Mol Vis 2010;16:862—73.

[32] Fairchild CJ, Chalmers RL, Begley CG. Clinically important difference in dry eye: change in IDEEL-symptom bother. Optom Vis Sci 2008;85:699—707.

[33] Armstrong RA, Eperjesi F, Gilmartin B. The application of analysis of variance (ANOVA) to different experimental designs in optometry. Ophthalmic Physiol Opt 2002;22:248—56.

[34] Armstrong RAH, AC. Statistical Analysis in Microbiology: StatNotes. Wiley-Blackwell; 2010.

[35] Schiffman RM, Christianson MD, Jacobsen G, Hirsch JD, Reis BL. Reliability and validity of the ocular surface disease index. Arch Ophthalmol 2000;118:615—21.

[36] Tian L, Qu JH, Zhang XY, Sun XG. Repeatability and reproducibility of noninvasive keratograph 5M measurements in patients with dry eye disease. J Ophthalmol 2016;2016:8013621.

[37] Chalmers RL, Begley CG, Caffery B. Validation of the 5-Item Dry Eye Questionnaire (DEQ-5): discrimination across self-assessed severity and aqueous tear deficient dry eye diagnoses. Cont Lens Anterior Eye 2010;33:55—60.

[38] Nichols JJ, Nichols KK, Puent B, Saracino M, Mitchell GL. Evaluation of tear film interference patterns and measures of tear break-up time. Optom Vis Sci 2002;79:363—9.

[39] Nichols KK, Mitchell GL, Zadnik K. The repeatability of clinical measurements of dry eye. Cornea 2004;23:272—85.

[40] Wei A, Le Q, Hong J, Wang W, Wang F, Xu J. Assessment of lower tear meniscus. Optom Vis Sci 2016;93:1420—5.

[41] Li J, Shen M, Wang J, Ma H, Tao A, Xu S, et al. Clinical significance of tear menisci in dry eye. Eye Contact Lens 2012;38:183—7.

[42] Wang J, Palakuru JR, Aquavella JV. Correlations among upper and lower tear menisci, noninvasive tear break-up time, and the Schirmer test. Am J Ophthalmol 2008;145:795—800.

[43] Wu S, Hong J, Tian L, Cui X, Sun X, Xu J. Assessment of bulbar redness with a newly developed keratograph. Optom Vis Sci 2015;92:892—9.

[44] Efron N. Grading scales for contact lens complications. Ophthalmic Physiol Opt 1998;18:182—6.

[45] Korb D, Herman J, Blackie C, Scaffidi R, Greiner J, Exford J, et al. Prevalence of lid wiper epitheliopathy in subjects with dry eye signs and symptoms. Cornea 2010;29:377—83.

[46] Masmali A, Alqahtani TA, Alharbi A, El-Hiti GA. Comparative study of repeatability of phenol red test versus Schirmer test in normal adults in Saudi Arabia. Eye Contact Lens 2014;40:127—31.

[47] Gokhale M, Stahl U, Jalbert I. In situ osmometry: validation and effect of sample collection technique. Optom Vis Sci 2013;90:359—65.

[48] Thue G, Sandberg S. Analytical performance specifications based on how clinicians use laboratory tests. Experiences from a post-analytical external quality assessment programme. Clin Chem Lab Med 2015;53:857—62.

[49] Fortes MB, Diment BC, Di Felice U, Gunn AE, Kendall JL, Esmaeelpour M, et al. Tear fluid osmolarity as a potential marker of hydration status. Med Sci Sports Exerc 2011;43:1590—7.

[50] Versura P, Bavelloni A, Grillini M, Fresina M, Campos EC. Diagnostic performance of a tear protein panel in early dry eye. Mol Vis 2013;19:1247—57.

[51] Zhou L, Beuerman RW, Chan CM, Zhao SZ, Li XR, Yang H. Identification of tear fluid biomarkers in dry eye syndrome using iTRAQ quantitative proteomics. J Proteome Res 2009;8:4889—905.

[52] Foulks GN. Challenges and pitfalls in clinical trials of treatments for dry eye. Ocul Surf 2003;1:20—30.

[53] The definition and classification of dry eye disease: report of the definition and classification subcommittee of the international dry eye workshop. Ocul Surf 2007;5:75—92.

[54] Begley C, Chalmers RL, Abetz L, Venkataraman K, Mertzanis P, Caffery BA, et al. The relationship between habitual patient-reported symptoms and clinical signs among patients with dry eye of varying severity. Invest Ophthalmol Vis Sci 2003;44:4753—61.

[55] Nichols KKNJ, Zadnik K. Frequency of dry eye diagnostic test procedures used in various modes of ophthalmic practice. Cornea 2000;19:477—82.

[56] Korb DR. Survey of preferred tests for diagnostis of the tear film and dry eye. Cornea 2000;19:483—6.

[57] Stapleton FJ, Alves M, Bunya V, Jalbert I, Lekhanont K, Malet F, et al. TFOS DEWS II epidemiology report. Ocul Surf 2017;15(3):334—68.

[58] Begley CG, Caffery B, Chalmers RL, Mitchell GL, Dry Eye Investigation Study G. Use of the dry eye questionnaire to measure symptoms of ocular irritation in patients with aqueous tear deficient dry eye. Cornea 2002;21:664—70.

[59] Camp A, Wellik SR, Tzu JH, Feuer W, Arheart KL, Sastry A, et al. Dry eye specific quality of life in veterans using glaucoma drops. Cont Lens Anterior Eye 2015;38:220—5.

[60] Galor A, Felix ER, Feuer W, Shalabi N, Martin ER, Margolis TP, et al. Dry eye symptoms align more closely to non-ocular conditions than to tear film parameters. Br J Ophthalmol 2015;99:1126—9.

[61] Fernandez CA, Galor A, Arheart KL, Musselman DL, Venincasa VD, Florez HJ, et al. Dry eye syndrome, posttraumatic stress disorder, and depression in an older male veteran population. Invest Ophthalmol Vis Sci 2013;54:3666—72.

[62] Sakane Y, Yokoi N, Uchino M, Dogru M, Oishi T, et al. Development and validation of the dry eye-related quality-of-life score questionnaire. JAMA Ophthalmol 2013;131:1331—8.

[63] Abetz L, Rajagopalan K, Mertzanis P, Begley C, Barnes R, Chalmers R, et al. Development and validation of the impact of dry eye on everyday life (IDEEL) questionnaire, a patient-reported outcomes (PRO) measure for the assessment of the burden of dry eye on patients. Health Qual Life Outcomes 2011;9:111.

[64] McMonnies CW, Ho A. Responses to a dry eye questionnaire from a normal population. J Am Optom Assoc 1987;58:588—91.

[65] Tang F, Wang J, Tang Z, Kang M, Deng Q, Yu J. Accuracy of McMonnies questionnaire as a screening tool for Chinese ophthalmic outpatients. PLoS One 2016;11:e0153047.

[66] Johnson ME, Murphy PJ. Measurement of ocular surface irritation on a linear interval scale with the ocular comfort index. Invest Ophthalmol Vis Sci 2007;48:4451—8.

[67] Chao C, Golebiowski B, Cui Y, Stapleton F. Development of a Chinese version of the ocular comfort index. Invest Ophthalmol Vis Sci 2014;55:3562—71.

[68] Golebiowski B, Badarudin N, Eden J, You J, Hampel U, Stapleton F. Does endogenous serum oestrogen play a role in meibomian gland dysfunction in postmenopausal women with dry eye? Br J Ophthalmol 2016;101(2):218—22. http://dx.doi.org/10.1136/bjophthalmol-2016-308473.

[69] Amparo F, Schaumberg DA, Dana R. Comparison of two questionnaires for dry eye symptom assessment: the ocular surface disease index and the symptom assessment in dry eye. Ophthalmology 2015;122:1498—503.

[70] Asiedu K, Kyei S, Boampong F, Ocansey S. Symptomatic Dry Eye and Its Associated Factors: A Study of University Undergraduate Students in Ghana. Eye Contact Lens. 2016.

[71] Baudouin C, Aragona P, Van Setten G, Rolando M, Irkec M, Benitez del Castillo J, et al. Diagnosing the severity of dry eye: a clear and practical algorithm. Br J Ophthalmol 2014;98:1168—76.

[72] Finis D, Pischel N, Konig C, JHayajneh, Schrader S, et al. Comparison of the OSDI and SPEED questionnaires for the evaluation of dry eye disease in clinical routine. Ophthalmologe 2014;111:1050—6.

[73] Miller KLWJ, Mink DR, Satram-Hoang S, Wilson SE, Perry HD, Asbell PA, et al. Minimal clinically important difference for the ocular surface disease index. Arch Ophthalmol 2010;128:94—101.

[74] Ogawa Y, Kim SK, Dana R, Clayton J, Jain S, Rosenblatt MI, et al. International chronic ocular graft-vs-host-disease (GVHD) consensus group: proposed diagnostic criteria for chronic GVHD (Part I). Sci Rep 2013;3:3419.

[75] Schaumberg DA, Gulati A, Mathers WD, Clinch T, Lemp MA, Nelson JD, et al. Development and validation of a short global dry eye symptom index. Ocul Surf 2007;5:50—7.

[76] Saboo US, Amparo F, Abud TB, Schaumberg DA, Dana R. Vision-related quality of life in patients with ocular graft-versus-host disease. Ophthalmology 2015;122:1669—74.

[77] Blackie CA, Solomon JD, Scaffidi RC, Greiner JV, Lemp MA, Korb DR. The relationship between dry eye symptoms and lipid layer thickness. Cornea 2009;28:789—94.

[78] Begley CGCR, Mitchell GL, Nichols KK, Caffery B, Simpson T, DuToit R, et al. Characterization of ocular surface symptoms from optometric practices in North America. Cornea 2001;20:610—8.

[79] Nichols JJ, Mitchell GL, Nichols KK, Chalmers R, Begley C. The performance of the contact lens dry eye questionnaire as a screening survey for contact lens-related dry eye. Cornea 2002;21:469—75.

[80] Chalmers RLBC, Moody K, Hickson-Curran S. Contact lens dry eye Questionnaire-8 and overall opinion of contact lenses. Optom Vis Sci 2012;89:1435—42.

[81] Chalmers RLKL, Hickson-Curran SB, Gleason WJ. Cutoff score and responsiveness of the 8-item Contact Lens Dry Eye Questionnaire (CLDEQ-8) in a

large daily disposable contact lens registry. Cont Lens Anterior Eye 2016;39(5):342–352. http://dx.doi.org/10.1016/j.clae.2016.04.005.

[82] Pesudovs KGE, Elliott DB. The contact lens impact on quality of life (CLIQ) questionnaire: development and validation. Invest Ophthalmol Vis Sci 2006;47:2789–96.

[83] Edurmus MYE, Abdalla YF, Hammersmith KM, Rapuano CJ, Cohen EJ. Contact lens related quality of life in patients with keratoconus. Eye Contact Lens 2009;35:123–7.

[84] Services USDoHaH. Patient-reported outcome measures: use in medical product development to support labelling claims. Guidance for industry. Available at: http://www.fda.gov/dowlloads/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/UCM193282.pdf. Center for Drug Evaluaiton and Research; Center for biologics Evaluation and Research; Center for Devices and Radiological Health

[85] Guillemin IBC, Chalmers R, Baudouin C, Arnould B. Appraisal of Patient-Reported Outcome instruments available for randomized clinical trials in dry eye: revisiting the standards. Ocul Surf 2012;10:84–99.

[86] Li M, Gong L, Chapin WJ, Zhu M. Assessment of vision-related quality of life in dry eye patients. Invest Ophthalmol Vis Sci 2012;53:5722–7.

[87] Bjerrum KB. Test and symptoms in keratoconjunctivitis sicca and their correlation. Acta Ophthalmol Scand 1996;74:436–41.

[88] Vitali C, Moutsopoulos HM, Bombardieri S. The European Community Study Group on diagnostic criteria for Sjögren's Syndrome. Sensitivity and specificity of tests for ocular and oral involvement in Sjogren's syndrome. Ann Rheum Dis 1994;53:637–47.

[89] Rajagopalan K, Abetz L, Mertzanis P, Espindle D, Begley C, Chalmers R, et al. Comparing the discriminative validity of two generic and one disease-specific health-related quality of life measures in a sample of patients with dry eye. Value Health 2005;8:168–74.

[90] Nichols KK, Mitchell GL, Zadnik K. Performance and repeatability of the NEI-VFQ-25 in patients with dry eye. Cornea 2002;21:578–83.

[91] Gonzalez-Perez M, Susi R, Antona B, Barrio A, Gonzalez E. The Computer-Vision Symptom Scale (CVSS17): development and initial validation. Invest Ophthalmol Vis Sci 2014;55:4504–11.

[92] Nilforoushan MR, Latkany RA, Speaker MG. Effect of artificial tears on visual acuity. Am J Ophthalmol 2005;140:830–5.

[93] van Landingham SW, West SK, Akpek EK, Munoz B, Ramulu PY. Impact of dry eye on reading in a population-based sample of the elderly: the Salisbury Eye Evaluation. Br J Ophthalmol 2014;98:639–44.

[94] Rolando M, Iester M, Macri A, Calabria G. Low spatial-contrast sensitivity in dry eyes. Cornea 1998;17:376–9.

[95] Huang FC, Tseng SH, Shih MH, Chen FK. Effect of artificial tears on corneal surface regularity, contrast sensitivity, and glare disability in dry eyes. Ophthalmology 2002;109:1934–40.

[96] Deschamps N, Ricaud X, Rabut G, Labbe A, Baudouin C, Denoyer A. The impact of dry eye disease on visual performance while driving. Am J Ophthalmol 2013;156:184–9.

[97] Ridder WH, Tomlinson A, Huang JF, Li J. Impaired visual performance in patients with dry eye. Ocul Surf 2011;9:42–55.

[98] Goto E, Yagi Y, Matsumoto Y, Tsubota K. Impaired functional visual acuity of dry eye patients. Am J Ophthalmol 2002;133:181–6.

[99] Ishida R, Kojima T, Dogru M, Kaido M, Matsumoto Y, Tanaka M, et al. The application of a new continuous functional visual acuity measurement system in dry eye syndromes. Am J Ophthalmol 2005;139:253–8.

[100] Kaido M, Dogru M, Yamada M, Sotozono C, Kinoshita S, Shimazaki J, et al. Functional visual acuity in Stevens-Johnson syndrome. Am J Ophthalmol 2006;142:917–22.

[101] Kaido M, Matsumoto Y, Shigeno Y, Ishida R, Dogru M, Tsubota K. Corneal fluorescein staining correlates with visual function in dry eye patients. Invest Ophthalmol Vis Sci 2011;52:9516–22.

[102] Kaido M, Toda I, Ishida R, Konagai M, Dogru M, Tsubota K. Age-related changes in functional visual acuity in healthy individuals. Jpn J Ophthalmol 2011;55:183–9.

[103] Hara S, Kojima T, Ishida R, Goto E, Matsumoto Y, Kaido M, et al. Evaluation of tear stability after surgery for conjunctivochalasis. Optom Vis Sci 2011;88:1112–8.

[104] Kaido M, Matsutani T, Negishi K, Dogru M, Tsubota K. Aged drivers may experience decreased visual function while driving. Asia Pac J Ophthalmol (Phila) 2013;2:150–8.

[105] Ibrahim OM, Dogru M, Kaido M, Kojima T, Fujishima H, Tsubota K. Functional visual acuity assessment of severe atopic keratoconjunctivitis. Cornea 2014;33(Suppl 11):S13–8.

[106] Kaido M, Kawashima M, Yokoi N, Fukui M, Ichihashi Y, Kato H, et al. Advanced dry eye screening for visual display terminal workers using functional visual acuity measurement: the Moriguchi study. Br J Ophthalmol 2015;99(11):1488–92. http://dx.doi.org/10.1136/bjophthalmol-2015-306640.

[107] Tutt R, Bradley A, Begley C, Thibos LN. Optical and visual impact of tear break-up in human eyes. Invest Ophthalmol Vis Sci 2000;41:4117–23.

[108] Liu H, Thibos L, Begley CG, Bradley A. Measurement of the time course of optical quality and visual deterioration during tear break-up. Invest Ophthalmol Vis Sci 2010;51:3318–26.

[109] Montes-Mico R, Caliz A, Alio JL. Wavefront analysis of higher order aberrations in dry eye patients. J Refract Surg 2004;20:243–7.

[110] Koh S, Maeda N, Hirohara Y, Mihashi T, Bessho K, Hori Y, et al. Serial measurements of higher-order aberrations after blinking in patients with dry eye. Invest Ophthalmol Vis Sci 2008;49:133–8.

[111] Denoyer A, Rabut G, Baudouin C. Tear film aberration dynamics and vision-related quality of life in patients with dry eye disease. Ophthalmology 2012;119:1811–8.

[112] Habay T, Majzoub S, Perrault O, Rousseau C, Pisella PJ. Objective assessment of the functional impact of dry eye severity on the quality of vision by double-pass aberrometry. J Fr Ophtalmol 2014;37:188–94.

[113] Koh S, Maeda N, Ikeda C, Asonuma S, Mitamura H, Oie Y, et al. Ocular forward light scattering and corneal backward light scattering in patients with dry eye. Invest Ophthalmol Vis Sci 2014;55:6601–6.

[114] Sweeney DFM, T J, Raju SR. Tear film stability: a review. Exp eye Res 2013;117:28–38.

[115] Lemp MA, Holly FJ, Iwata S, Dohlman CH. The precorneal tear film. I. Factors in spreading and maintaining a continuous tear film over the corneal surface. Arch Ophthalmol 1970;83:89–94.

[116] Norn M. Desiccation of the precorneal tear film I. Corneal wetting time. Acta Ophthalmol 1969;47:865–80.

[117] Mengher LSB, A J, Tonge SR, Gilbert DJ. Effect of fluorescein instillation on the pre-corneal tear film stability. Curr Eye Res 1985;4:9–12.

[118] Mooi JK, Wang MT, Lim J, Muller A, Craig JP. Minimising instilled volume reduces the impact of fluorescein on clinical measurements of tear film stability. Cont Lens Anterior Eye 2017.

[119] Pult H, Riede-Pult BH. A new modified fluorescein strip: its repeatability and usefulness in tear film break-up time analysis. Cont Lens Anterior Eye 2012;35:35–8.

[120] Kim KT, Kim JH, Kong YT, Chae JB, Hyung S. Reliability of a new modified tear breakup time method: dry tear breakup time. Graefes Arch Clin Exp Ophthalmol 2015;253:1355–61.

[121] Korb DR, Greiner JV, Herman J. Comparison of fluorescein break-up time measurement reproducibility using standard fluorescein strips versus the Dry Eye Test (DET) method. Cornea 2001;20:811–5.

[122] Lemp MA, Hamill JR. Factors affecting tear film breakup in normal eyes. Arch Ophthalmol 1973;89:103–5.

[123] Abelson MBO, G 3rd W, Nally LA, Welch D, Krenzer K. Alternative reference values for tear film break up time in normal and dry eye populations. Adv Exp Med Biol 2002;506:1121–5.

[124] Abelson RL, K J, Rodriguez J, Johnston P, Angjeli E, Ousler G, et al. A single-center study evaluating the effect of the controlled adverse environment (CAE(SM)) model on tear film stability. Clin Ophthalmol 2012;6:1865–72.

[125] Sullivan BD, Crews LA, Sonmez B, Paz MF, Comert E, Charoenrook V. Clinical utility of objective tests for dry eye disease: variability over time and implications for clinical trials and disease management. Cornea 2012:31.

[126] Cebreiro E, Ramos L, Mosquera A, Barreira N, Penedo MFG. Automation of the tear film break-up time test. In: 4th international symposium on applied sciences in biomedical and communication technologies. Barcelona: ISABEL'11; 2011.

[127] Ramos L, Barreira N, Mosquera A, Penedo MG, Yebra-Pimentel E, García-Resúa C. Analysis of parameters for the automatic computation of the tear film break-up time test based on CCLRU standards. Comput Methods Programs Biomed 2014;113:715–24.

[128] Cardona G, Seres C, Quevedo L, Auge M. Knowledge and use of tear film evaluation tests by Spanish practitioners. Optom Vis Sci 2011;88:1106–11.

[129] Downie LE, Keller PR, Vingrys AJ. An evidence-based analysis of Australian optometrists' dry eye practices. Optom Vis Sci 2013;90:1385–95.

[130] Turner AW, Layton CJ, Bron AJ. Survey of eye practitioners' attitudes towards diagnostic tests and therapies for dry eye disease. Clin Exp Ophthalmol 2005;33:351–5.

[131] Graham JE, McGilligan VE, Berrar D, Leccisotti A, Moore JE, Bron AJ, et al. Attitudes towards diagnostic tests and therapies for dry eye disease. Ophthalmic Res 2010;43:11–7.

[132] Smith J, Nichols KK, Baldwin EK. Current patterns in the use of diagnostic tests in dry eye evaluation. Cornea 2008;27:656–62.

[133] Wang MT, Murphy PJ, Blades KJ, Craig JP. Comparison of non-invasive tear film stability measurement techniques. Clin Exp Optom 2017 (in press).

[134] Mengher LS, Bron AJ, Tonge SR, Gilbert DJ. A non-invasive instrument for clinical assessment of the pre-corneal tear film stability. Curr Eye Res 1985;4:1–7.

[135] Hirji N, Patel S, Callander M. Human tear film pre-rupture phase time (TP-RPT) - a non-invasive technique for evaluating the pre-corneal tear film using a novel keratometer mire. Ophthal Physiol Opt 1989;9:139–42.

[136] Liu Z, Pflugfelder SC. Corneal surface regularity and the effect of artificial tears in aqueous tear deficiency. Ophthalmology 1999;106:939–43.

[137] Goto T, Zheng X, Okamoto S, Ohashi Y. Tear film stability analysis system: introducing a new application for videokeratography. Cornea 2004;23:S65–70.

[138] Goto T, Zheng X, Klyce SD, Kataoka H, Uno T, Karon M, et al. A new method for tear film stability analysis using videokeratography. Am J Ophthalmol 2003;135:607–12.

[139] Kojima T, Ishida R, Dogru M, Goto E, Takano Y, Matsumoto Y. A new noninvasive tear stability analysis system for the assessment of dry eyes. Invest Ophthalmol Vis Sci 2004:45.

[140] Gumus K, Crockett CH, Rao K, Yeu E, Weikert MP, Shirayama M, et al. Noninvasive assessment of tear stability with the tear stability analysis system in tear dysfunction patients. Invest Ophthalmol Vis Sci 2011:52:

456—61.

[141] Best NDL, Wolffsohn JS. Clinical evaluation of the Oculus keratograph. Cont Lens Anterior Eye 2012;35:171—4.

[142] Hong J, Sun X, Wei A, Cui X, Li Y, Qian T, et al. Assessment of tear film stability in dry eye with a newly developed keratograph. Cornea 2013;32: 716—21.

[143] Abdelfattah NS, Dastiridou A, Sadda SR, Lee OL. Noninvasive imaging of tear film dynamics in eyes with ocular surface disease. Cornea 2015;34(Suppl 10):S48—52.

[144] Alonso-Caneiro D, Iskander DR, Collins MJ. Tear film surface quality with soft contact lenses using dynamic-area high-speed videokeratoscopy. Eye Contact Lens 2009;35:227—31.

[145] Iskander DR, Collins MJ. Applications of high-speed videokeratoscopy. Clin Exp Optom 2005;88:223—31.

[146] Kopf M, Yi F, Iskander DR, Collins MJ, Shaw AJ, Straker B. Tear film surface quality with soft contact lenses using dynamic videokeratoscopy. J Optom 2008;1:14—21.

[147] Downie LE. Automated tear film surface quality breakup time as a novel clinical marker for tear hyperosmolarity in dry eye disease. Invest Ophthalmol Vis Sci 2015;56:7260—8.

[148] Craig JP, Tomlinson A. Importance of the lipid layer in human tear film stability and evaporation. Optom Vis Sci 1997;74:8—13.

[149] Doane MG. An instrument for in vivo tear film interferometry. Optom Vis Sci 1989;66:383—8.

[150] Guillon JP. Use of the Tearscope Plus and attachments in the routine examination of the marginal dry eye contact lens patient. Adv Exp Med Biol 1998;438:859—67.

[151] Maissa C, Guillon M. Tear film dynamics and lipid layer characteristics—effect of age and gender. Cont Lens Anterior Eye 2010;33:176—82.

[152] Yokoi N, Komuro A. Non-invasive methods of assessing the tear film. Exp Eye Res 2004;78:399—407.

[153] Finis D, Pischel N, Schrader S, Geerling G. Evaluation of lipid layer thickness measurement of the tear film as a diagnostic tool for Meibomian gland dysfunction. Cornea 2013;32:1549—53.

[154] Guillon M, Styles E, Guillon JP, Maissa C. Preocular tear film characteristics of nonwearers and soft contact lens wearers. Optom Vis Sci 1997;74:273—9.

[155] Cho P, Douthwaite W. The relation between invasive and noninvasive tear break-up time. Optom Vis Sci 1995;72:17—22.

[156] Craig JP, Singh I, Tomlinson A, Morgan PB. The role of tear physiology in ocular surface temperature. Eye (Lond) 2000;14(Pt 4):635—41.

[157] Tan JH, Ng EYK, Rajendra Acharya U, Chee C. Infrared thermography on ocular surface temperature: a review. Infrared Phys Technol 2009;52: 97—108.

[158] Purslow C, Wolffsohn J. The relation between physical properties of the anterior eye and ocular surface temperature. Optom Vis Sci 2007;84: 197—201.

[159] Fujishima H, Toda I, Yamada M, Sato N, Tsubota K. Corneal temperature in patients with dry eye evaluated by infrared radiation thermometry. Br J Ophthalmol 1996;80:29—32.

[160] Kamao T, Yamaguchi M, Kawasaki S, Mizoue S, Shiraishi A, Ohashi Y. Screening for dry eye with newly developed ocular surface thermographer. Am J Ophthalmol 2011;151:782—91.

[161] Su TY, Hwa CK, Liu PH, Wu MH, Chang DO, Su PF, et al. Noncontact detection of dry eye using a custom designed infrared thermal image system. J Biomed Opt 2011;16:046009.

[162] Klamann MKJ, Maier AKB, Gonnermann J, Klein JP, Pleyer U. Measurement of dynamic ocular surface temperature in healthy subjects using a new thermography device. Curr Eye Res 2012;37:678—83.

[163] Szczesna DH, Alonso-Caneiro D, Iskander DR, Read SA, Collins MJ. Predicting dry eye using noninvasive techniques of tear film surface assessment. Invest Ophthalmol Vis Sci 2011;52:751—6.

[164] Abreu K, Callan C, Kottaiyan R, Zhang A, Yoon G, Aquavella JV, et al. Temperatures of the ocular surface, lid, and periorbital regions of Sjogren's, evaporative, and aqueous-deficient dry eyes relative to normals. Ocul Surf 2016;14:64—73.

[165] Li W, Graham AD, Selvin S, Lin MC. Ocular surface cooling corresponds to tear film thinning and breakup. Optom Vis Sci 2015;92. e248—e56.

[166] Su TY, Chang SW, Yang CJ, Chiang HK, Ho WT, Chang SW, Chiang HK. Thermographic evaluation of tear film break-up time to study tear film stability. Int J Therm Sci 2014;99:36—40.

[168] Versura P, Giannaccare G, Fresina M, Campos EC. Subjective discomfort symptoms are related to low corneal temperature in patients with evaporative dry eye. Cornea 2015;34:1079—85.

[169] Gilbard JP, Farris RL, Santamaria II J. Osmolarity of tear microvolumes in keratoconjunctivitis sicca. Arch Ophthalmol 1978;96:677—81.

[170] Keech A, Senchyna M, Jones L. Impact of time between collection and collection method on human tear fluid osmolarity. Curr Eye Res 2013;38: 428—36.

[171] Sullivan B. Challenges in using signs and symptoms to evaluate new biomarkers of dry eye disease. Ocul Surf 2014;12:2—9.

[172] Liu H, Begley C, Chen M, Bradley A, Bonanno J, McNamara NA, et al. A link between tear instability and hyperosmolarity in dry eye. Invest Ophthalmol Vis Sci 2009;50:3671—9.

[173] Peng CC, Cerretani C, Braun RJ, Radke CJ. Evaporation-driven instability of the precorneal tear film. Adv Colloid Interface Sci 2014;206:250—64.

[174] Willcox MDP, Argüeso P, Georgiev G, Holopainen J, Laurie G, Millar T, et al. TFOS DEWS II tear film report. Ocul Surf 2017;15(3):369—406.

[175] Abusharha AA, Pearce EI. The effect of low humidity on the human tear film. Cornea 2013;32:429—34.

[176] Hamano H, Hori M, Mitsunaga S. Measurement of evaporation rate of water from the precorneal tear film and contact lenses. Contacto 1981;25:7—14.

[177] Trees GR, Tomlinson A. Effect of artificial tear solutions and saline on tear film evaporation. Optom Vis Sci 1990;67:886—90.

[178] Rolando M, Refojo MF. Tear evaporimeter for measuring water evaporation rate from the tear film under controlled conditions in humans. Exp Eye Res 1983;36:25—33.

[179] Rolando M, Refojo MF, Kenyon KR. Increased tear evaporation in eyes with keratoconjunctivitis sicca. Arch Ophthalmol 1983;101:557—8.

[180] Tsubota K, Yamada M. Tear evaporation from the ocular surface. Invest Ophthalmol Vis Sci 1992;33:2942—50.

[181] Tomlinson A, Cedarstaff TH. Tear evaporation from the human eye: the effects of contact lens wear. J Br Contact Lens Assoc 1982;5:141—4. 6-7.

[182] Mathers WD, Binarao G, Petroll M. Ocular water evaporation and the dry eye. A new measuring device. Cornea 1993;12:335—40.

[183] Mathers WD, Daley TE. Tear flow and evaporation in patients with and without dry eye. Ophthalmology 1996;103:664—9.

[184] Guillon M, Maissa C. Contact lens wear affects tear film evaporation. Eye Contact Lens 2008;34:326—30.

[185] Craig JP, Willcox MD, Argueso P, Maissa C, Stahl U, Tomlinson A, et al. The TFOS International Workshop on Contact Lens Discomfort: report of the contact lens interactions with the tear film subcommittee. Invest Ophthalmol Vis Sci 2013;54. TFOS123—56.

[186] Abusharha AA, Pearce EI, Fagehi R. Effect of ambient temperature on the human tear film. Eye Contact Lens 2015.

[187] Uchiyama E, Aronowicz JD, Butovich IA, McCulley JP. Increased evaporative rates in laboratory testing conditions simulating airplane cabin relative humidity: an important factor for dry eye syndrome. Eye Contact Lens 2007;33: 174—6.

[188] Wojtowicz JC, McCulley JP. Assessment and impact of the time of day on aqueous tear evaporation in normal subjects. Eye Contact Lens 2009;35: 117—9.

[189] Petznick A, Tan JH, Boo SK, Lee SY, Acharya UR, Tong L. Repeatability of a new method for measuring tear evaporation rates. Optom Vis Sci 2013;90: 366—71.

[190] Rohit A, Ehrmann K, Naduvilath T, Willcox M, Stapleton F. Validating a new device for measuring tear evaporation rates. Ophthalmic Physiol Opt 2014;34:53—62.

[191] Tan JH, Ng EYK, Acharya UR. Evaluation of tear evaporation from ocular surface by functional infrared thermography. Med Phys 2010;37:6022—34.

[192] Purslow C, Wolffsohn JS, Santodomingo-Rubido J. The effect of contact lens wear on dynamic ocular surface temperature. Cont Lens Anterior Eye 2005;28:29—36.

[193] Bron AJ, Abelson MB, Ousler G, Pearce E, Tomlinson A, Yokoi N, et al. Methodologies to diagnose and monitor dry eye disease: report of the diagnostic methodology subcommittee of the international Dry Eye Workshop. Ocul Surf 2007;2007(5):108—52.

[194] Best N, Drury L, Wolffsohn JS. Predicting success with silicone-hydrogel contact lenses in new wearers. Cont Lens Anterior Eye 2013;36:232—7.

[195] Elliott M, Fandrich H, Simpson T, Fonn D. Analysis of the repeatability of tear break-up time measurement techniques on asymptomatic subjects before, during and after contact lens wear. Cont Lens Anterior Eye 1998;21:98—103.

[196] Wang J, Aquavella J, Palakuru J, Chung S, Feng C. Relationships between central tear film thickness and tear menisci of the upper and lower eyelids. Invest Ophthalmol Vis Sci 2006;47:4349—55.

[197] Holly FJ. Physical chemistry of the normal and disordered tear film. Trans Ophthalmol Soc U K 1985;104(Pt 4):374—80.

[198] Mainstone JC, Bruce AS, Golding TR. Tear meniscus measurement in the diagnosis of dry eye. Curr Eye Res 1996;15:653—61.

[199] Golding TR, Bruce AS, Mainstone JC. Relationship between tear-meniscus parameters and tear-film breakup. Cornea 1997;16:649—61.

[200] Oguz H, Yokoi N, Kinoshita S. The height and radius of the tear meniscus and methods for examining these parameters. Cornea 2000;19:497—500.

[201] Yokoi N, Bron A, Tiffany J, Brown N, Hsuan J, Fowler C. Reflective meniscometry: a non-invasive method to measure tear meniscus curvature. Br J Ophthalmol 1999;83:92—7.

[202] Yokoi N, Bron AJ, Tiffany JM, Kinoshita S. Reflective meniscometry: a new field of dry eye assessment. Cornea 2000;19:S37—43.

[203] Bandlitz S, Purslow C, Murphy PJ, Pult H. The relationship between tear meniscus regularity and conjunctival folds. Optom Vis Sci 2014;91:1037—44.

[204] Bandlitz S, Purslow C, Murphy PJ, Pult H, Bron AJ. A new portable digital meniscometer. Optom Vis Sci 2014;91:e1—8.

[205] Bandlitz S, Purslow C, Murphy PJ, Pult H. Comparison of a new portable digital meniscometer and optical coherence tomography in tear meniscus radius measurement. Acta Ophthalmol 2014;92. e112—8.

[206] Bandlitz S, Purslow C, Murphy PJ, Pult H. Time course of changes in tear meniscus radius and blink rate after instillation of artificial tears. Invest Ophthalmol Vis Sci 2014;55:5842—7.

[207] Akiyama R, Usui T, Yamagami S. Diagnosis of dry eye by tear meniscus measurements using anterior segment swept source optical coherence tomography. Cornea 2015;34(Suppl 11):S115—20.

[208] Altan-Yaycioglu R, Sizmaz S, Canan H, Coban-Karatas M. Optical coherence tomography for measuring the tear film meniscus: correlation with schirmer test and tear-film breakup time. Curr Eye Res 2013;38:736—42.

[209] Arriola-Villalobos P, Fernandez-Vigo JI, Diaz-Valle D, Peraza-Nieves JE, Fernandez-Perez C, Benitez-Del-Castillo JM. Assessment of lower tear meniscus measurements obtained with Keratograph and agreement with Fourier-domain optical-coherence tomography. Br J Ophthalmol 2015;99:1120—5.

[210] Baek J, Doh SH, Chung SK. Comparison of tear meniscus height measurements obtained with the keratograph and fourier domain optical coherence tomography in dry eye. Cornea 2015;34:1209—13.

[211] Bartuzel MM, Szczesna-Iskander DH, Iskander DR. Automatic dynamic tear meniscus measurement in optical coherence tomography. Biomed Opt Express 2014;5:2759—68.

[212] Canan H, Altan-Yaycioglu R, Ulas B, Sizmaz S, Coban-Karatas M. Interexaminer reproducibility of optical coherence tomography for measuring the tear film meniscus. Curr Eye Res 2014;39:1145—50.

[213] Chan HH, Zhao Y, Tun TA, Tong L. Repeatability of tear meniscus evaluation using spectral-domain Cirrus(R) HD-OCT and time-domain Visante(R) OCT. Cont Lens Anterior Eye 2015;38:368—72.

[214] Chen Q, Zhang X, Cui L, Huang Q, Chen W, Ma H, et al. Upper and lower tear menisci in Sjogren's syndrome dry eye. Invest Ophthalmol Vis Sci 2011;52: 9373—8.

[215] Cui L, Shen M, Wang J, Jiang J, Li M, Chen D, et al. Age-related changes in tear menisci imaged by optical coherence tomography. Optom Vis Sci 2011;88: 1214—9.

[216] Czajkowski G, Kaluzny BJ, Laudencka A, Malukiewicz G, Kaluzny JJ. Tear meniscus measurement by spectral optical coherence tomography. Optom Vis Sci 2012;89:336—42.

[217] Huang ZP, Meng H. Application of tear meniscus measurement by anterior segment optical coherence tomography in the diagnosis of dry eye. Eye Sci 2012;27:217—9.

[218] Ibrahim OM, Dogru M, Takano Y, Satake Y, Wakamatsu TH, Fukagawa K, et al. Application of visante optical coherence tomography tear meniscus height measurement in the diagnosis of dry eye disease. Ophthalmology 2010;117: 1923—9.

[219] Qiu X, Gong L, Lu Y, Jin H, Robitaille M. The diagnostic significance of Fourier-domain optical coherence tomography in Sjogren syndrome, aqueous tear deficiency and lipid tear deficiency patients. Acta Ophthalmol 2012;90. e359—66.

[220] Qiu X, Gong L, Sun X, Jin H. Age-related variations of human tear meniscus and diagnosis of dry eye with Fourier-domain anterior segment optical coherence tomography. Cornea 2011;30543—9.

[221] Shen M, Li J, Wang J, Ma H, Cai C, Tao A, et al. Upper and lower tear menisci in the diagnosis of dry eye. Invest Ophthalmol Vis Sci 2009;50:2722—6.

[222] Shen M, Wang J, Tao A, Chen Q, Lin S, Qu J, et al. Diurnal variation of upper and lower tear menisci. Am J Ophthalmol 2008;145:801—6.

[223] Su TY, Ho WT, Lu CY, Chang SW, Chiang HK. Correlations among ocular surface temperature difference value, the tear meniscus height, Schirmer's test and fluorescein tear film break up time. Br J Ophthalmol 2015;99:482—7.

[224] Tittler EH, Bujak MC, Nguyen P, Zhang X, Li Y, Yiu SC, et al. Between-grader repeatability of tear meniscus measurements using Fourier-domain OCT in patients with dry eye. Ophthalmic Surg Lasers Imaging 2011;42:423—7.

[225] Wang CX, Liu YZ, Yuan J, Li BB, Zhou SY. Application of anterior segment optical coherence tomography for measuring the tear meniscus height in the diagnosis of dry eye diseases. Zhonghua Yan Ke Za Zhi 2009;45:616—20.

[226] Zhou S, Li Y, Lu AT, Liu P, Tang M, Yiu SC, et al. Reproducibility of tear meniscus measurement by Fourier-domain optical coherence tomography: a pilot study. Ophthalmic Surg Lasers Imaging 2009;40:442—7.

[227] Pult H, Riede-Pult BH. Impact of conjunctival folds on central tear meniscus height. Invest Ophthalmol Vis Sci 2015;56:1459—66.

[228] de Monchy I, Gendron G, Miceli C, Pogorzalek N, Mariette X, Labetoulle M. Combination of the Schirmer I and phenol red thread tests as a rescue strategy for diagnosis of ocular dryness associated with Sjogren's syndrome. Invest Ophthalmol Vis Sci 2011;52:5167—73.

[229] Cho P. The cotton thread test: a brief review and a clinical study of its reliability on Hong Kong-Chinese. Optom Vis Sci 1993;70:804—8.

[230] Tomlinson A, Blades KJ, Pearce EI. What does the phenol red thread test actually measure? Optom Vis Sci 2001;78:142—6.

[231] Miller WL, Doughty MJ, Narayanan S, Leach NE, Tran A, Gaume AL, et al. A comparison of tear volume (by tear meniscus height and phenol red thread test) and tear fluid osmolality measures in non-lens wearers and in contact lens wearers. Eye Contact Lens 2004;30:132—7.

[232] Sakamoto R, Bennett ES, Henry VA, Paragina S, Narumi T, Izumi Y, et al. The phenol red thread test: a cross-cultural study. Invest Ophthalmol Vis Sci 1993;34:3510—4.

[233] Vashisht S, Singh S. Evaluation of Phenol Red Thread test versus Schirmer test in dry eyes: a comparative study. Int J Appl Basic Med Res 2011;1:40—2.

[234] Patel S, Farrell J, Blades KJ, Grierson DJ. The value of a phenol red impregnated thread for differentiating between the aqueous and non-aqueous deficient dry eye. Ophthalmic Physiol Opt 1998;18:471—6.

[235] Pult H, Purslow C, Murphy PJ. The relationship between clinical signs and dry eye symptoms. Eye (Lond) 2011;25:502—10.

[236] Doughty MJ, Whyte J, Li W. The phenol red thread test for lacrimal volume—does it matter if the eyes are open or closed? Ophthalmic Physiol Opt 2007;27:482—9.

[237] Li N, Deng XG, He MF. Comparison of the Schirmer I test with and without topical anesthesia for diagnosing dry eye. Int J Ophthalmol 2012;5:478—81.

[238] Tsubota K, Kaido M, Yagi Y, Fujihara T, Shimmura S. Diseases associated with ocular surface abnormalities: the importance of reflex tearing. Br J Ophthalmol 1999;83:89—91.

[239] Serin D, Karsloglu S, Kyan A, Alagoz G. A simple approach to the repeatability of the Schirmer test without anesthesia: eyes open or closed? Cornea 2007;26:903—6.

[240] Bitton E, Wittich W. Influence of eye position on the Schirmer tear test. Cont Lens Anterior Eye 2014;37:257—61.

[241] Karampatakis V, Karamitsos A, Skriapa A, Pastiadis G. Comparison between normal values of 2- and 5-minute Schirmer test without anesthesia. Cornea 2010;29:497—501.

[242] van Bijsterveld OP. Diagnostic tests in the Sicca syndrome. Arch Ophthalmol 1969;82:10—4.

[243] Dogru M, Ishida K, Matsumoto Y, Goto E, Ishioka M, Kojima T, et al. Strip meniscometry: a new and simple method of tear meniscus evaluation. Invest Ophthalmol Vis Sci 2006;47:1895—901.

[244] Ibrahim OM, Dogru M, Ward SK, Matsumoto Y, Wakamatsu TH, Ishida K, et al. The efficacy, sensitivity, and specificity of strip meniscometry in conjunction with tear function tests in the assessment of tear meniscus. Invest Ophthalmol Vis Sci 2011;52:2194—8.

[245] Vitali C, Bombardieri S, Jonsson R, Moutsopoulos HM, Alexander EL, Carsons SE, et al. Classification criteria for Sjogren's syndrome: a revised version of the European criteria proposed by the American-European Consensus Group. Ann Rheum Dis 2002;61:554—8.

[246] Potvin R, Makari S, Rapuano CJ. Tear film osmolarity and dry eye disease: a review of the literature. Clin Ophthalmol 2015;9:2039—47.

[247] Jackson DC, Zeng W, Wong CY, Mifsud EJ, Williamson NA, Ang CS, et al. Tear interferon-gamma as a biomarker for evaporative dry eye disease. Invest Ophthalmol Vis Sci 2016;57:4824—30.

[248] Bunya VY, Fuerst NM, Pistilli M, McCabe BE, Salvo R, Macchi I, et al. Variability of tear osmolarity in patients with dry eye. JAMA Ophthalmol 2015;133:662—7.

[249] Versura P, Profazio V, Campos EC. Performance of tear osmolarity compared to previous diagnostic tests for dry eye diseases. Curr Eye Res 2010;35: 553—64.

[250] Schargus M, Meyer-ter-Vehn T, Menrath J, Grigoleit GU, Geerling G. Correlation between tear film osmolarity and the disease score of the international chronic ocular graft-versus-host-disease consensus group in hematopoietic stem cell transplantation patients. Cornea 2015;34:911—6.

[251] Khanal S, Tomlinson A, McFadyen A, CDiaper. Dry eye diagnosis. Invest Ophthalmol Vis Sci 2008;49:1407—14.

[252] Tomlinson A, McCann LC, Pearce EI. Comparison of human tear film osmolarity measured by electrical impedance and freezing point depression techniques. Cornea 2010;29:1036—41.

[253] Masmali AM, Purslow C, Murphy PJ. The tear ferning test: a simple clinical technique to evaluate the ocular tear film. Clin Exp Optom 2014;97: 399—406.

[254] Masmali AM, Al-Qhtani S, Al-Gasham TM, El-Hiti GA, Purslow C, Murphy PJ. Application of a new grading scale for tear ferning in non-dry eye and dry eye subjects. Cont Lens Anterior Eye 2015;38:39—43.

[255] Rolando M, Baldi F, Zingirian M. The effect of hyperosmolarity on tear mucus ferning. Fortschr Ophthalmol 1986;83:644—6.

[256] Vaikoussis E, Georgiou P, Nomicarios D. Tear mucus ferning in patients with Sjogren's syndrome. Documenta Ophthalmol Adv Ophthalmol 1994;87: 145—51.

[257] Norn M. Quantitative tear ferning. Clinical investigations. Acta Ophthalmol (Copenh) 1994;72:369—72.

[258] Maragou M, Vaikousis E, Ntre A, Koronis N, Georgiou P, Hatzidimitriou E, et al. Tear and saliva ferning tests in Sjogren's syndrome (SS). Clin Rheumatol 1996;15:125—32.

[259] Albach KA, Lauer M, Stolze HH. Diagnosis of keratoconjunctivitis sicca in rheumatoid arthritis. The value of various tests. Ophthalmologe 1994;91: 229—34.

[260] Puderbach S, Stolze HH. Tear ferning and other lacrimal tests in normal persons of different ages. Int Ophthalmol 1991;15:391—5.

[261] Ravazzoni L, Ghini C, Macri A, Rolando M. Forecasting of hydrophilic contact lens tolerance by means of tear ferning test. Graefes Arch Clin Exp Ophthalmol 1998;236:354—8.

[262] Evans KS, North RV, Purslow C. Tear ferning in contact lens wearers. Ophthalmic Physiol Opt 2009;29:199—204.

[263] Downie LE, Keller PR. A pragmatic approach to dry eye diagnosis: evidence into practice. Optom Vis Sci 2015;92:1189—97.

[264] Niimi J, Tan B, Chang J, Zhou Y, Ghanekar A, Wong M, et al. Diurnal pattern of tear osmolarity and its relationship to corneal thickness and deswelling. Cornea 2013;32:1305—10.

[265] Bron AJ, Tomlinson A, Foulks GN, Pepose JS, Baudouin C, Geerling G, et al. Rethinking dry eye disease: a perspective on clinical implications. Ocul Surf 2014;12:S1—31.

[266] Bron AJ, Argueso P, Irkec M, Bright FV. Clinical staining of the ocular surface: mechanisms and interpretations. Prog Retin eye Res 2015;44:36—61.

[267] Feenstra RP, Tseng SC. Comparison of fluorescein and rose bengal staining. Ophthalmology 1992;99:605—17.

[268] Glasgow BJ. Fluorescence lifetime imaging microscopy reveals quenching of

fluorescein within corneal epithelium. Exp Eye Res 2016;147:12—9.

[269] Khan-Lim D, Berry M. Still confused about rose bengal? Curr Eye Res 2004;29:311—7.

[270] Argueso P, Tisdale A, Spur-Michaud S, Sumiyoshi M, Gipson IK. Mucin characteristics of human corneal-limbal epithelial cells that exclude the rose bengal anionic dye. Invest Ophthalmol Vis Sci 2006;47:113—9.

[271] Manning FJ, Wehrly SR, Foulks GN. Patient tolerance and ocular surface staining characteristics of lissamine green versus rose bengal. Ophthalmology 1995;102:1953—7.

[272] Korb DR, Herman JP, Finnemore VM, Exford JM, Blackie CA. An evaluation of the efficacy of fluorescein, rose bengal, lissamine green, and a new dye mixture for ocular surface staining. Eye Contact Lens 2008;34:61—4.

[273] Kim J, Foulks GN. Evaluation of the effect of lissamine green and rose bengal on human corneal epithelial cells. Cornea 1999;18:328—32.

[274] Chodosh J, Dix RD, Howell RC, Stroop WG, Tseng SC. Staining characteristics and antiviral activity of sulforhodamine B and lissamine green B. Invest Ophthalmol Vis Sci 1994;35:1046—58.

[275] Versura P, Frigato M, Cellini M, Mule R, Malavolta N, Campos EC. Diagnostic performance of tear function tests in Sjogren's syndrome patients. Eye (Lond) 2007;21:229—37.

[276] Toda I, Tsubota K. Practical double vital staining for ocular surface evaluation. Cornea 1993;12:366—7.

[277] Yoon KC, Im SK, Kim HG, You IC. Usefulness of double vital staining with 1% fluorescein and 1% lissamine green in patients with dry eye syndrome. Cornea 2011;30:972—6.

[278] Korb DR, Herman JP, Solomon JD, Greiner JV, Blackie CA. Lid wiper staining and sequential fluorescein instillation. Invest Ophthalmol Vis Sci 2006;47. ARVO E-Abstract: 242.

[279] Peterson RC, Wolffsohn JS, Fowler CW. Optimization of anterior eye fluorescein viewing. Am J Ophthalmol 2006;142:572—5.

[280] Hamrah P, Alipour F, Jiang S, Sohn JH, Foulks GN. Optimizing evaluation of Lissamine Green parameters for ocular surface staining. Eye (Lond) 2011;25:1429—34.

[281] Lemp MA. Report of the National eye institute/industry workshop on clinical trials in dry eyes. CLAO J 1995;21:221—32.

[282] Barr JT, Schechtman KB, Fink BA, Pierce GE, Pensyl CD, Zadnik K, et al. Corneal scarring in the collaborative longitudinal evaluation of keratoconus (CLEK) study: baseline prevalence and repeatability of detection. Cornea 1999;18:34—46.

[283] Bron AJ, Evans VE, Smith JA. Grading of corneal and conjunctival staining in the context of other dry eye tests. Cornea 2003;22:640—50.

[284] Miyata K, Amano S, Sawa M, Nishida T. A novel grading method for superficial punctate keratopathy magnitude and its correlation with corneal epithelial permeability. Arch Ophthalmol 2003;121:1537—9.

[285] Whitcher JP, Shiboski CH, Shiboski SC, Heidenreich AM, Kitagawa K, Zhang S, et al. A simplified quantitative method for assessing keratoconjunctivitis sicca from the Sjogren's Syndrome International Registry. Am J Ophthalmol 2010;149:405—15.

[286] Rose-Nussbaumer J, Lietman TM, Shiboski CH, Shiboski SC, Bunya VY, Akpek EK, et al. Inter-grader agreement of the ocular staining score in the Sjogren's international clinical collaborative alliance (SICCA) registry. Am J Ophthalmol 2015;160:1150—3.

[287] Tole DM, McKelvie PA, Daniell M. Reliability of impression cytology for the diagnosis of ocular surface squamous neoplasia employing the Biopore membrane. Br J Ophthalmol 2001;85:154—8.

[288] Mrugacz M, Kasacka I, Bakunowicz-Lazarczyk A, Kaczmarski M, Kulak W. Impression cytology of the conjunctival epithelial cells in patients with cystic fibrosis. Eye (Lond) 2008;22:1137—40.

[289] Brignole F, Pisella PJ, De Saint Jean M, Goldschild M, Goguel A, Baudouin C. Flow cytometric analysis of inflammatory markers in KCS: 6-month treatment with topical cyclosporin A. Invest Ophthalmol Vis Sci 2001;42:90—5.

[290] Yoon KC, Heo H, Im SK, You IC, Kim YH, Park YG. Comparison of autologous serum and umbilical cord serum eye drops for dry eye syndrome. Am J Ophthalmol 2007;144:86—92.

[291] Nelson JD, Havener VR, Cameron JD. Cellulose acetate impressions of the ocular surface. Dry eye states. Arch Ophthalmol 1983;101:1869—72.

[292] Tseng SC. Staging of conjunctival squamous metaplasia by impression cytology. Ophthalmology 1985;92:728—33.

[293] Blades K, Doughty MJ, Patel S. Pilot study on the use of impression cytology specimens for quantitative assessment of the surface area of bulbar conjunctival cells. Optom Vis Sci 1998;75:591—9.

[294] Zuazo F, Lopez-Ponce D, Salinas-Toro D, Valenzuela F, Sans-Puroja J, Srur M, et al. Conjunctival impression cytology in patients with normal and impaired OSDI scores. Arch Soc Esp Oftalmol 2014;89:391—6.

[295] Pult H, Tosatti SGP, Spencer ND, Asfour J-M, Ebenhoch M, Murphy PJ. Spontaneous blinking from a tribological viewpoint. Ocul Surf 2015;13:236—49.

[296] Gumus K, Pflugfelder SC. Increasing prevalence and severity of conjunctivochalasis with aging detected by anterior segment optical coherence tomography. Am J Ophthalmol 2013;155. 238—42.e2.

[297] Höh H, Schirra F, Kienecker C, Ruprecht KW. Lid-parallel conjunctival folds (LIPCOF): a definite diagnostic sign of dry eye. Ophthalmologe 1995;92:802—8.

[298] Berry M, Pult H, Purslow C, Murphy PJ. Mucins and ocular signs in symptomatic and asymptomatic contact lens wear. Optom Vis Sci 2008;85:

E930—8.

[299] Pult H, Purslow C, Berry M, Murphy PJ. Clinical tests for successful contact lens wear: relationship and predictive potential. Optom Vis Sci 2008;85:E924—9.

[300] Meller D, Tseng SC. Conjunctivochalasis: literature review and possible pathophysiology. Surv Ophthalmol 1998;43:225—32.

[301] Pult H, Riede-Pult BH, Murphy PJ. The relation between blinking and conjunctival folds and dry eye symptoms. Optom Vis Sci 2013;90:1034—9.

[302] Pult H, Murphy PJ, Purslow C. A novel method to predict dry eye symptoms in new contact lens wearers. Optom Vis Sci 2009;86:E1042—50.

[303] Németh J, Fodor E, Lang Z, Kosina-Hagyó K, Berta A, Komár T, et al. Lid-parallel conjunctival folds (LIPCOF) and dry eye: a multicentre study. Br J Ophthalmol 2012.

[304] Sickenberger W, Pult H, Sickenberger B. LIPCOF and contact lens wearers - a new tool of forecast subjective dryness and degree of comfort of contact lens wearers. Contactologia 2000;22:74—9.

[305] Markoulli M, Carnt N, Jalbert I, Keay L, Naduvilath T, Papas E. Resolution and clinical characteristics of conjunctival "Flaps". Invest Ophthalmol Vis Sci 2007;48. ARVO E-Abstract: 5391.

[306] Thota S, Perrigin J, Miller W, Leach N, Bergmanson J, Back A. Conjunctival Flaps in silicone hydrogel lens wearers. Invest Ophthalmol Vis Sci 2006;47. ARVO E-Abstract: 82.

[307] Graham AD, Truong TN, Lin MC. Conjunctival epithelial flap in continuous contact lens wear. Optom Vis Sci 2009;86. e324—e31.

[308] Veres A, Tapaszto B, Kosina-Hagyo K, Somfai GM, Nemeth J. Imaging lid-parallel conjunctival folds with OCT and comparing its grading with the slit lamp classification in dry eye patients and normal subjects. Invest Ophthalmol Vis Sci 2011;52:2945—51.

[309] Tapaszto B, Veres A, Kosina-Hagyo K, Somfai GM, Nemeth J. OCT imaging of lid-parallel conjunctival folds in soft contact lens wearers. Optom Vis Sci 2011.

[310] Lopez Garcia JS, Garcia Lozano I, Martinez Garchitorena J. Lacunar folds study in dry eye diagnosis. Arch Soc Esp Oftalmol 2003;78:21—7.

[311] Viso E, Rodriguez-Ares MT, Boveda FJ, Tourino R, Gude F. Prevalence of conjunctival shrinkage and its association with dry eye disease: results from a population-based study in Spain. Cornea 2014;33:442—7.

[312] Alhatem A, Cavalcanti B, Hamrah P. In vivo confocal microscopy in dry eye disease and related conditions. Semin Ophthalmol 2012;27:138—48.

[313] Villani E, Mantelli F, Nucci P. In-vivo confocal microscopy of the ocular surface: ocular allergy and dry eye. Curr Opin Allergy Clin Immunol 2013;13:569—76.

[314] Erdelyi B, Kraak R, Zhivov A, Guthoff R, Nemeth J. In vivo confocal laser scanning microscopy of the cornea in dry eye. Graefes Arch Clin Exp Ophthalmol 2007;245:39—44.

[315] Villani E, Magnani F, Viola F, Santaniello A, Scorza R, Nucci P, et al. In vivo confocal evaluation of the ocular surface morpho-functional unit in dry eye. Optom Vis Sci 2013;90:576—86.

[316] Wakamatsu TH, Sato EA, Matsumoto Y, Ibrahim OM, Dogru M, Kaido M, et al. Conjunctival in vivo confocal scanning laser microscopy in patients with Sjogren syndrome. Invest Ophthalmol Vis Sci 2010;51:144—50.

[317] Villani E, Galimberti D, Viola F, Mapelli C, Ratiglia R. The cornea in Sjogren's syndrome: an in vivo confocal study. Invest Ophthalmol Vis Sci 2007;48:2017—22.

[318] Kojima T, Matsumoto Y, Dogru M, Tsubota K. The application of in vivo laser scanning confocal microscopy as a tool of conjunctival in vivo cytology in the diagnosis of dry eye ocular surface disease. Mol Vis 2010;16:2457—64.

[319] Benitez del Castillo JM, Wasfy MA, Fernandez C, Garcia-Sanchez J. An in vivo confocal masked study on corneal epithelium and subbasal nerves in patients with dry eye. Invest Ophthalmol Vis Sci 2004;45:3030—5.

[320] Benitez-Del-Castillo JM, Acosta MC, Wassfi MA, Diaz-Valle D, Gegundez JA, Fernandez C, et al. Relation between corneal innervation with confocal microscopy and corneal sensitivity with noncontact esthesiometry in patients with dry eye. Invest Ophthalmol Vis Sci 2007;48:173—81.

[321] Villani E, Galimberti D, Viola F, Mapelli C, Del Papa N, Ratiglia R. Corneal involvement in rheumatoid arthritis: an in vivo confocal study. Invest Ophthalmol Vis Sci 2008;49:560—4.

[322] Tuisku IS, Konttinen YT, Konttinen LM, Tervo TM. Alterations in corneal sensitivity and nerve morphology in patients with primary Sjogren's syndrome. Exp Eye Res 2008;86:879—85.

[323] Zhang M, Chen J, Luo L, Xiao Q, Sun M, Liu Z. Altered corneal nerves in aqueous tear deficiency viewed by in vivo confocal microscopy. Cornea 2005;24:818—24.

[324] Labbe A, Alalwani H, Van Went C, Brasnu E, Georgescu D, Baudouin C. The relationship between subbasal nerve morphology and corneal sensation in ocular surface disease. Invest Ophthalmol Vis Sci 2012;53:4926—31.

[325] Labbe A, Liang Q, ZWang, Zhang Y, Xu L, Baudouin C, et al. Corneal nerve structure and function in patients with non-sjogren dry eye: clinical correlations. Invest Ophthalmol Vis Sci 2013;54:5144—50.

[326] Villani E, Beretta S, Galimberti D, Viola F, Ratiglia R. In vivo confocal microscopy of conjunctival roundish bright objects: young, older, and Sjogren subjects. Invest Ophthalmol Vis Sci 2011;52:4829—32.

[327] Hong J, Zhu W, Zhuang H, Xu J, Sun X, Le Q, et al. In vivo confocal microscopy of conjunctival goblet cells in patients with Sjogren's syndrome dry eye. Br J Ophthalmol 2010;94:1454—8.

[328] Le QH, Wang WT, Hong JX, Sun XH, Zheng TY, Zhu WQ, et al. An in vivo

confocal microscopy and impression cytology analysis of goblet cells in patients with chemical burns. Invest Ophthalmol Vis Sci 2010;51:1397–400.

[329] Messmer EM, Torres Suarez E, Mackert MI, Zapp DM, Kampik A. In vivo confocal microscopy in blepharitis. Klin Monbl Augenheilkd 2005;222:894–900.

[330] Hong J, Zhu W, Zhuang H, Xu J, Sun X, Le Q, et al. In vivo confocal microscopy of conjunctival goblet cells in patients with Sjogren syndrome dry eye. Br J Ophthalmol 2009;94:1454–8.

[331] Colorado LH, Alzahrani Y, Pritchard N, Efron N. Assessment of conjunctival goblet cell density using laser scanning confocal microscopy versus impression cytology. Cont Lens Anterior Eye 2016;39:221–6.

[332] Nishida T. Neurotrophic mediators and corneal wound healing. Ocul Surf 2005;3:194–202.

[333] Belmonte C, Nichols JJ, Begley C, Bereiter D, Brock JA, Cox S, et al. TFOS DEWS II pain and sensation report. Ocul Surf 2017;15(3):407–40.

[334] Cox SM, Nichols JJ. Association between meibomian gland testing and ocular surface sensitivity. Cornea 2015;34:1187–92.

[335] Wei Y, Asbell PAThe core mechanism of dry eye disease is inflammation. Eye Contact Lens 2014;40:248–56.

[336] Pflugfelder SC, Huang AJ, Feuer W, Chuchovski PT, Pereira IC, Tseng SC. Conjunctival cytologic features of primary Sjogren's syndrome. Ophthalmology 1990;97:985–91.

[337] Raphael M, Bellefqih S, Piette JC, Le Hoang P, Debre P, Chomette G. Conjunctival biopsy in Sjogren's syndrome: correlations between histological and immunohistochemical features. Histopathology 1988;13:191–202.

[338] Papas EB. Key factors in the subjective and objective assessment of conjunctival erythema. Invest Ophthalmol Vis Sci 2000;41:687–91.

[339] Fieguth P, Simpson T. Automated measurement of bulbar redness. Invest Ophthalmol Vis Sci 2002;43:340–7.

[340] Amparo F, Wang H, Emami-Naeini P, Karimian P, Dana R. The Ocular Redness Index: a novel automated method for measuring ocular injection. Invest Ophthalmol Vis Sci 2013;54:4821–6.

[341] Peterson RC, Wolffsohn JS. Objective grading of the anterior eye. Optom Vis Sci 2009;86:273–8.

[342] Peterson RC, Wolffsohn JS. Sensitivity and reliability of objective image analysis compared to subjective grading of bulbar hyperaemia. Br J Ophthalmol 2007;91:1464–6.

[343] Wolffsohn JS. Incremental nature of anterior eye grading scales determined by objective image analysis. Br J Ophthalmol 2004;88:1434–8.

[344] Wolffsohn JS, Purslow C. Clinical monitoring of ocular physiology using digital image analysis. Cont Lens Anterior Eye 2003;26:27–35.

[345] Acera A, Rocha G, Vecino E, Lema I, Duran JA. Inflammatory markers in the tears of patients with ocular surface disease. Ophthalmic Res 2008;40:315–21.

[346] Chotikavanich S, de Paiva CS, Li de Q, Chen JJ, Bian F, Farley WJ, et al. Production and activity of matrix metalloproteinase-9 on the ocular surface increase in dysfunctional tear syndrome. Invest Ophthalmol Vis Sci 2009;50:3203–9.

[347] Hadassah J, Bhuvaneshwari N, Rao U, Sehgal PK. Evaluation of succinylated collagen bandage lenses in corneal healing by the expression of matrix metalloproteinases (MMP-2 and MMP-9) in tear fluid. Ophthalmic Res 2009;42:64–72.

[348] Kaufman HE. The practical detection of mmp-9 diagnoses ocular surface disease and may help prevent its complications. Cornea 2013;32:211–6.

[349] Tong L, Thumboo J, Tan YK, Wong TY, Albani S. The eye: a window of opportunity in rheumatoid arthritis? Nat Rev Rheumatol 2014;10:552–60.

[350] Meadows JF, Dionne K, Nichols KK. Differential profiling of t-cell cytokines as measured by protein microarray across dry eye subgroups. Cornea 2016;35:329–35.

[351] Quah JH, Tong L, Barbier S. Patient acceptability of tear collection in the primary healthcare setting. Optom Vis Sci 2014;91:452–8.

[352] Wei Y, Gadaria-Rathod N, Epstein S, Asbell P. Tear cytokine profile as a noninvasive biomarker of inflammation for ocular surface diseases: standard operating procedures. Invest Ophthalmol Vis Sci 2013;54:8327–36.

[353] Le Guezennec X, Quah J, Tong L, Kim N. Human tear analysis with miniaturized multiplex cytokine assay on "wall-less" 96-well plate. Mol Vis 2015;21:1151–61.

[354] Benito MJ, Gonzalez-Garcia MJ, Teson M, Garcia N, Fernandez I, Calonge M, et al. Intra- and inter-day variation of cytokines and chemokines in tears of healthy subjects. Exp Eye Res 2014;120:43–9.

[355] Malesinski R, Bakunowicz-Lazarczyk A, Wysocka J. The role of chemokines CCL3/MIP-1 alfa and CCL4/MIP-1 beta in pathogenesis of dry eye syndrome. Klin Ocz 2008;110:277–9.

[356] Mrugacz M. CCL4/MIP-1beta levels in tear fluid and serum of patients with cystic fibrosis. J Interferon Cytokine Res 2010;30:509–12.

[357] Yoon KC, Park CS, You IC, Choi HJ, Lee KH, Im SK, et al. Expression of CXCL9, -10, -11, and CXCR3 in the tear film and ocular surface of patients with dry eye syndrome. Invest Ophthalmol Vis Sci 2010;51:643–50.

[358] Epstein SP, Gadaria-Rathod N, Wei Y, Maguire MG, Asbell PA. HLA-DR expression as a biomarker of inflammation for multicenter clinical trials of ocular surface disease. Exp Eye Res 2013;111:95–104.

[359] Baudouin C, Liang H, Riancho L, Ismail D, Deniaud M, Amrane M, et al. Correlation between the inflammatory marker HLA DR and signs and symptoms in moderate to severe dry eye disease. Invest Ophthalmol Vis Sci 2015;56. 298–CO183.

[360] Williams GP, Tomlins PJ, Denniston AK, Southworth HS, Sreekantham S, Curnow SJ, et al. Elevation of conjunctival epithelial CD45INTCD11b(+)CD16(+)CD14(-) neutrophils in ocular Stevens-Johnson syndrome and toxic epidermal necrolysis. Invest Ophthalmol Vis Sci 2013;54:4578–85.

[361] Sanchez MA, Arriola-Villalobos P, Torralbo-Jimenez P, Giron N, de la Heras B, Herrero Vanrell R, et al. The effect of preservative-free HP-Guar on dry eye after phacoemulsification: a flow cytometric study. Eye (Lond) 2010;24:1331–7.

[362] Guyette N, Williams L, Tran MT, Than T, Bradley J, Kehinde L, et al. Comparison of low-abundance biomarker levels in capillary-collected non-stimulated tears and washout tears of aqueous-deficient and normal patients. Invest Ophthalmol Vis Sci 2013;54:3729–37.

[363] Mantopoulos D, Cruzat A, Hamrah P. In vivo imaging of corneal inflammation: new tools for clinical practice and research. Semin Ophthalmol 2010;25:178–85.

[364] Mastropasqua L, Nubile M, Lanzini M, Carpineto P, Ciancaglini M, Pannellini T, et al. Epithelial dendritic cell distribution in normal and inflamed human cornea: in vivo confocal microscopy study. Am J Ophthalmol 2006;142:736–44.

[365] Zhivov A, Stachs O, Kraak R, Stave J, Guthoff RF. In vivo confocal microscopy of the ocular surface. Ocul Surf 2006;4:81–93.

[366] Villani E, Viola F, Sala R, Salvi M, Mapelli C, Curro N, et al. Corneal involvement in Graves' orbitopathy: an in vivo confocal study. Invest Ophthalmol Vis Sci 2010;51:4574–8.

[367] Wakamatsu TH, Okada N, Kojima T, Matsumoto Y, Ibrahim OM, Dogru M, et al. Evaluation of conjunctival inflammatory status by confocal scanning laser microscopy and conjunctival brush cytology in patients with atopic keratoconjunctivitis (AKC). Mol Vis 2009;15:1611–9.

[368] Villani E, Beretta S, De Capitani M, Galimberti D, Viola F, Ratiglia R. In vivo confocal microscopy of meibomian glands in Sjogren's syndrome. Invest Ophthalmol Vis Sci 2011;52:933–9.

[369] Villani E, Galimberti D, Del Papa N, Nucci P, Ratiglia R. Inflammation in dry eye associated with rheumatoid arthritis: cytokine and in vivo confocal microscopy study. Innate Immun 2013;19:420–7.

[370] Kheirkhah A, Rahimi Darabad R, Cruzat A, Hajrasouliha AR, Witkin D, Wong N, et al. Corneal epithelial immune dendritic cell alterations in subtypes of dry eye disease: a pilot in vivo confocal microscopic study. Invest Ophthalmol Vis Sci 2015;56:7179–85.

[371] Villani E, Garoli E, Termine V, Pichi F, Ratiglia R, Nucci P. Corneal confocal microscopy in dry eye treated with corticosteroids. Optom Vis Sci 2015;92.e290–5.

[372] Qazi Y, Kheirkhah A, Blackie C, Cruzat A, Trinidad M, Williams C, et al. In vivo detection of clinically non-apparent ocular surface inflammation in patients with meibomian gland dysfunction-associated refractory dry eye symptoms: a pilot study. Eye (Lond) 2015;29:1099–110.

[373] D'Souza S, Tong L. Practical issues concerning tear protein assays in dry eye. Eye Vis 2014:1.

[374] Tong L, Zhou XY, Jylha A, Aapola U, Liu DN, Koh SK, et al. Quantitation of 47 human tear proteins using high resolution multiple reaction monitoring (HR-MRM) based-mass spectrometry. J Proteomics 2015;115:36–48.

[375] Lam SM, Tong L, Duan X, Petznick A, Wenk MR, Shui G. Extensive characterization of human tear fluid collected using different techniques unravels the presence of novel lipid amphiphiles. J Lipid Res 2014;55:289–98.

[376] Tong L, Petznick A. Correlation between tear matrix metalloproteinases and the Schirmer's test. Invest Ophthalmol Vis Sci 2012;53:1592.

[377] Jones L, Downie LE, Korb DR, Benitez-del-Castillo JM, Dana R, Deng SX, et al. TFOS DEWS II management and therapy report. Ocul Surf 2017;15(3):580–634.

[378] Novack GD, Asbell P, Barabino S, Bergamini MVW, Ciolino JB, Foulks GN, et al. TFOS DEWS II clinical trial design report. Ocul Surf 2017;15(3):635–55.

[379] Korb DR, Greiner JV, Herman JP, Hebert E, Finnemore VM, Exford JM, et al. Lid-wiper epitheliopathy and dry-eye symptoms in contact lens wearers. CLAO J 2002;28:211–6.

[380] Bron AJ, de Paiva CS, Chauhan S, Bonini S, Gabison EE, Jain S, et al. TFOS DEWS II pathophysiology report. Ocul Surf 2017;15(3):441–515.

[381] Knop N, Korb DR, Blackie CA, Knop E. The lid wiper contains goblet cells and goblet cell crypts for ocular surface lubrication during the blink. Cornea 2012;31:668–79.

[382] Navascues-Cornago M, Maldonado-Codina C, Morgan PB. Mechanical sensitivity of the human conjunctiva. Cornea 2014;33:855–9.

[383] Jones L, Varikooty J, Keir N, Soong F, Patel P. The evaluation of lid wiper epitheliopathy in contact lens wearers in a controlled low humidity environmental exposure chamber. ARVO Meet Abstr 2013;54:5475.

[384] Morgan PB, Petropoulos IN, Read ML, Malik RA, Maldonado-Codina C. In vivo confocal microscopy of the lid margin area of contact lens wearers. BCLA Conf 2013. Manchester2013.

[385] Varikooty J, Srinivasan S, Subbaraman L, Woods CA, Fonn D, Simpson TL, et al. Variations in observable lid wiper epitheliopathy (LWE) staining patterns in wearers of silicone hydrogel lenses. Contact Lens Anterior Eye 2015;38:471–6.

[386] Rubio EG. Evaluation of upper eye lid inner marginstaining after using lubricating eye drops. Contact Lens Anterior Eye 2011;34(Supplement 1):S17.

[387] Schmidt TA, Sullivan DA, Knop E, Richards SM, Knop N, Liu S, et al. Transcription, translation, and function of lubricin, a boundary lubricant, at the

ocular surface. JAMA Ophthalmol 2013;131:766—76.

[388] Guillon M, Maissa C. Assessment of contact lens wearers' lid margins with lissamine green. Invest Ophthalmol Vis Sci 2009;50. 6343-.

[389] Shiraishi A, Yamaguchi M, Ohashi Y. Prevalence of upper- and lower-lid-wiper epitheliopathy in contact lens wearers and non-wearers. Eye Contact Lens 2014.

[390] Stahl UG, Delaveris A, Madigan M, Jalbert I. Dallos Award Winner Lid wiper epitheliopathy: exploring the links to comfort and osmolality in contact lens wear. Cont Lens Anterior Eye 2011;34:S18.

[391] Korb DR, Herman JP, Greiner JV, Scaffidi RC, Finnemore VM, Exford JM, et al. Lid wiper epitheliopathy and dry eye symptoms. Eye Contact Lens 2005;31: 2—8.

[392] Rubio EG. Dry eye and upper lid margin staining. Contact Lens Spectr 2012;27:44—7.

[393] Bron AJ, Tiffany JM, Gouveia SM, Yokoi N, Voon LW. Functional aspects of the tear film lipid layer. Exp Eye Res 2004;78:347—60.

[394] Yokoi N, Yamada H, Mizukusa Y, Bron AJ, Tiffany JM, Kato T, et al. Rheology of tear film lipid layer spread in normal and aqueous tear-deficient dry eyes. Invest Ophthalmol Vis Sci 2008;49:5319—24.

[395] Olsen T. Reflectometry of the precorneal film. Acta Ophthalmol (Copenh) 1985;63:432—8.

[396] Goto E, Endo K, Suzuki A, Fujikura Y, Matsumoto Y, Tsubota K. Tear evaporation dynamics in normal subjects and subjects with obstructive meibomian gland dysfunction. Invest Ophthalmol Vis Sci 2003;44:533—9.

[397] Remeseiro B, Penas M, Barreira N, Mosquera A, Novo J, Garcia-Resua C. Automatic classification of the interferential tear film lipid layer using colour texture analysis. Comput Methods Programs Biomed 2013;111:93—103.

[398] Goto E, Tseng SC. Differentiation of lipid tear deficiency dry eye by kinetic analysis of tear interference images. Arch Ophthalmol 2003;121:173—80.

[399] King-Smith PE, Fink BA, Fogt N. Three interferometric methods for measuring the thickness of layers of the tear film. Optom Vis Sci 1999;76: 19—32.

[400] Licznerski TJ, Kasprzak HT, Kowalik W. Analysis of shearing interferograms of tear film using fast fourier transforms. J Biomed Opt 1998;3:32—7.

[401] Szczesna DH, Iskander DR. Robust estimation of tear film surface quality in lateral shearing interferometry. J Biomed Opt 2009;14:064039.

[402] Szczesna DH, Kasprzak HT. Numerical analysis of interferograms for evaluation of tear film build-up time. Ophthalmic Physiol Opt 2009;29:211—8.

[403] Szczesna-Iskander DH, Iskander DR, Read SA, Alonso-Caneiro D. Noninvasive in vivo assessment of soft contact lens type on tear film surface quality. Invest Ophthalmol Vis Sci 2012;53:525—31.

[404] Jester JV, Rife L, Nii D, Luttrull JK, Wilson L, Smith RE. In vivo biomicroscopy and photography of meibomian glands in a rabbit model of meibomian gland dysfunction. Invest Ophthalmol Vis Sci 1982;22:660—7.

[405] Robin JB, Jester JV, Nobe J, Nicolaides N, Smith RE. In vivo transillumination biomicroscopy and photography of meibomian gland dysfunction. A clinical study. Ophthalmology 1985;92:1423—6.

[406] Mathers WD, Shields WJ, Sachdev MS, Petroll WM, Jester JV. Meibomian gland dysfunction in chronic blepharitis. Cornea 1991;10:277—85.

[407] Mathers WD, Daley T, Verdick R. Video imaging of the meibomian gland. Arch Ophthalmol 1994;112:448—9.

[408] Yokoi N, Komuro A, Yamada H, Maruyama K, Kinoshita S. A newly developed video-meibography system featuring a newly designed probe. Jpn J Ophthalmol 2007;51:53—6.

[409] Arita R, Itoh K, Inoue K, Amano S. Noncontact infrared meibography to document age-related changes of the meibomian glands in a normal population. Ophthalmology 2008/05/03:911—5. ed2008.

[410] Arita R, Itoh K, Maeda S, Maeda K, Amano S. A newly developed noninvasive and mobile pen-shaped meibography system. Cornea 2012.

[411] Pult H, Riede-Pult BH. Non-contact meibography: keep it simple but effective. Cont Lens Anterior Eye 2012;35:77—80.

[412] Srinivasan S, Menzies K, Sorbara L, Jones L. Infrared imaging of meibomian gland structure using a novel keratograph. Optom Vis Sci 2012;89:788—94.

[413] Nichols JJ, Berntsen DA, Mitchell GL, Nichols KK. An assessment of grading scales for meibography images. Cornea 2005;24:382—8.

[414] Shimazaki J, Sakata M, Tsubota K. Ocular surface changes and discomfort in patients with meibomian gland dysfunction. Arch Ophthalmol 1995;113: 1266—70.

[415] Mathers WD, Billborough M. Meibomian gland function and giant papillary conjunctivitis. Am J Ophthalmol 1992;114:188—92.

[416] Pflugfelder SC, Tseng SC, Sanabria O, Kell H, Garcia CG, Felix C, et al. Evaluation of subjective assessments and objective diagnostic tests for diagnosing tear-film disorders known to cause ocular irritation. Cornea 1998;17:38—56.

[417] Koh YW, Celik T, Lee HK, Petznick A, Tong L. Detection of meibomian glands and classification of meibography images. J Biomed Opt 2012;17:086008.

[418] Pult H, Riede-Pult BH, Nichols JJ. Relation between upper and lower lids' meibomian gland morphology, tear film, and dry eye. Optom Vis Sci 2012;89:E310—5.

[419] Arita R, Suehiro J, Haraguchi T, Shirakawa R, Tokoro H, Amano S. Objective image analysis of the meibomian gland area. Br J Ophthalmol 2014;98: 746—55.

[420] Ban Y, Shimazaki-Den S, Tsubota K, Shimazaki J. Morphological evaluation of meibomian glands using noncontact infrared meibography. Ocul Surf 2013;11:47—53.

[421] Arita R, Suehiro J, Haraguchi T, Maeda S, Maeda K, Tokoro H, et al. Topical

[422] Arita R, Morishige N, Shirakawa R, Sato Y, Amano S. Effects of eyelid warming devices on tear film parameters in normal subjects and patients with meibomian gland dysfunction. Ocul Surf 2015;13:321—30.

[423] Arita R, Itoh K, Maeda S, Maeda K, Furuta A, Fukuoka S, et al. Proposed diagnostic criteria for obstructive meibomian gland dysfunction. Ophthalmology 2009;116. 2058—63 e1.

[424] Finis D, Ackermann P, Pischel N, Konig C, Hayajneh J, Borrelli M, et al. Evaluation of meibomian gland dysfunction and local distribution of meibomian gland atrophy by non-contact infrared meibography. Curr Eye Res 2015;40:982—9.

[425] Lemp MA, Crews LA, Bron AJ, Foulks GN, Sullivan BD. Distribution of aqueous-deficient and evaporative dry eye in a clinic-based patient cohort: a retrospective study. Cornea 2012;31:472—8.

[426] Eom Y, Lee JS, Kang SY, Kim HM, Song JS. Correlation between quantitative measurements of tear film lipid layer thickness and meibomian gland loss in patients with obstructive meibomian gland dysfunction and normal controls. Am J Ophthalmol 2013;155. 1104—10 e2.

[427] Arita R, Morishige N, Koh S, Shirakawa R, Kawashima M, Sakimoto T, et al. Increased tear fluid production as a compensatory response to meibomian gland loss: a multicenter cross-sectional study. Ophthalmology 2015;122: 925—933.

[428] Pult H, Riede-Pult B. Comparison of subjective grading and objective assessment in meibography. Cont Lens Anterior Eye 2013;36:22—7.

[429] Arita R, Itoh K, Maeda S, Maeda K, Tomidokoro A, Amano S. Efficacy of diagnostic criteria for the differential diagnosis between obstructive meibomian gland dysfunction and aqueous deficiency dry eye. Jpn J Ophthalmol 2010;54:387—91.

[430] Arita R, Itoh K, Inoue K, Kuchiba A, Yamaguchi T, Amano S. Contact lens wear is associated with decrease of meibomian glands. Ophthalmology 2009;116: 379—84.

[431] Bron AJ, Benjamin L, Snibson GR. Meibomian gland disease. Classification and grading of lid changes. Eye (Lond) 1991;5(Pt 4):395—411.

[432] Mathers WD, Shields WJ, Sachdev MS, Petroll WM, Jester JV. Meibomian gland morphology and tear osmolarity: changes with Accutane therapy. Cornea 1991;10:286—90.

[433] Blackie CA, Korb DR. The diurnal secretory characteristics of individual meibomian glands. Cornea 2010;29:34—8.

[434] Shimazaki J, Goto E, Ono M, Shimmura S, Tsubota K. Meibomian gland dysfunction in patients with Sjogren syndrome. Ophthalmology 1998;105: 1485—8.

[435] Henriquez AS, Korb DR. Meibomian glands and contact lens wear. Br J Ophthalmol 1981;65:108—11.

[436] Blackie CA, Korb DR. Recovery time of an optimally secreting meibomian gland. Cornea 2009;28:293—7.

[437] Korb DR, Blackie CA. Meibomian gland diagnostic expressibility: correlation with dry eye symptoms and gland location. Cornea 2008;27:1142—7.

[438] Knop E, Knop N, Zhivov A, Kraak R, Korb DR, Blackie C, et al. The lid wiper and muco-cutaneous junction anatomy of the human eyelid margins: an in vivo confocal and histological study. J Anat 2011;218:449—61.

[439] Knop E, Korb DR, Blackie CA, Knop N. The lid margin is an underestimated structure for preservation of ocular surface health and development of dry eye disease. Dev Ophthalmol 2010;45:108—22.

[440] Randon M, Liang H, El Hamdaoui M, Tahiri R, Batellier L, Denoyer A, et al. In vivo confocal microscopy as a novel and reliable tool for the diagnosis of Demodex eyelid infestation. Br J Ophthalmol 2015;99:336—41.

[441] Kojima T, Ishida R, Sato EA, Kawakita T, Ibrahim OM, Matsumoto Y, et al. In vivo evaluation of ocular demodicosis using laser scanning confocal microscopy. Invest Ophthalmol Vis Sci 2011;52:565—9.

[442] Randon M, Liang H, Abbas R, Michee S, Denoyer A, Baudouin C, et al. A new classification for meibomian gland diseases with in vivo confocal microscopy. J Fr Ophtalmol 2016;39:239—47.

[443] Villani E, Ceresara G, Beretta S, Magnani F, Viola F, Ratiglia R. In vivo confocal microscopy of meibomian glands in contact lens wearers. Invest Ophthalmol Vis Sci 2011;52:5215—9.

[444] Ban Y, Ogawa Y, Ibrahim OM, Tatematsu Y, Kamoi M, Uchino M, et al. Morphologic evaluation of meibomian glands in chronic graft-versus-host disease using in vivo laser confocal microscopy. Mol Vis 2011;17:2533—43.

[445] Ibrahim OM, Matsumoto Y, Dogru M, Adan ES, Wakamatsu TH, Shimazaki J, et al. In vivo confocal microscopy evaluation of meibomian gland dysfunction in atopic-keratoconjunctivitis patients. Ophthalmology 2012;119: 1961—8.

[446] Agnifili L, Fasanella V, Costagliola C, Ciabattoni C, Mastropasqua R, Frezzotti P, et al. In vivo confocal microscopy of meibomian glands in glaucoma. Br J Ophthalmol 2013;97:343—9.

[447] Villani E, Canton V, Magnani F, Viola F, Nucci P, Ratiglia R. The aging Meibomian gland: an in vivo confocal study. Invest Ophthalmol Vis Sci 2013;54: 4735—40.

[448] Matsumoto Y, Shigeno Y, Sato EA, Ibrahim OM, Saiki M, Negishi K, et al. The evaluation of the treatment response in obstructive meibomian gland disease by in vivo laser confocal microscopy. Graefes Arch Clin Exp Ophthalmol 2009;247:821—9.

[449] Villani E, Garoli E, Canton V, Pichi F, Nucci P, Ratiglia R. Evaluation of a novel eyelid-warming device in meibomian gland dysfunction unresponsive to

traditional warm compress treatment: an in vivo confocal study. Int Ophthalmol 2015;35:319–23.

[450] Linton RG, Curnow DH, Riley WJ. The meibomian glands: an investigation into the secretion and some aspects of the physiology. Br J Ophthalmol 1961;45:718–23.

[451] Knop N, Knop E. Meibomian glands. Part I: anatomy, embryology and histology of the Meibomian glands. Ophthalmologe 2009;106:872–83.

[452] McMonnies CW. Incomplete blinking: exposure keratopathy, lid wiper epitheliopathy, dry eye, refractive surgery, and dry contact lenses. Cont Lens Anterior Eye 2007;30:37–51.

[453] Cruz AA, Garcia DM, Pinto CT, Cechetti SP. Spontaneous eyeblink activity. Ocul Surf 2011;9:29–41.

[454] Doane MG. Blinking and the mechanics of the lacrimal drainage system. Ophthalmology 1981;88:844–51.

[455] Carney LG, Hill RM. The nature of normal blinking patterns. Acta Ophthalmol 1982;60:427–33.

[456] Forst G. Structure of the tear film during the blinking process. Ophthalmic Physiol Opt 1987;7:81–3.

[457] Nakamori K, Odawara M, Nakajima T, Mizutani T, Tsubota K. Blinking is controlled primarily by ocular surface conditions. Am J Ophthalmol 1997;124:24–30.

[458] Rambold H, Sprenger A, Helmchen C. Effects of voluntary blinks on saccades, vergence eye movements, and saccade-vergence interactions in humans. J Neurophysiol 2002;88:1220–33.

[459] Varikooty J, Srinivasan S, Chan A, Subbaraman L, Woods CA, Simpson T, et al. Clinical manifestations of upper lid staining in adapted silicone hydrogel lens wearers. http://siliconehydrogels.org/pdf/posters/jan_07/jan_07.pdf (accessed May 2017).

[460] Knop E, Knop N, Schirra F. Meibomian glands. Part II: physiology, characteristics, distribution and function of meibomian oil. Ophthalmologe 2009;106:884–92.

[461] Craig JP, Wang MT, Kim D, Lee JM. Exploring the predisposition of the asian eye to development of dry eye. Ocul Surf 2016;14:385–92.

[462] Abelson MB, Holly FJ. A tentative mechanism for inferior punctate keratopathy. Am J Ophthalmol 1977;83:866–9.

[463] Doane MG. Interactions of eyelids and tears in corneal wetting and the dynamics of the normal human eyeblink. Am J Ophthalmol 1980;89:507–16.

[464] Collins MJ, Iskander DR, Saunders A, Hook S, Anthony E, Gillon R. Blinking patterns and corneal staining. Eye Contact Lens 2006;32:287–93.

[465] Doughty MJ. Consideration of three types of spontaneous eyeblink activity in normal humans: during reading and video display terminal use, in primary gaze, and while in conversation. Optom Vis Sci 2001;78:712–25.

[466] King DC, Michels KM. Muscular tension and the human blink rate. J Exp Psychol 1957;53:113–6.

[467] Zaman ML, Doughty MJ, Button NF. The exposed ocular surface and its relationship to spontaneous eyeblink rate in elderly caucasians. Exp Eye Res 1998;67:681–6.

[468] Sun WS, Baker RS, Chuke JC, Rouholiman BR, Hasan SA, Gaza W, et al. Age-related changes in human blinks. Passive and active changes in eyelid kinematics. Invest Ophthalmol Vis Sci 1997;38:92–9.

[469] Blackie CA, Korb DR. A novel lid seal evaluation: the Korb-Blackie light test. Eye Contact Lens 2015;41:98–100.

[470] Tsubota K, Hata S, Okusawa Y, Egami F, Ohtsuki T, Nakamori K. Quantitative videographic analysis of blinking in normal subjects and patients with dry eye. Arch Ophthalmol 1996;114:715–20.

[471] Karson CN, Burns RS, LeWitt PA, Foster NLR.P.Newman. Blink rates and disorders of movement. Neurology 1984;34:677–8.

[472] Tsubota K, Nakamori K. Dry eyes and video display terminals. N Engl J Med 1993;328:584.

[473] Pult H, Murphy P, Riede-Pult BH. Velocity of upper lid in spontaneous complete blinks and dry eye. BCLA Conf 2014. Birmingham.

[474] Pult H. What actually happens to your contact lens when you blink. BCLA Conf 2015. Liverpool.

[475] Pult H, Korb DR, Murphy PJ, Riede-Pult BH, Blackie C. A new model of central lid margin apposition and tear film mixing in spontaneous blinking. Contact Lens Anterior Eye 2015;38:173–80.

[476] Norn MS. Conjunctival sensitivity in normal eyes. Acta Ophthalmol (Copenh) 1973;51:58–66.

[477] Norn MS. Conjunctival sensitivity in pathological cases, with simultaneous measurement of corneal and lid margin sensitivity. Acta Ophthalmol (Copenh) 1975;53:450–7.

[478] McGowan DP, Lawrenson JG, Ruskell GL. Touch sensitivity of the eyelid margin and palpebral conjunctiva. Acta Ophthalmol (Copenh) 1994;72:57–60.

[479] Golebiowski B, Chim K, So J, Jalbert I. Lid margins: sensitivity, staining, meibomian gland dysfunction, and symptoms. Optom Vis Sci 2012;89:1443–9.

[480] Tomlinson A, Bron AJ, Korb DR, Amano S, Paugh JR, Pearce EI, et al. The international workshop on meibomian gland dysfunction: report of the diagnosis subcommittee. Invest Ophthalmol Vis Sci 2011;52:2006–49.

[481] Woods CADK, Jones L, Fonn D. Patient use of smartphones to communicate subjective data in clinical trials. Optom Vis Sci 2011;88:290–4.

[482] Lienert JP, Tarko L, Uchino M, Christen WG, Schaumberg DA. Long-term Natural History of Dry Eye Disease from the Patient's Perspective. Ophthalmology 2016;123:425–33.

[483] Geerling G, Tauber J, Baudouin C, E.Goto, Matsumoto Y, O'Brien T, et al. The international workshop on meibomian gland dysfunction: report of the subcommittee on management and treatment of meibomian gland dysfunction. Invest Ophthalmol Vis Sci 2011;52:2050–64.

[484] Efron N, Brennan NA, Morgan PB, Wilson T. Lid wiper epitheliopathy. Prog Retin Eye Res 2016;53:140–74.

[485] Walker PM, Lane KJ, Ousler 3rd GW, Abelson MB. Diurnal variation of visual function and the signs and symptoms of dry eye. Cornea 2010;29:607–12.

[486] Doughty MJ, Fonn D, Richter D, Simpson T, Caffery B, Gordon K. A patient questionnaire approach to estimating the prevalence of dry eye symptoms in patients presenting to optometric practices across Canada. Optom Vis Sci 1997;74:624–31.

[487] Voulgarelis M, Skopouli FN. Clinical, immunologic, and molecular factors predicting lymphoma development in Sjogren's syndrome patients. Clin Rev Allergy Immunol 2007;32:265–74.

[488] Bonini S. Atopic keratoconjunctivitis. Allergy 2004;59(Suppl 78):71–3.

[489] Kubicka-Trzaska A, Romanowska-Dixon B. Dry eye syndrome and allergic conjunctivitis–epidemics of XXI century–diagnostic problems and management. Przeglad Lek 2009;66:967–71.

[490] Akil H, Celik F, Ulas F, Kara IS. Dry Eye Syndrome and Allergic Conjunctivitis in the Pediatric Population. Middle East Afr J Ophthalmol 2015;22:467–71.

[491] Bousquet J, Schunemann HJ, Fonseca J, Samolinski B, Bachert C, Canonica GW, et al. MACVIA-ARIA Sentinel NetworK for allergic rhinitis (MASK-rhinitis): the new generation guideline implementation. Allergy 2015;70:1372–92.

[492] Seidman MD, Gurgel RK, Lin SY, Schwartz SR, Baroody FM, Bonner JR, et al. Clinical practice guideline: Allergic rhinitis. Otolaryngol Head Neck Surg 2015;152:S1–43.

[493] Bonini M, Gramiccioni C, Fioretti D, Ruckert B, Rinaldi M, Akdis C, et al. Asthma, allergy and the Olympics: a 12-year survey in elite athletes. Curr Opin Allergy Clin Immunol 2015;15:184–92.

[494] Welch D, Ousler 3rd GW, Nally LA, Abelson MB, Wilcox KA. Ocular drying associated with oral antihistamines (loratadine) in the normal population-an evaluation of exaggerated dose effect. Adv Exp Med Biol 2002;506:1051–5.

[495] Gomes JAP, Azar DT, Baudouin C, Efron N, Hirayama M, Horwath-Winter J, et al. TFOS DEWS II iatrogenic report. Ocul Surf 2017;15(3):516–43.

[496] Bielory L. Ocular toxicity of systemic asthma and allergy treatments. Curr Allergy Asthma Rep 2006;6:299–305.

[497] Friedman T, Friedman Z, Neumann E. Giant papillary conjunctivitis following cataract extraction. Ann Ophthalmol 1984;16:50–2.

[498] Chen JJ, Applebaum DS, Sun GS, Pflugfelder SC. Atopic keratoconjunctivitis: A review. J Am Acad Dermatol 2014;70:569–75.

[499] Cornish KS, Gregory ME, Ramaesh K. Systemic cyclosporin A in severe atopic keratoconjunctivitis. Eur J Ophthalmol 2010;20:844–51.

[500] Akpek EK, Dart JK, Watson S, Christen W, Dursun D, Yoo S, et al. A randomized trial of topical cyclosporin 0.05% in topical steroid-resistant atopic keratoconjunctivitis. Ophthalmology 2004;111:476–82.

[501] Akova YA, Rodriguez A, Foster CS. Atopic keratoconjunctivitis. Ocular Immunol Inflamm 1994;2:125–44.

[502] Onguchi T, Dogru M, Okada N, Kato NA, Tanaka M, Takano Y, et al. The impact of the onset time of atopic keratoconjunctivitis on the tear function and ocular surface findings. Am J Ophthalmol 2006;141:569–71.

[503] Dogru M, Katakami C, Nakagawa N, Tetsumoto K, Yamamoto M. Impression cytology in atopic dermatitis. Ophthalmology 1998;105:1478–84.

[504] Power WJ, Tugal-Tutkun I, Foster CS. Long-term follow-up of patients with atopic keratoconjunctivitis. Ophthalmology 1998;105:637–42.

[505] Guglielmetti S, Dart JK, Calder V. Atopic keratoconjunctivitis and atopic dermatitis. Curr Opin Allergy Clin Immunol 2010;10:478–85.

[506] Bielory B, Bielory L. Atopic dermatitis and keratoconjunctivitis. Immunol Allergy Clin N Am 2010;30:323–36.

[507] Villani E, Strologo MD, Pichi F, Luccarelli SV, De Cilla S, Serafino M, et al. Dry Eye in Vernal Keratoconjunctivitis: A Cross-Sectional Comparative Study. Medicine 2015;94:e1648.

[508] Vichyanond P, Pacharn P, Pleyer U, Leonardi A. Vernal keratoconjunctivitis: a severe allergic eye disease with remodeling changes. Pediatr Allergy Immunol 2014;25:314–22.

[509] Bonini S, Coassin M, Aronni S, Lambiase A. Vernal keratoconjunctivitis. Eye (Lond) 2004;18:345–51.

[510] Lee SW, Lee SC, Jin KH. Conjunctival Inclusion Cysts in Long-standing Chronic Vernal Keratoconjunctivitis. Korean J Ophthalmol 2007;21:251–4.

[511] Leonardi A, Piliego F, Castegnaro A, Lazzarini D, La Gloria Valerio A, Mattana P, et al. Allergic conjunctivitis: a cross-sectional study. Clin Exp Immunol 2015;45:1118–25.

[512] Maranhao AG, Soares CC, Albuquerque MC, Santos N. Molecular epidemiology of adenovirus conjunctivitis in Rio de Janeiro, Brazil, between 2004 and 2007. Rev Inst Med Trop Sao Paulo 2009;51:227–9.

[513] Lynch 3rd JP, Fishbein M, Echavarria M. Adenovirus. Seminars Respir Crit Care Med 2011;32:494–511.

[514] Alfonso SA, Fawley JD, Alexa Lu X. Conjunctivitis. Prim Care 2015;42:325–45.

[515] Sheppard JD, Wertheimer MLScoper SV. Modalities to decrease stromal herpes simplex keratitis reactivation rates. Arch Ophthalmol 2009;127:852–6.

[516] Yawn BP, Wollan PC, St Sauver JL, Butterfield LC. Herpes zoster eye complications: rates and trends. Mayo Clin Proc 2013;88:562–70.

[517] Henle G, Henle W, Clifford P, Diehl V, Kafuko GW, Kirya BG, et al. Antibodies to Epstein-Barr virus in Burkitt's lymphoma and control groups. J Natl Cancer Inst 1969;43:1147–57.

[518] Wong KW, D'Amico DJ, Hedges 3rd TR, Soong HK, Schooley RT, Kenyon KR. Ocular involvement associated with chronic Epstein-Barr virus disease. Arch Ophthalmol 1987;105:788–92.

[519] Matoba AY. Ocular disease associated with Epstein-Barr virus infection. Surv Ophthalmol 1990;35:145–50.

[520] Sambursky R, Trattler W, Tauber S, Starr C, Friedberg M, Boland T, et al. Sensitivity and specificity of the AdenoPlus test for diagnosing adenoviral conjunctivitis. JAMA Ophthalmol 2013;131:17–22.

[521] Orden Martinez B, Martinez Ruiz R, Millan Perez R. Bacterial conjunctivitis: most prevalent pathogens and their antibiotic sensitivity. An Pediatr (Barc) 2004;61:32–6.

[522] Sugita G, Hotomi M, Sugita R, Kono M, Togawa A, Yamauchi K, et al. Genetic characteristics of Haemophilus influenzae and Streptococcus pneumoniae isolated from children with conjunctivitis-otitis media syndrome. J Infect Chemother 2014;20:493–7.

[523] Pflugfelder SC, Karpecki PM, Perez VL. Treatment of blepharitis: recent clinical trials. Ocul Surf 2014;12:273–84.

[524] Panel ACEDP. Blepharitis PPP. Am Acad Ophthalmol 2013.

[525] McCulley JP, Dougherty JM, Deneau DG. Classification of chronic blepharitis. Ophthalmology 1982;89:1173–80.

[526] Baum J. Clinical manifestations of dry eye states. Trans Ophthalmol Soc U. K 1985;104(Pt 4):415–23.

[527] Post CF, Juhlin E. Demodex Folliculorum and Blepharitis. Arch Dermatol 1963;88:298–302.

[528] Hom MM, Mastrota KM, Schachter SE. Demodex. Optom Vis Sci 2013;90: e198–205.

[529] Liu J, Sheha H, Tseng SC. Pathogenic role of Demodex mites in blepharitis. Curr Opin Allergy Clin Immunol 2010;10:505–10.

[530] Kamoun B, Fourati M, Feki J, Mlik M, Karray F, Trigui A, et al. Blepharitis due to Demodex: myth or reality? J Fr Ophtalmol 1999;22:525–7.

[531] Gutgesell VJ, Stern GA, Hood CI. Histopathology of meibomian gland dysfunction. Am J Ophthalmol 1982;94:383–7.

[532] Arrua M, Samudio M, Farina N, Cibils D, Laspina F, Sanabria R, et al. Comparative study of the efficacy of different treatment options in patients with chronic blepharitis. Arch Soc Esp Oftalmol 2015;90:112–8.

[533] Thode AR, Latkany RA. Current and Emerging Therapeutic Strategies for the Treatment of Meibomian Gland Dysfunction (MGD). Drugs 2015;75: 1177–85.

[534] Jalbert I, Rejab S. Increased numbers of Demodex in contact lens wearers. Optom Vis Sci 2015;92:671–8.

[535] Norn MS. Incidence of Demodex Folliculorum on skin of lids and nose. Acta Ophthalmol 1982;60:575–83.

[536] Liang LY, Ding XH, Tseng SCG. High Prevalence of Demodex brevis Infestation in Chalazia. Am J Ophthalmol 2014;157:342–8.

[537] Wolf R, Ophir J, Avigad J, Lengy J, Krakowski A. The hair follicle mites (Demodex spp.). Could they be vectors of pathogenic microorganisms? Acta Derm Venereol 1988;68:535–7.

[538] Bevins CL, Liu FT. Rosacea: skin innate immunity gone awry? Nat Med 2007;13:904–6.

[539] Gao YY, Di Pascuale MA, Elizondo A, Tseng SC. Clinical treatment of ocular demodecosis by lid scrub with tea tree oil. Cornea 2007;26:136–43.

[540] Kheirkhah A, Casas V, Li W, Raju VK, Tseng SC. Corneal manifestations of ocular demodex infestation. Am J Ophthalmol 2007;143:743–9.

[541] Murube J. Demodex hominis. Ocul Surf 2015.

[542] Mastrota KM. Method to identify Demodex in the eyelash follicle without epilation. Optom Vis Sci 2013;90. e172–4.

[543] Krasny J, Hruba D, Netukova M. The role of Chlamydia pneumoniae in the etiology of keratoconjunctivitis sicca (KCS). Recent Pat Inflamm Allergy Drug Discov 2014;8:216–22.

[544] Blodi BA, Byrne KA, Tabbara KF. Goblet cell population among patients with inactive trachoma. Int Ophthalmol 1988;12:41–5.

[545] Kim BY, Riaz KM, Bakhtiari P, Chan CC, Welder JD, Holland EJ, et al. Medically reversible limbal stem cell disease: clinical features and management strategies. Ophthalmology 2014;121:2053–8.

[546] Pisella PJ, Pouliquen P, Baudouin C. Prevalence of ocular symptoms and signs with preserved and preservative free glaucoma medication. Br J Ophthalmol 2002;86:418–23.

[547] Dua HS, Watson NJ, Mathur RM, Forrester JV. Corneal epithelial cell migration in humans: 'hurricane and blizzard keratopathy'. Eye (Lond) 1993;7(Pt 1):53–8.

[548] Nelson JD. Superior limbic keratoconjunctivitis (SLK). Eye (Lond) 1989;3(Pt 2):180–9.

[549] Culbertson WW, Tseng SC. Corneal disorders in floppy eyelid syndrome. Cornea 1994;13:33–42.

[550] Di Pascuale MA, Espana EM, Kawakita T, Tseng SC. Clinical characteristics of conjunctivochslasis with or without aqueous teas deficiency. Br J Ophthalmol 2004;88:388–92.

[551] Murube J. Characteristics and etiology of conjunctivochalasis: historical perspective. Ocul Surf 2005;3:7–14.

[552] Saw VP, Dart JK. Ocular mucous membrane pemphigoid: diagnosis and management strategies. Ocul Surf 2008;6:128–42.

[553] Sobolewska B, Deuter C, Zierhut M. Current medical treatment of ocular mucous membrane pemphigoid. Ocul Surf 2013;11:259–66.

[554] Kohanim S, Palioura S, Saeed HN, Akpek EK, Amescua G, Basu S, et al. Stevens-Johnson Syndrome/Toxic Epidermal Necrolysis–A Comprehensive Review and Guide to Therapy. I. Systemic Disease. Ocul Surf 2016;14:2–19.

[555] Zaidman GW, Geeraets R, Paylor RR, Ferry AP. The histopathology of filamentary keratitis. Arch Ophthalmol 1985;103:1178–81.

[556] Albietz J, Sanfilippo P, Troutbeck R, Lenton LM. Management of filamentary keratitis associated with aqueous-deficient dry eye. Optom Vis Sci 2003;80: 420–30.

[557] Schwartz GS, Harrison AR, Holland EJ. Etiology of immune stromal (interstitial) keratitis. Cornea 1998;17:278–81.

[558] Heigle TJ, Pflugfelder SC. Aqueous tear production in patients with neurotrophic keratitis. Cornea 1996;15:135–8.

[559] Goins KM. New insights into the diagnosis and treatment of neurotrophic keratopathy. Ocul Surf 2005;3:96–110.

[560] Siu GD, Young AL, Jhanji V. Alternatives to corneal transplantation for the management of bullous keratopathy. Curr Opin Ophthalmol 2014;25: 347–52.

[561] Henrich CF, Ramulu PY, Akpek EK. Association of dry eye and inflammatory systemic diseases in a tertiary care-based sample. Cornea 2014;33:819–25.

[562] Kruszka P, O'Brian RJ. Diagnosis and management of Sjogren syndrome. Am Fam Physician 2009;79:465–70.

[563] Baldini C, Pepe P, Luciano N, Ferro F, Talarico R, Grossi S, et al. A clinical prediction rule for lymphoma development in primary Sjogren's syndrome. J Rheumatol 2012;39:804–8.

[564] Shiboski SC, Shiboski CH, Criswell L, Baer A, Challacombe S, Lanfranchi H, et al. American College of Rheumatology classification criteria for Sjogren's syndrome: a data-driven, expert consensus approach in the Sjogren's International Collaborative Clinical Alliance cohort. Arthritis Care Res (Hoboken) 2012;64:475–87.

[565] Matossian C, Micucci J. Characterization of the serological biomarkers associated with Sjogren's syndrome in patients with recalcitrant dry eye disease. Clin Ophthalmol 2016;10:1329–34.

[566] Aakre BM, Doughty MJ. Are there differences between 'visual symptoms' and specific ocular symptoms associated with video display terminal (VDT) use? Cont Lens Anterior Eye 2007;30:174–82.

[567] Blehm C, Vishnu S, Khattak A, Mitra S, Yee RW. Computer vision syndrome: a review. Surv Ophthalmol 2005;50:253–62.

[568] Chu C, Rosenfield M, Portello JK, Benzoni JA, Collier JD. A comparison of symptoms after viewing text on a computer screen and hardcopy. Ophthalmic Physiol Opt 2011;31:29–32.

[569] Chu CA, Rosenfield M, Portello JK. Blink patterns: reading from a computer screen versus hard copy. Optom Vis Sci 2014;91:297–302.

[570] Balasubramaniam SC, Raja H, Nau CB, Shen JF, Schornack MM. Ocular Graft-Versus-Host Disease: A Review. Eye Contact Lens 2015;41:256–61.

[571] Ferrara JL, Levine JE, Reddy P, Holler E. Graft-versus-host disease. Lancet (Lond) 2009;373:1550–61.

[572] Hessen M, Akpek EK. Ocular graft-versus-host disease. Curr Opin Allergy Clin Immunol 2012;12:540–7.

[573] Lin X, Cavanagh HD. Ocular manifestations of graft-versus-host disease: 10 years' experience. Clin Ophthalmol 2015;9:1209–13.

[574] Papas EB, Ciolino JB, Jacobs D, Miller WL, Pult H, Sahin A, et al. The TFOS International Workshop on Contact Lens Discomfort: report of the management and therapy subcommittee. Invest Ophthalmol Vis Sci 2013;54. Tfos183–203.

[575] Ayaki M, Kawashima M, Negishi K, Tsubota K. High prevalence of sleep and mood disorders in dry eye patients: survey of 1,000 eye clinic visitors. Neuropsychiatr Dis Treat 2015;11:889–94.

[576] Szakats I, Sebestyen M, Nemeth J, Birkas E, Purebl G. The Role of Health Anxiety and Depressive Symptoms in Dry Eye Disease. Curr Eye Res 2015: 1–6.

[577] Labbe A, Wang YX, Jie Y, Baudouin C, Jonas JB, Xu L. Dry eye disease, dry eye symptoms and depression: the Beijing Eye Study. Br J Ophthalmol 2013;97: 1399–403.

[578] Hallak JA, Tibrewal S, Jain S. Depressive Symptoms in Patients With Dry Eye Disease: A Case-Control Study Using the Beck Depression Inventory. Cornea 2015;34:1545–50.

[579] Na KS, Han K, Park YG, Na C, Joo CK. Depression, Stress, Quality of Life, and Dry Eye Disease in Korean Women: A Population-Based Study. Cornea 2015;34:733–8.

[580] van der Vaart R, Weaver MA, Lefebvre C, Davis RM. The association between dry eye disease and depression and anxiety in a large population-based study. Am J Ophthalmol 2015;159:470–4.

[581] Galor A, Covington D, Levitt AE, McManus KT, Seiden B, Felix ER, et al. Neuropathic Ocular Pain due to Dry Eye Is Associated With Multiple Comorbid Chronic Pain Syndromes. J Pain 2016;17:310–8.

[582] Galor A, Feuer W, Lee DJ, Florez H, Faler AL, Zann KL, et al. Depression, post-traumatic stress disorder, and dry eye syndrome: a study utilizing the national United States Veterans Affairs administrative database. Am J Ophthalmol 2012;154. 340–6 e2.

[583] Vickers ER, Cousins MJ. Neuropathic orofacial pain. Part 2-Diagnostic procedures, treatment guidelines and case reports. Aust Endod J 2000;26: 53–63.

[584] Morreale M, Marchione P, Giacomini P, Pontecorvo S, Marianetti M, Vento C, et al. Neurological involvement in primary Sjogren syndrome: a focus on

central nervous system. PLoS One 2014;9:e84605.

[585] Epstein LC, Masse G, Harmatz JS, Scott TM, Papas AS, Greenblatt DJ. Characterization of cognitive dysfunction in Sjogren's syndrome patients. Clin Rheumatol 2014;33:511–21.

[586] Strömbeck B, Ekdahl C, Manthorpe R, Wikström I, Jacobsson L. Health-related quality of life in primary Sjögren's syndrome, rheumatoid arthritis and fibromyalgia compared to normal population data using SF-36. Scand J Rheumatol 2000;29:20–8.

[587] von Thun Und Hohenstein-Blaul N, Funke S, Grus FH. Tears as a source of biomarkers for ocular and systemic diseases. Exp Eye Res 2013;117:126–37.

[588] Willcox MD, Zhao Z, Naduvilath T, Lazon de la Jara P. Cytokine changes in tears and relationship to contact lens discomfort. Mol Vis 2015;21:293–305.

[589] Laguna M, Holgado M, Hernandez AL, Santamaria B, Lavin A, Soria J, et al. Antigen-Antibody Affinity for Dry Eye Biomarkers by Label Free Biosensing.

Comparison with the ELISA Technique. Sensors (Basel) 2015;15:19819–29.

[590] Macri A, Scanarotti C, Bassi AM, Giuffrida S, Sangalli G, Traverso CE, et al. Evaluation of oxidative stress levels in the conjunctival epithelium of patients with or without dry eye, and dry eye patients treated with preservative-free hyaluronic acid 0.15 % and vitamin B12 eye drops. Graefes Arch Clin Exp Ophthalmol 2015;253:425–30.

[591] Braun RJ, Gewecke NR, Begley CG, King-Smith PE, Siddique JI. A model for tear film thinning with osmolarity and fluorescein. Invest Ophthalmol Vis Sci 2014;55:1133–42.

[592] Varikooty J, Keir N, Simpson T. Estimating tear film spread and stability through tear hydrodynamics. Optom Vis Sci 2012;89:E1119–24.

[593] Shaheen BS, Bakir M, Jain S. Corneal nerves in health and disease. Surv Ophthalmol 2014;59:263–85.